

10ths Scientific ESSOMM Winter Conference

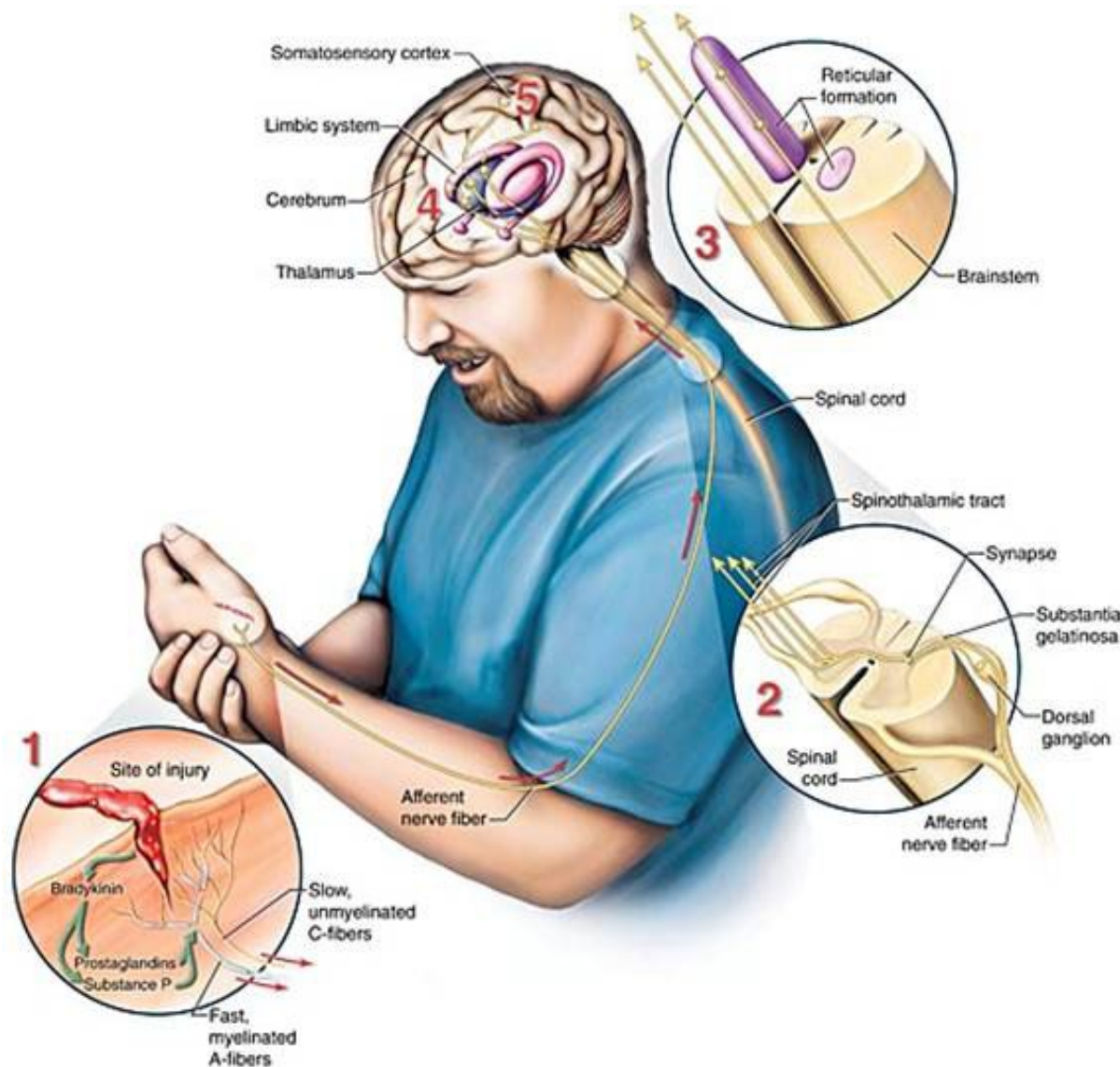
Lech/Austria 11-14 December 2023

Endogenous pain inhibitory mechanisms

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Definition of pain and nociception



„Pain is an unpleasant sensory and emotional experience associated with actual or potential tissue damage, or described in terms of such damage.“

International Association for the Study of Pain, 1979

Nociception: all peripheral and central neuronal processes that can lead to pain

Pain: sensation, perception, cognition, experience; pain requires consciousness

Adressed topics

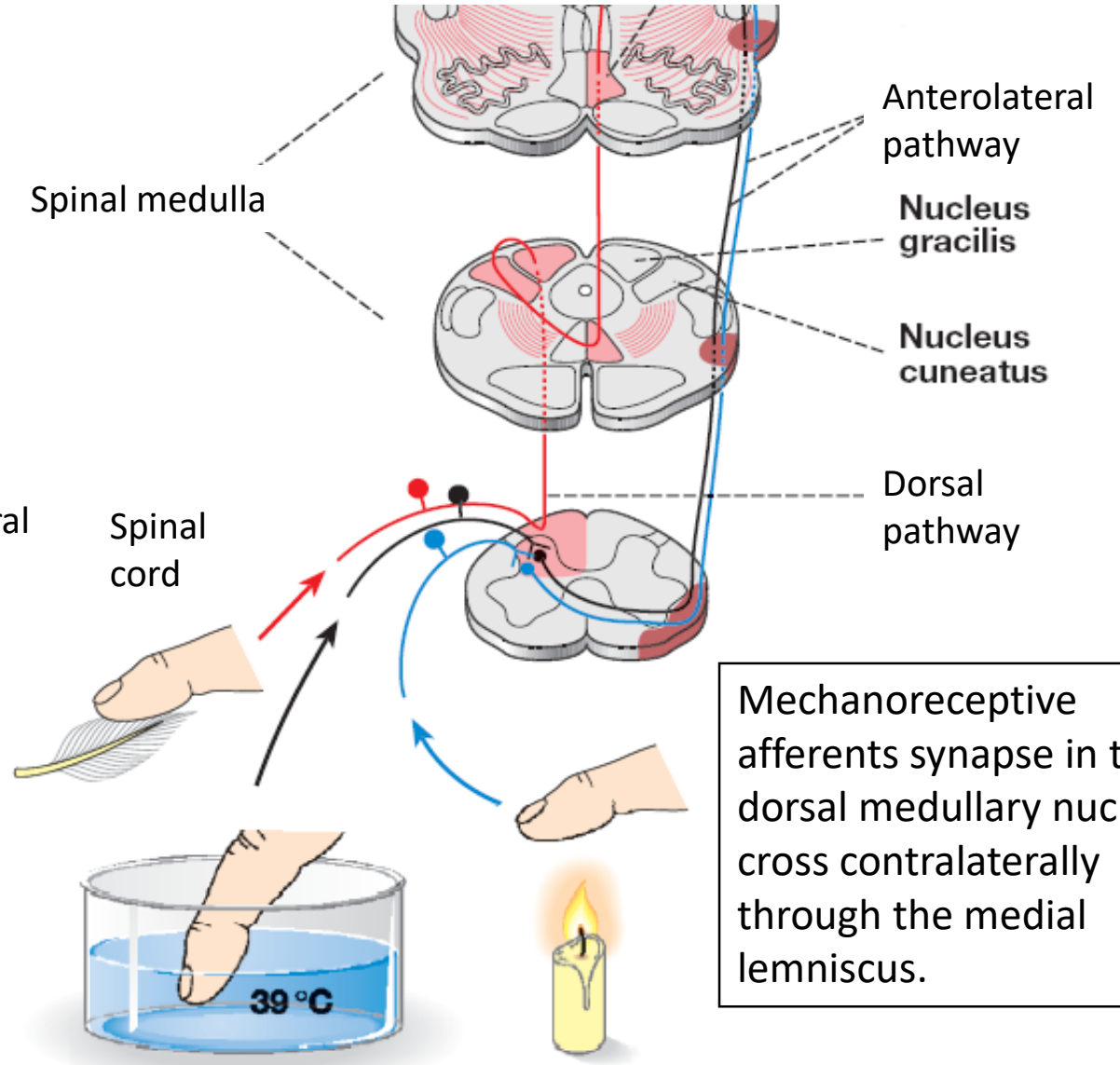
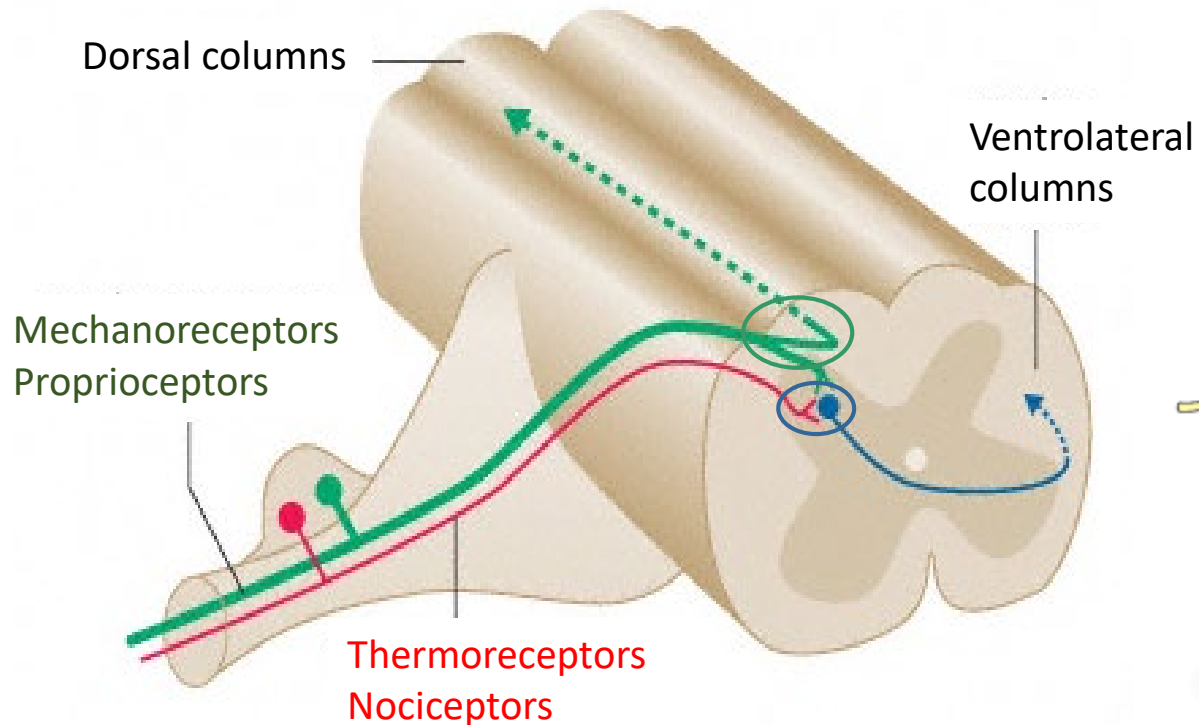
- **Retrospection:** Basic organization of ascending and descending pain modulating pathways
- **Recollection:** Molecular mechanisms of nociceptive transmission and central sensitization
- **Controlling spinal nociception:** Interneurons and molecular antinociceptive mechanisms
- **Descending pathways:** On and Off cells in the rostral ventromedial medulla
- **Supraspinal antinociceptive mechanisms:** Diffuse inhibitory control systems (DNIC)
- **Central pain inhibition:** Stress- and exercise-induced hypoalgesia and acupuncture

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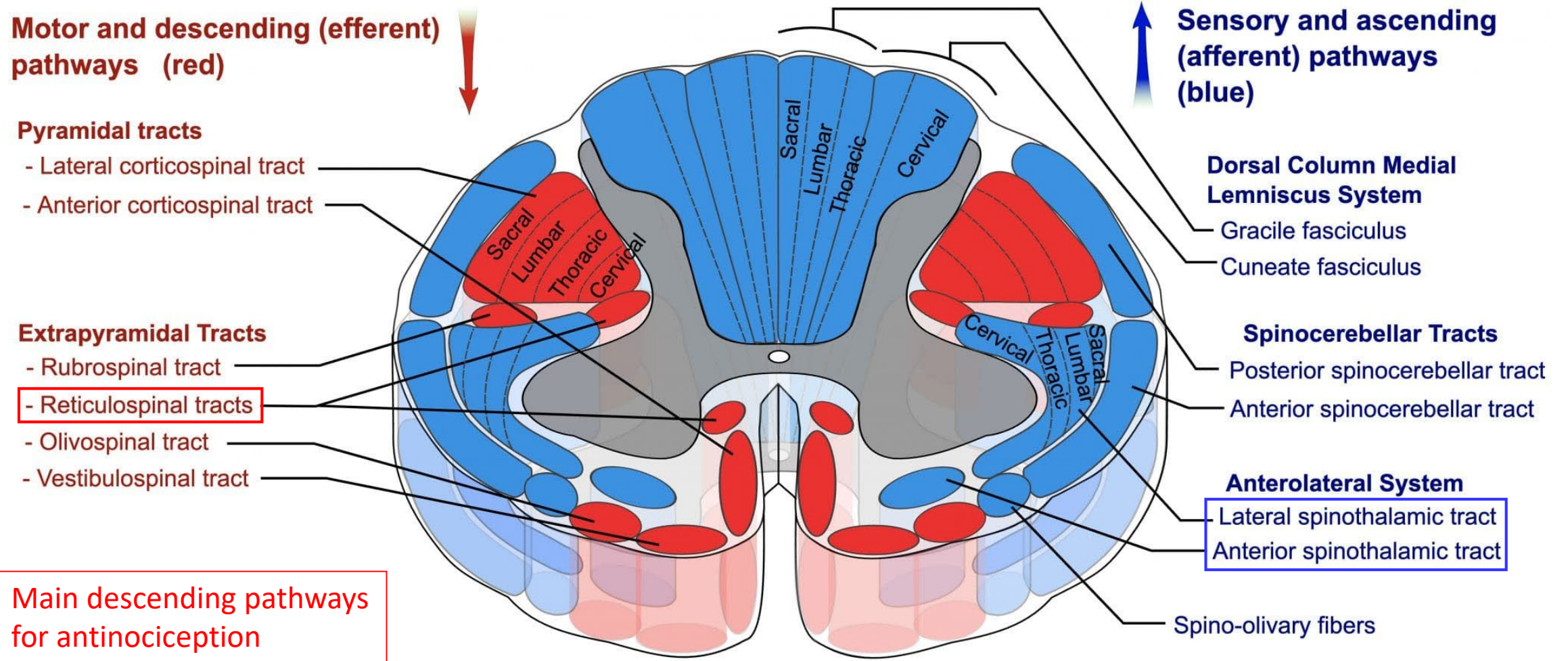
Ascending sensory spinal pathways

Mechanoreceptive afferents ($A\beta$ fibers) send collaterals into the dorsal horn; the main pathways ascent in the ipsilateral dorsal columns. Nociceptive and thermoreceptive afferents ($A\delta$ and C fibers) synapse in the dorsal horn on second order neurons, which ascend in the antero-lateral columns.



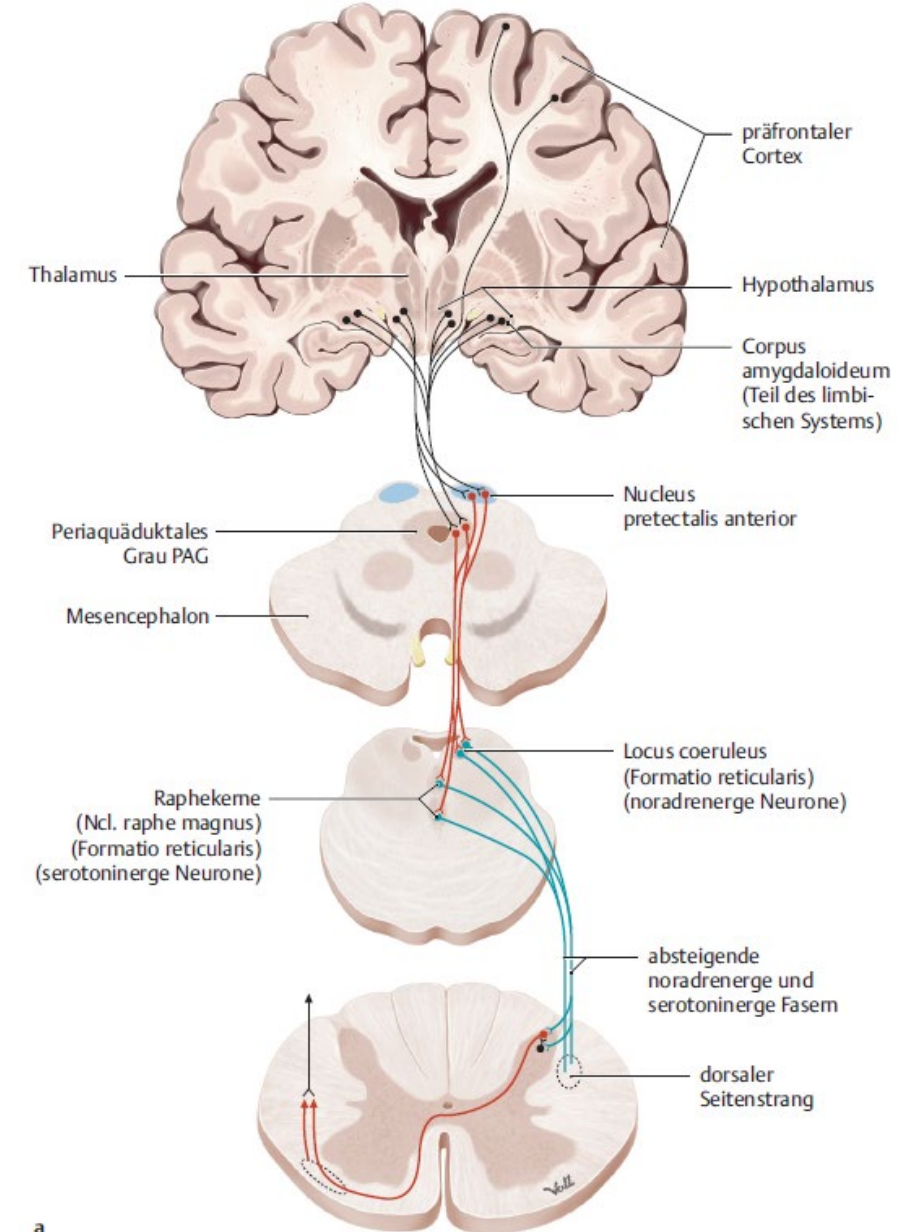
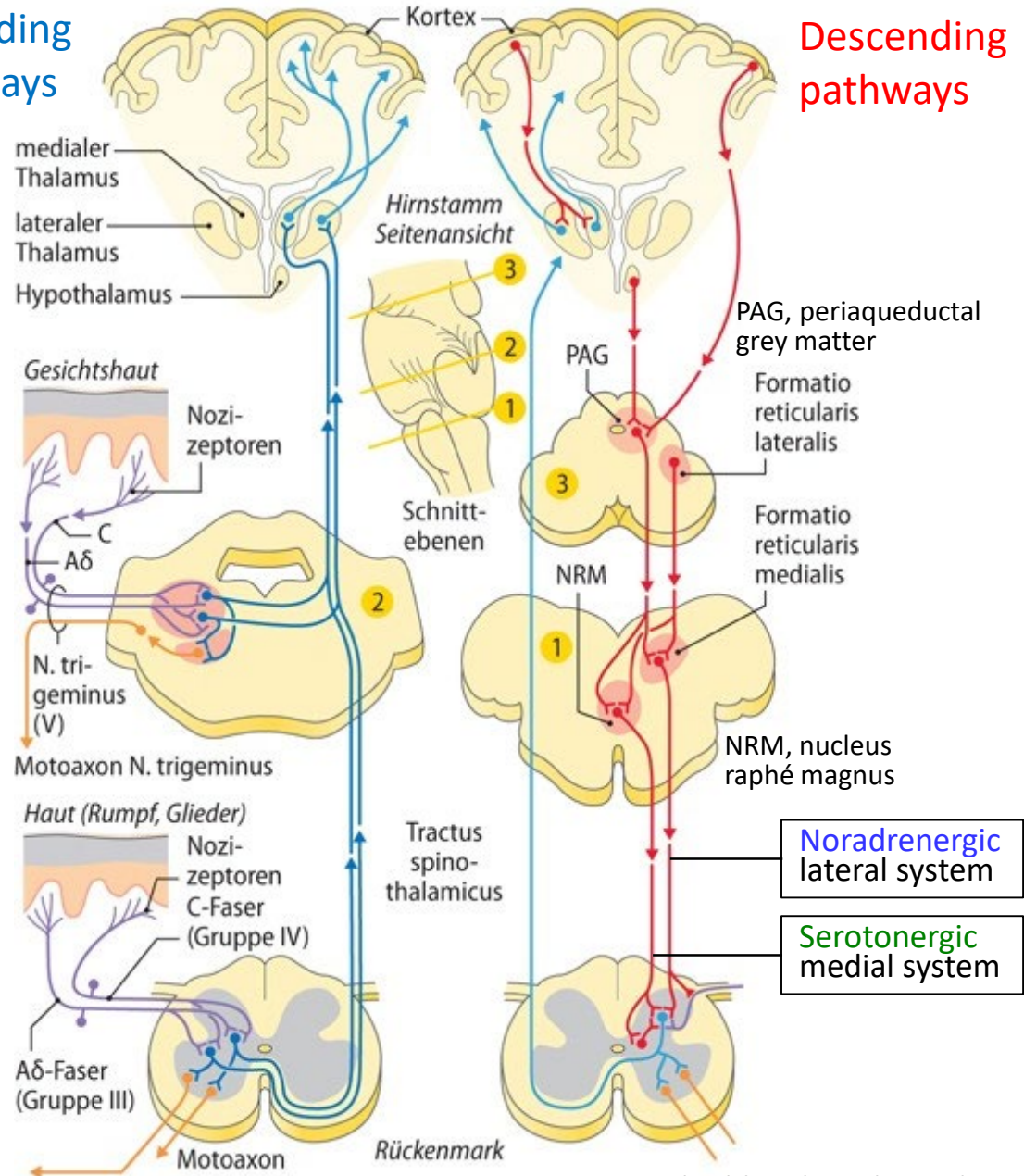
Mechanoreceptive afferents synapse in the dorsal medullary nuclei cross contralaterally through the medial lemniscus.

Descending and ascending spinal pathways

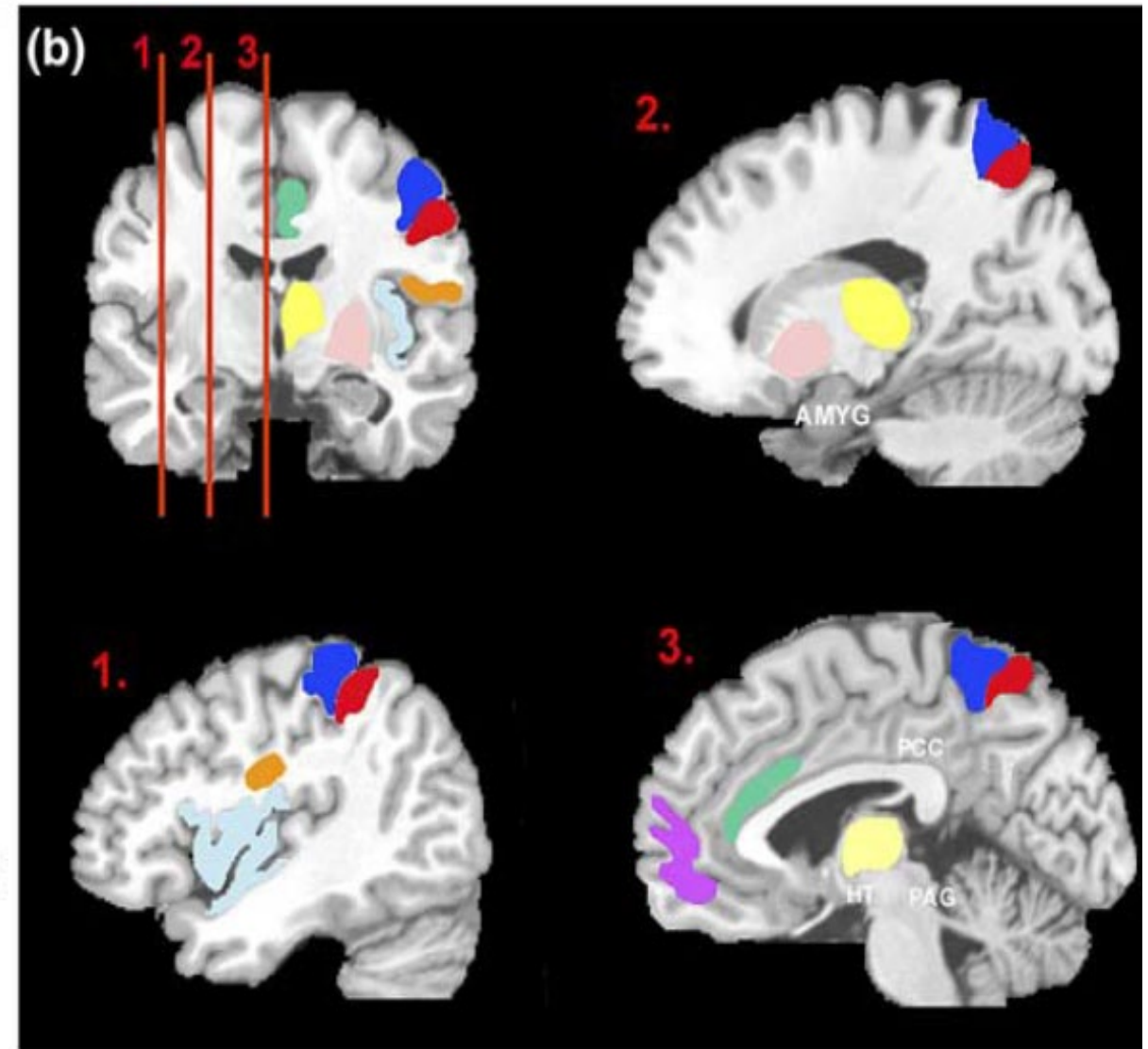
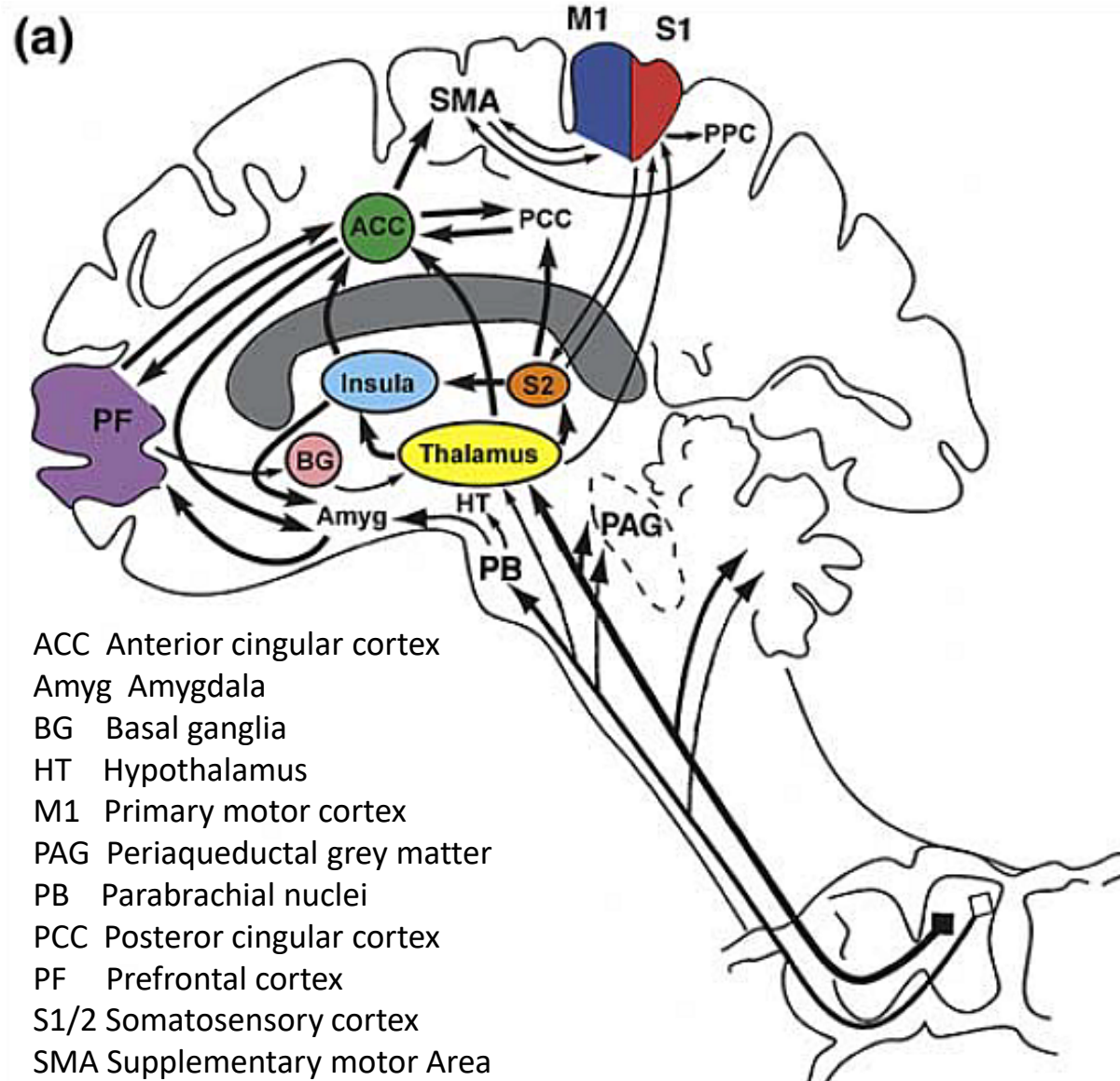


Ascending and descending spinal pathways

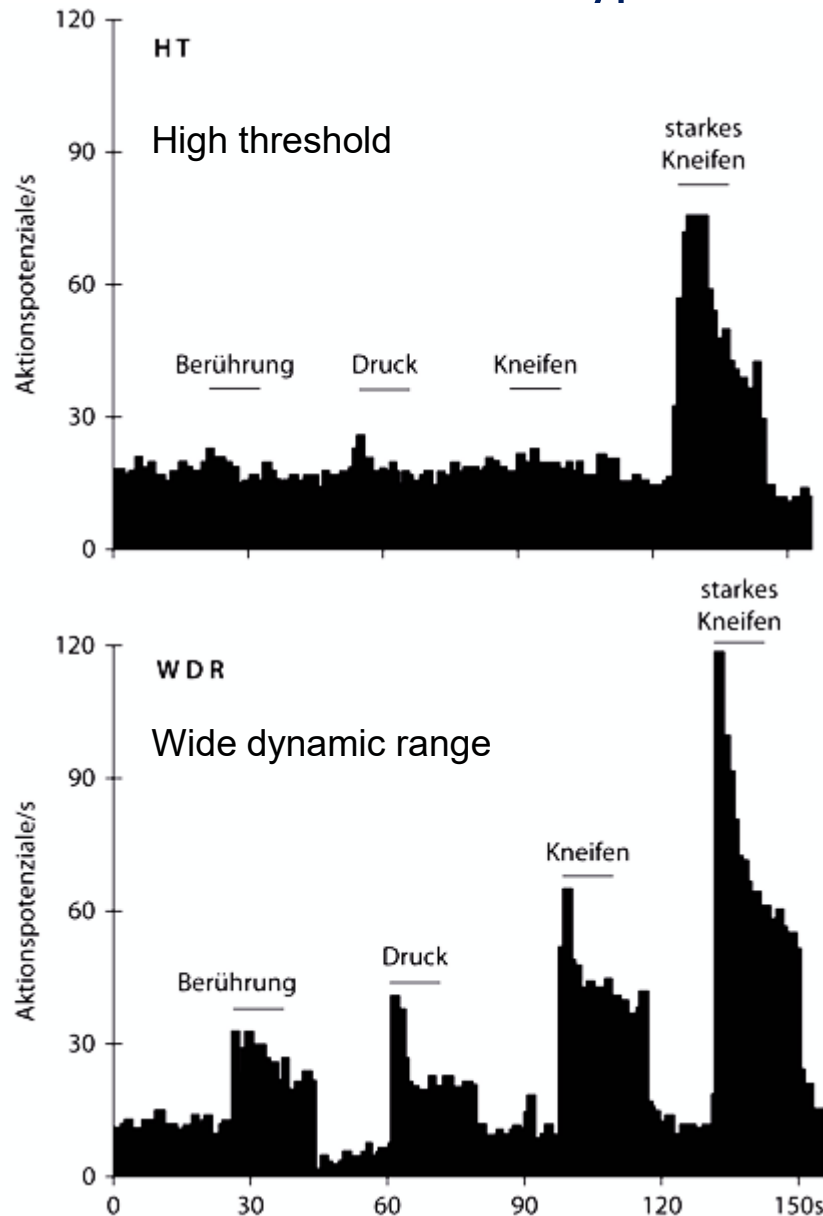
Ascending pathways



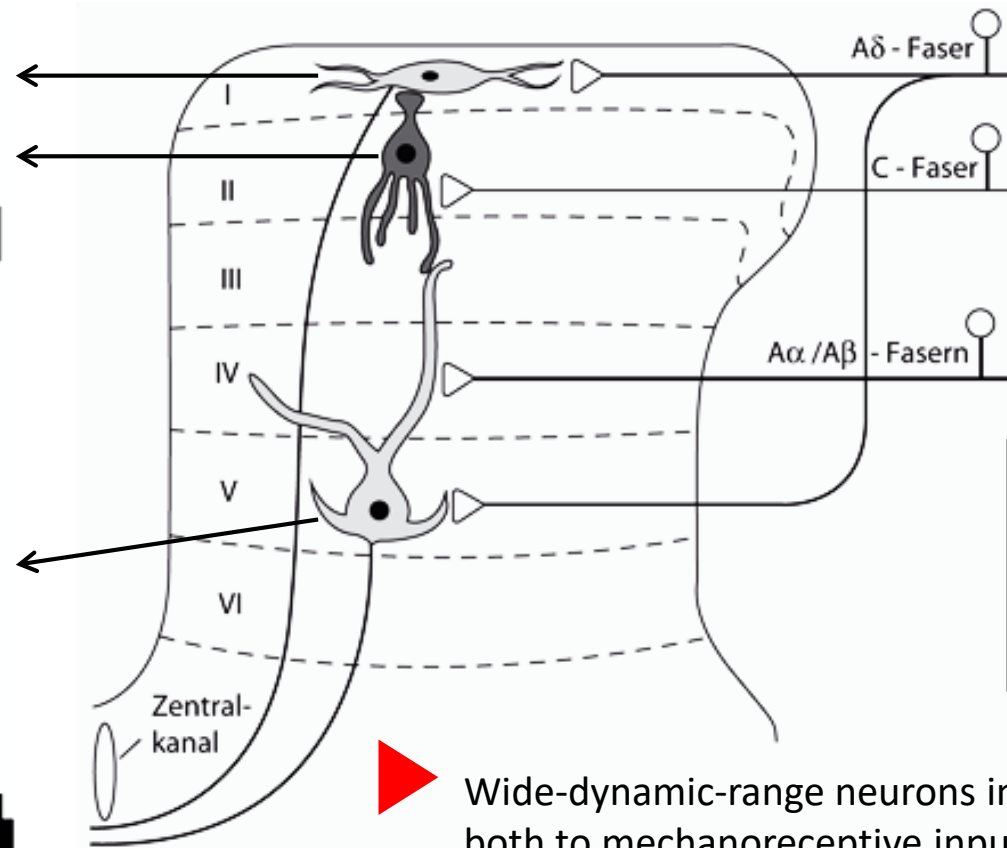
Ascending pathways and cerebral areas for pain processing



Basic types of nociceptive neurons in the spinal dorsal horn



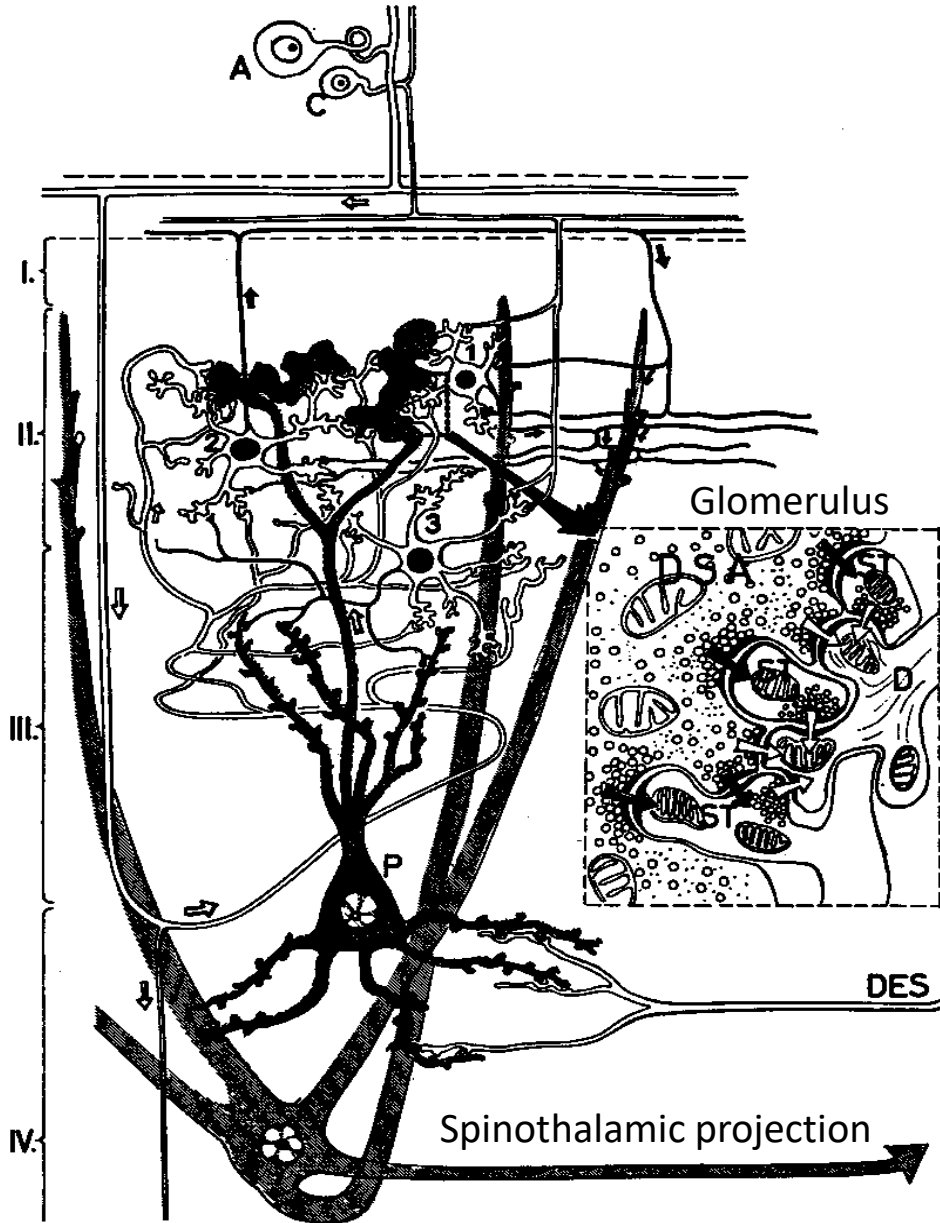
▶ Nociception-specific neurons in the superficial laminae respond specifically to nociceptive input from afferent C- and A δ -fibers.



The WDR neurons are the decisive target neurons for endogenous pain control.

▶ Wide-dynamic-range neurons in deep laminae respond both to mechanoreceptive input from afferent A-fibers as well as to nociceptive input from A δ -fibers.

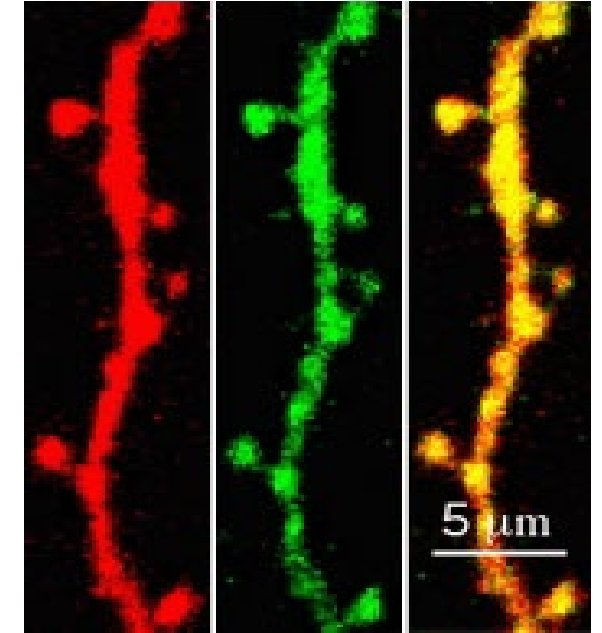
Afferent wiring in the spinal dorsal horn and synaptic spines



Willis & Coggeshall 1991

▶ The synaptic wiring in the dorsal horn is very complex. Deep laminae neurons are connected to superficial laminae by their dendrites. Synaptic contacts can be reciprocal with both pre- and postsynaptic zones. The functional properties of these glomeruli are only rudimentally known.

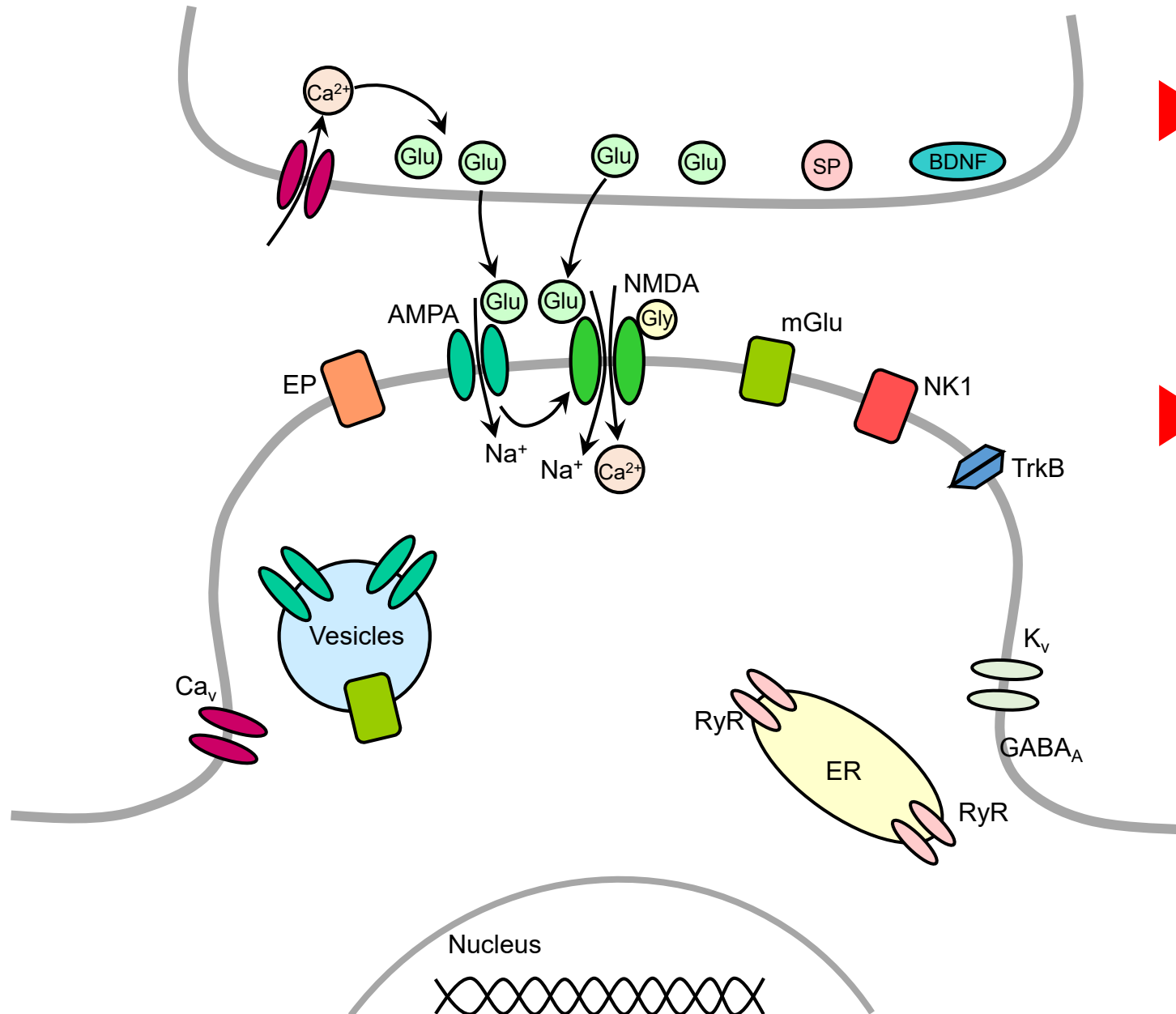
▶ Spinothalamic projection neurons are preferably in lamina IV. Descending projections (DES) are indirectly connected with projection neurons through interneurons.



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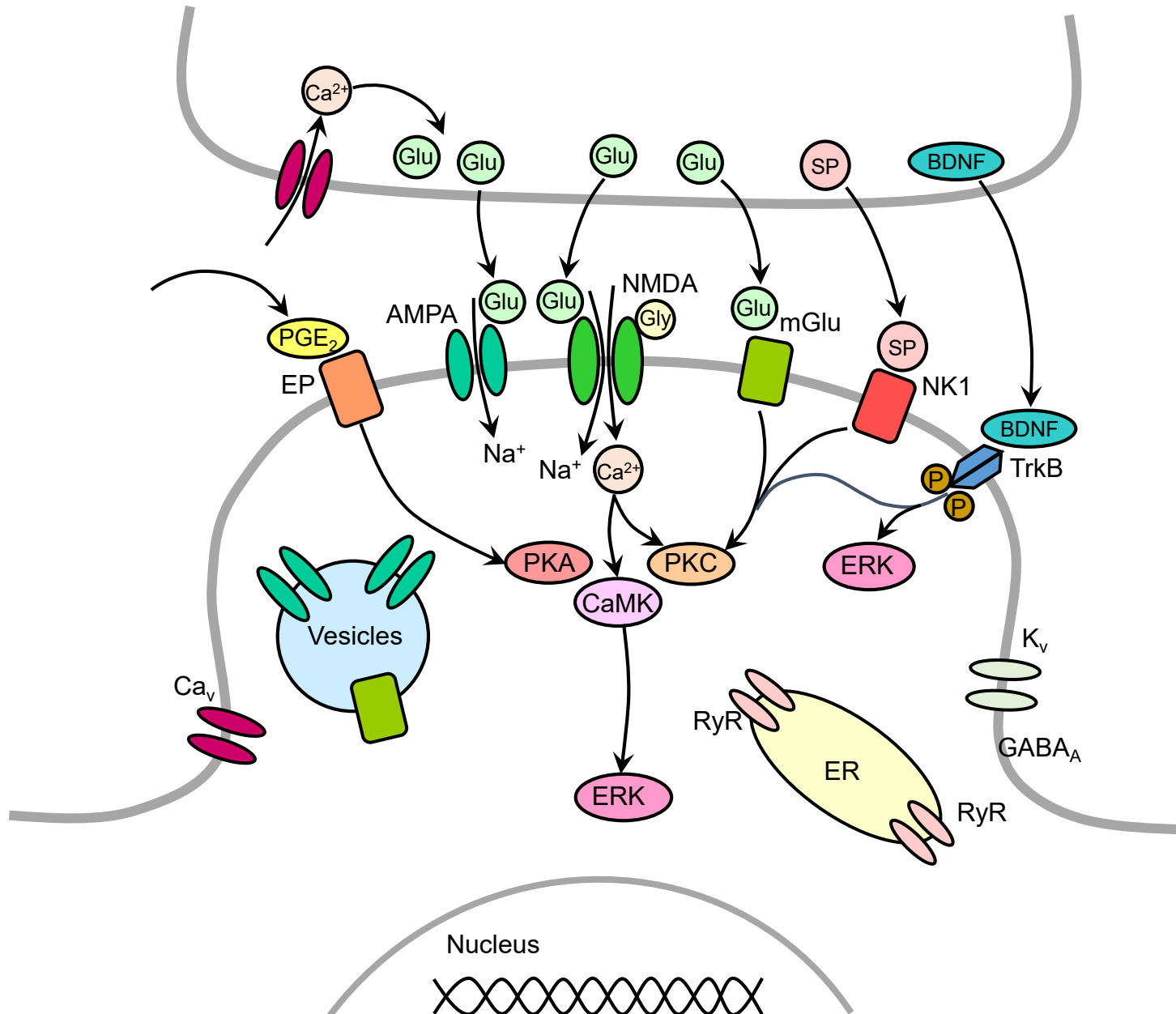
Central sensitization step 1: Activation of glutamate receptors



▶ Neurotransmitter (glutamate) release from presynaptic terminals is activated by calcium influx through voltage-dependent calcium channels.

▶ Glutamate is activating different types of glutamate receptor channels of the postsynaptic neuron leading to sodium influx and hence postsynaptic potential as well as calcium influx through N-methyl-D-aspartate channels.

Central sensitization step 2: Activation of G-protein-coupled receptors

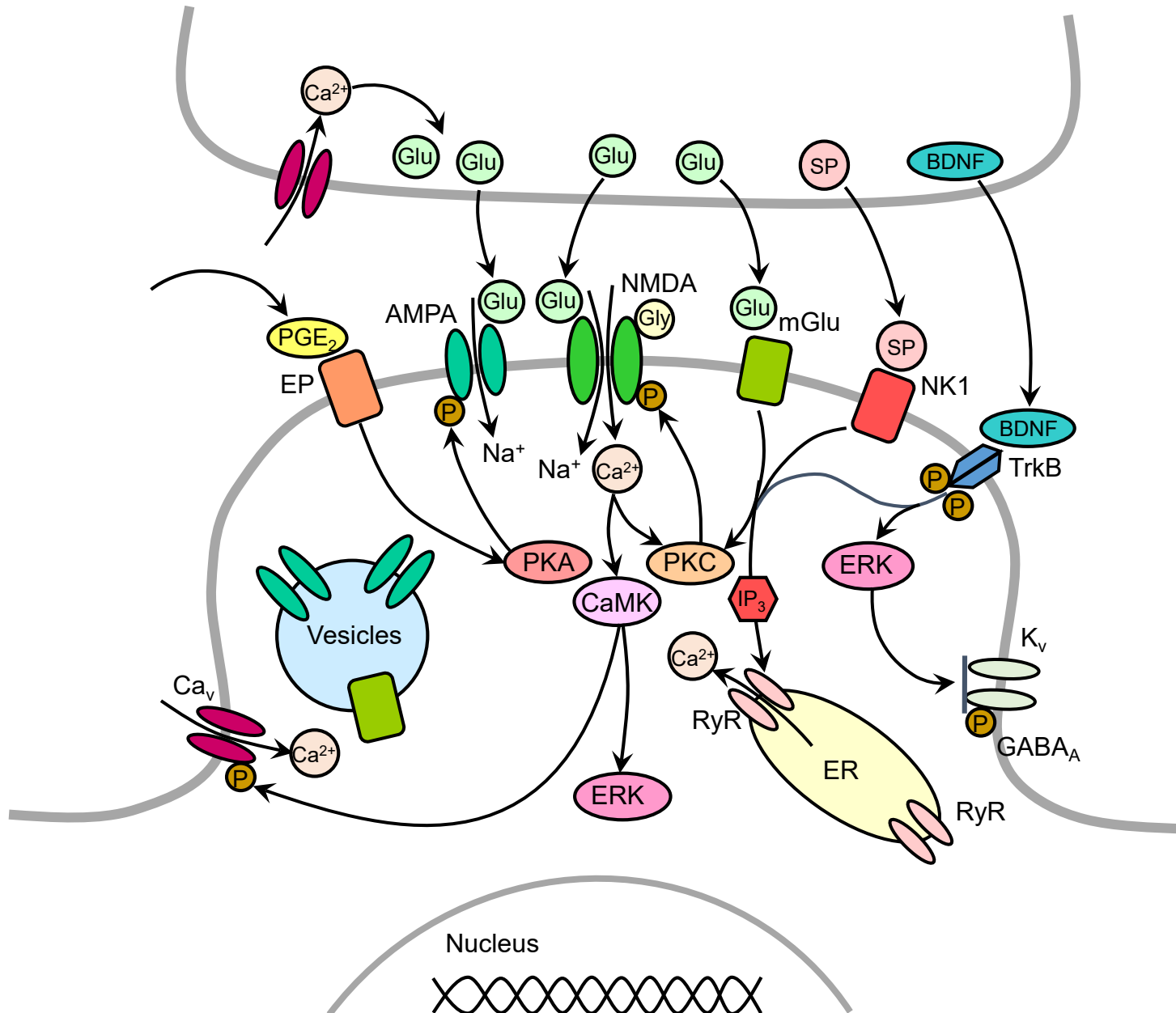


► Postsynaptic metabotropic receptors like prostaglandin receptors (EP), metabotropic glutamate receptors or neurokinin-1 receptors (binding substance P) are G-protein-coupled; through metabolic cascades, they activate different kinds of protein kinases.

Brain-derived neurotrophic factor (BDNF) activates the tyrosine receptor kinase B (TrkB), an autophosphorylating receptor.

All cascades can end up with the activation of the extracellularly regulated kinase (ERK).

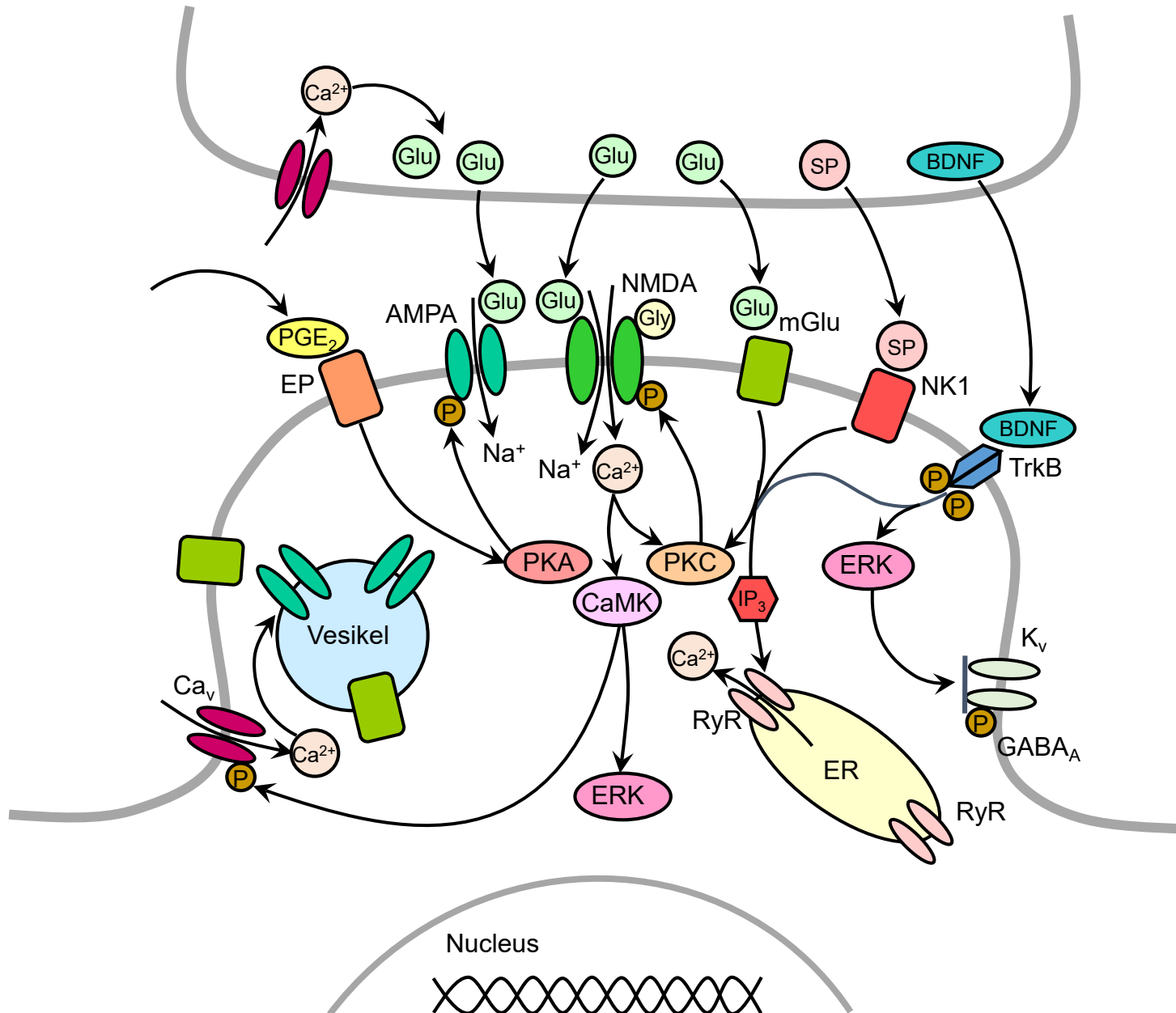
Central sensitization step 3: Metabolic cascades and action of protein kinases



► The activated protein kinases (PKA, PKC, CaMK) phosphorylate receptor channels and voltage-dependent ion channels, which changes the ion conduction and leads *summa summarum* to the facilitation of postsynaptic activation.

Intracellular increases in intracellular calcium by opening of voltage-dependent calcium channels or ryanodine receptor channels in the endoplasmic reticulum foster calcium-dependent processes.

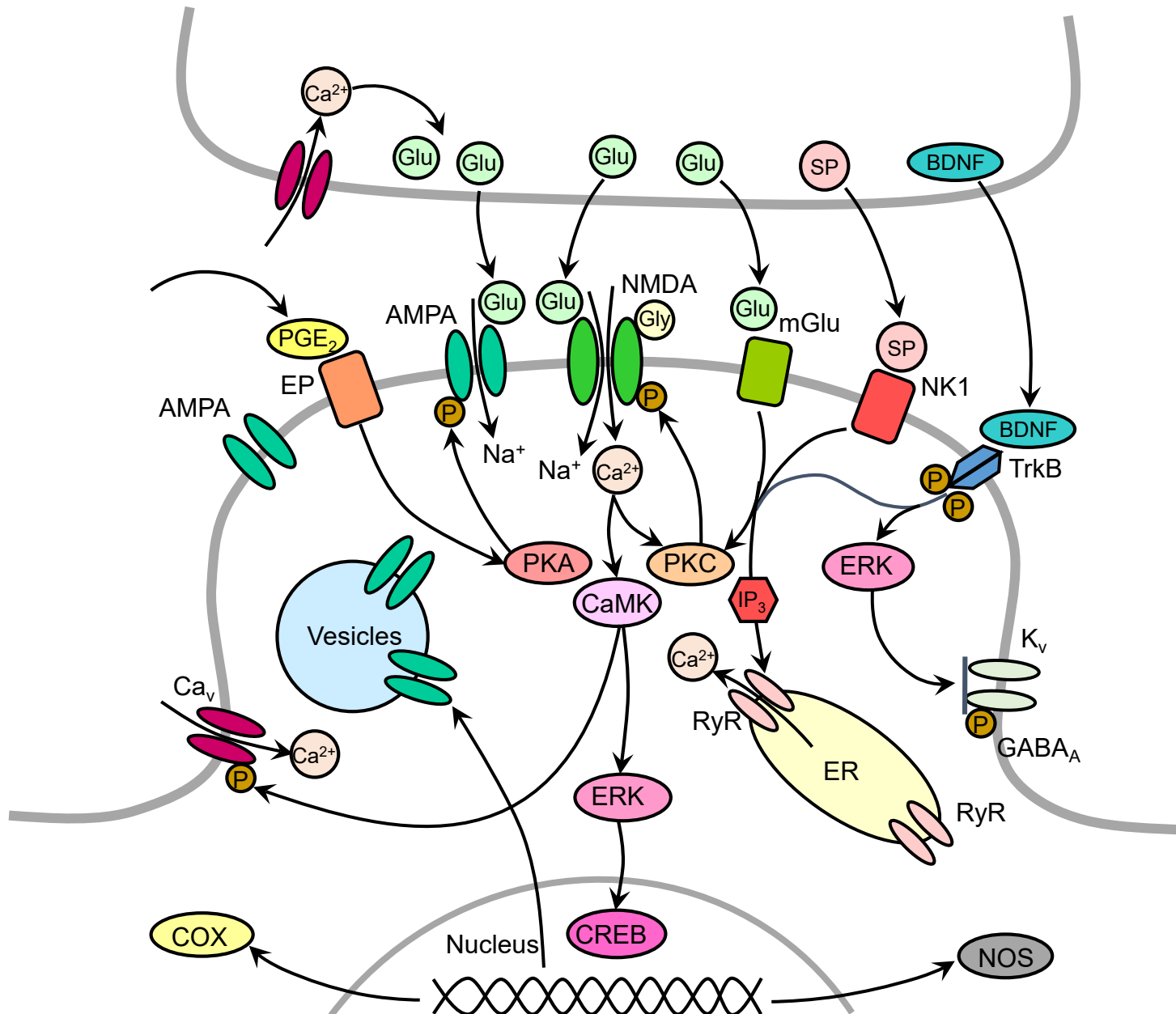
Central sensitization step 4: Recruitment of channels and receptors



▶ Long-lasting increase in calcium levels induce the integration of ion channels and receptors of different kinds (particularly glutamate receptor/channels) into the postsynaptic membrane increasing the postsynaptic responses to transmitter release.

This is the first plastic change increasing synaptic function for a long time.

Central sensitization step 5: *De novo* expression of channels and enzymes

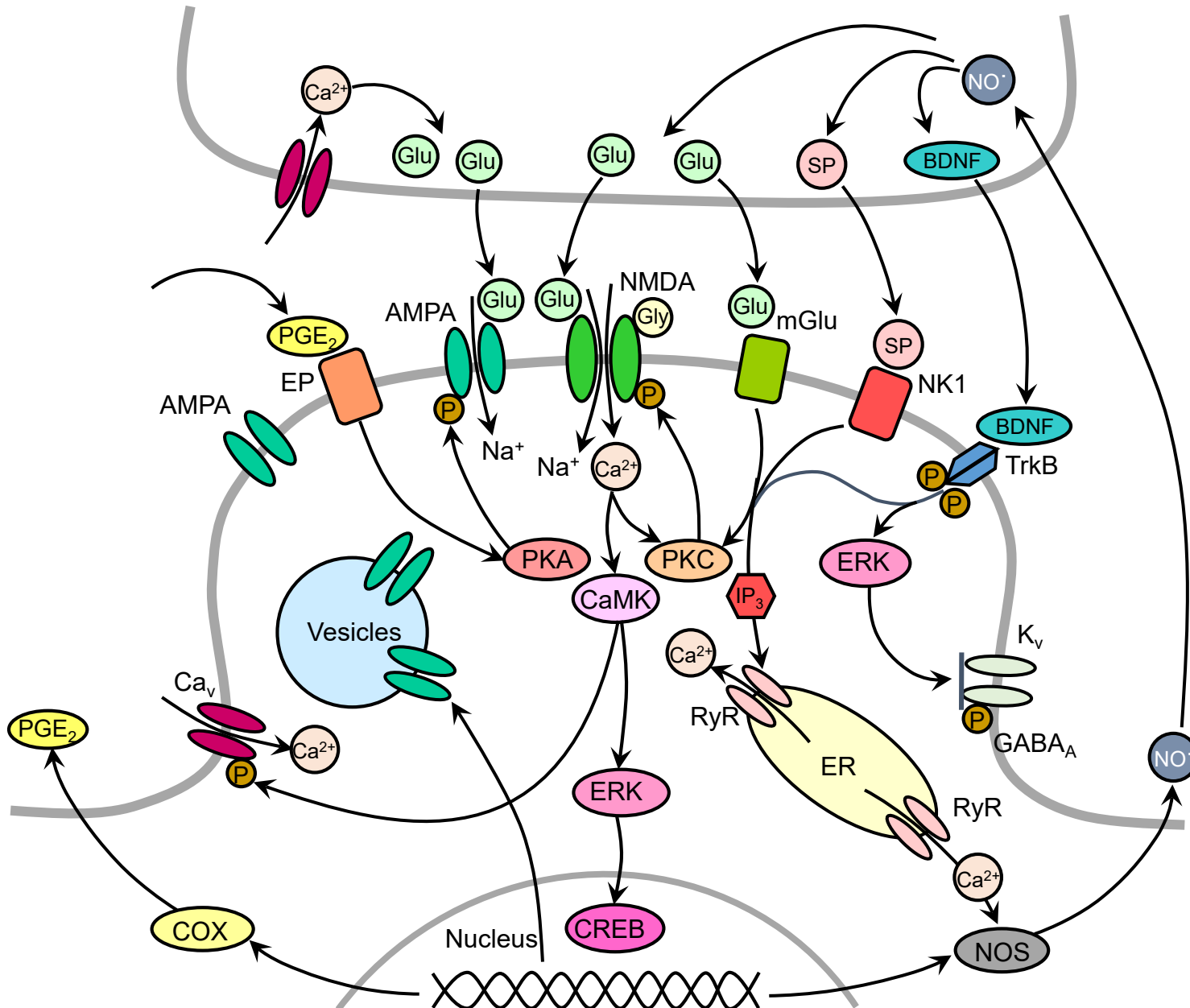


► Long-lasting increase in calcium levels induce also the activation of the extracellularly regulated kinase (ERK), which leads to the production of transcription factors like the cAMP response element binding protein (CREB).

The CREB induces transcription of genes encoding proteins for new channels and receptors that are stored in vesicles.

This is another important step of postsynaptic plasticity, which cannot be reversed immediately.

Central sensitization step 6: Production of nociceptive mediators and „retrograde transmitters“

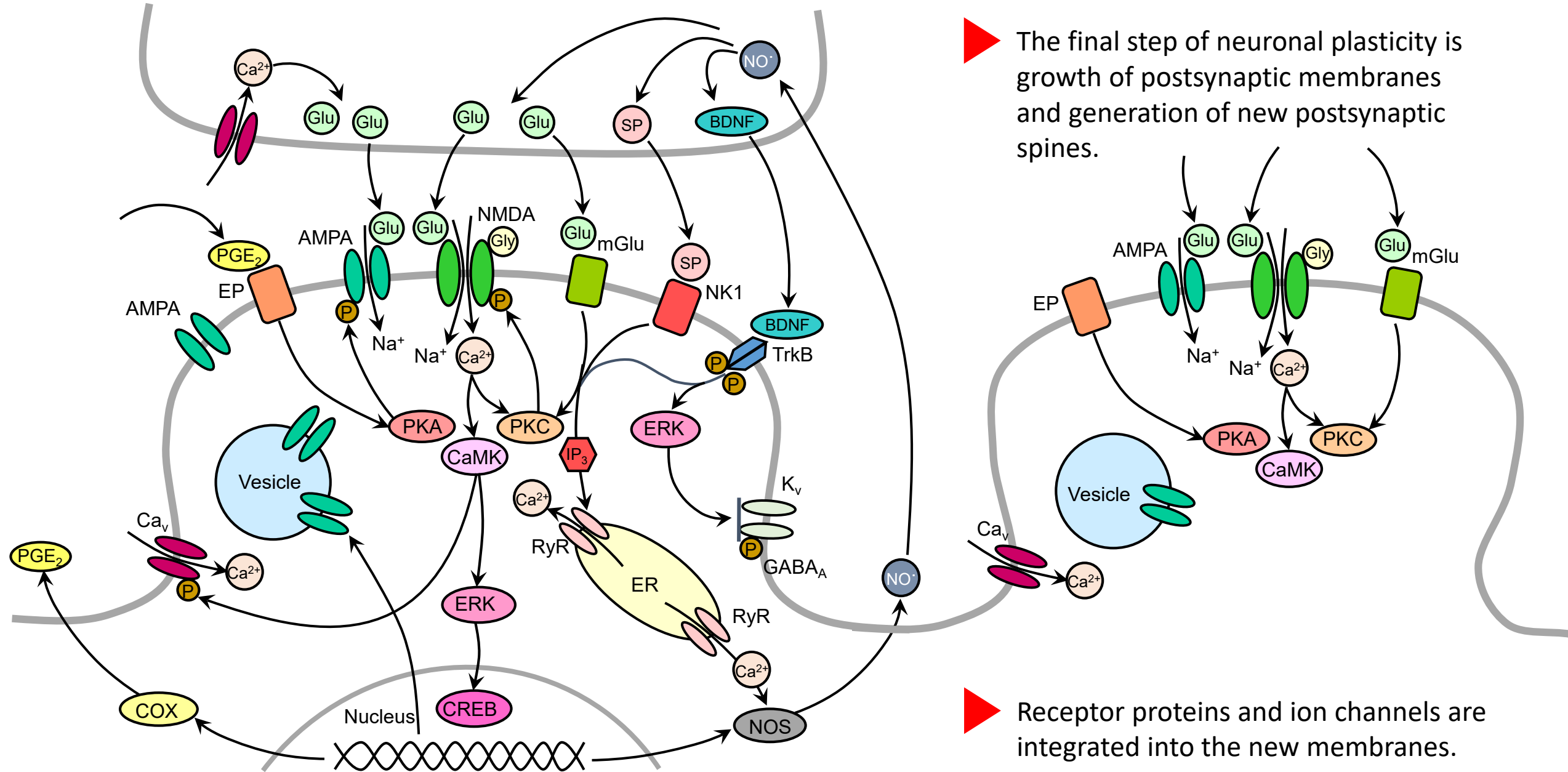


► Gene expression includes also transcription of enzymes, which give rise to the production of nociceptive mediators like prostaglandins (PGE₂) and „retrograde transmitters“ like nitric oxide (NO).

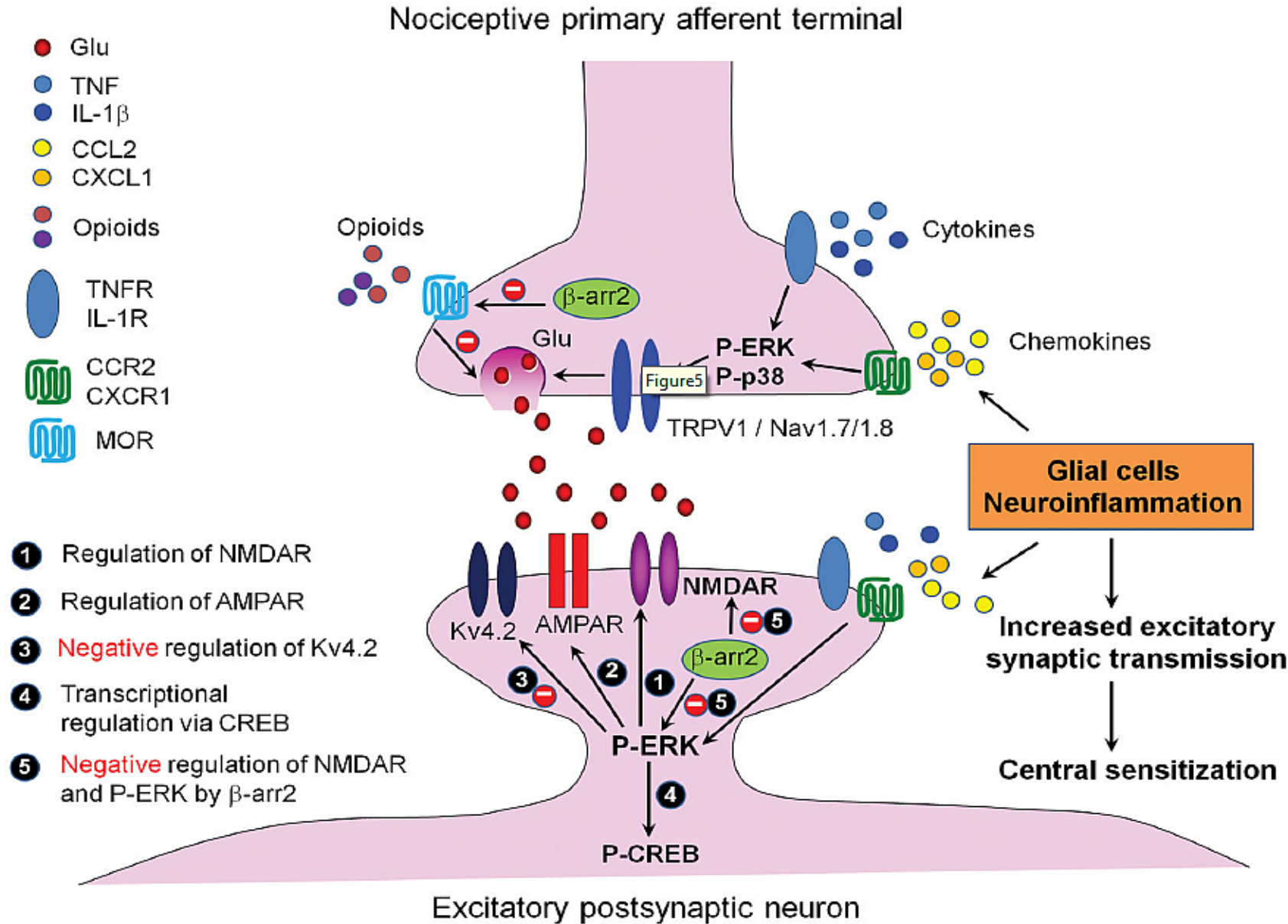
NO is rapidly diffusing through cell membranes, reaches the presynaptic terminals and may facilitate the production and release of neurotransmitters.

PGE₂ can contribute to reciprocal sensitization of neurons. Glial cells may be other targets of these mediators.

Central sensitization step 7 (final): Formation of new postsynaptic spines – plasticity of synapses

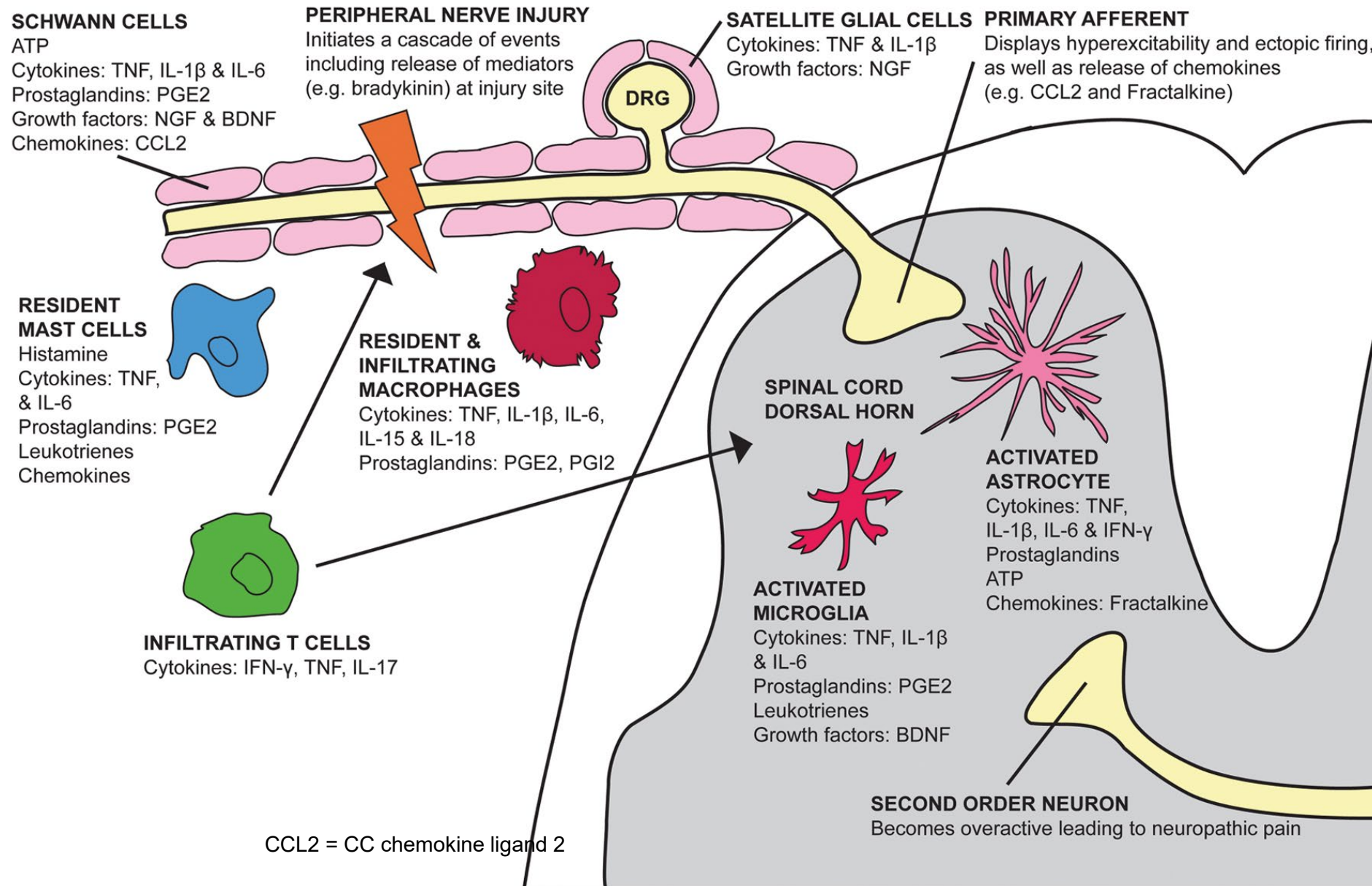


Neuroinflammation and central sensitization involving glial cells (microglia, astrocytes)



▶ Glial cells involved in neuroinflammation can produce different kinds of chemokines (cytokines) that act both pre- and postsynaptically on central neurons by regulating ion channels and G-protein-coupled receptors.

Central sensitization mechanisms in neuropathic pain

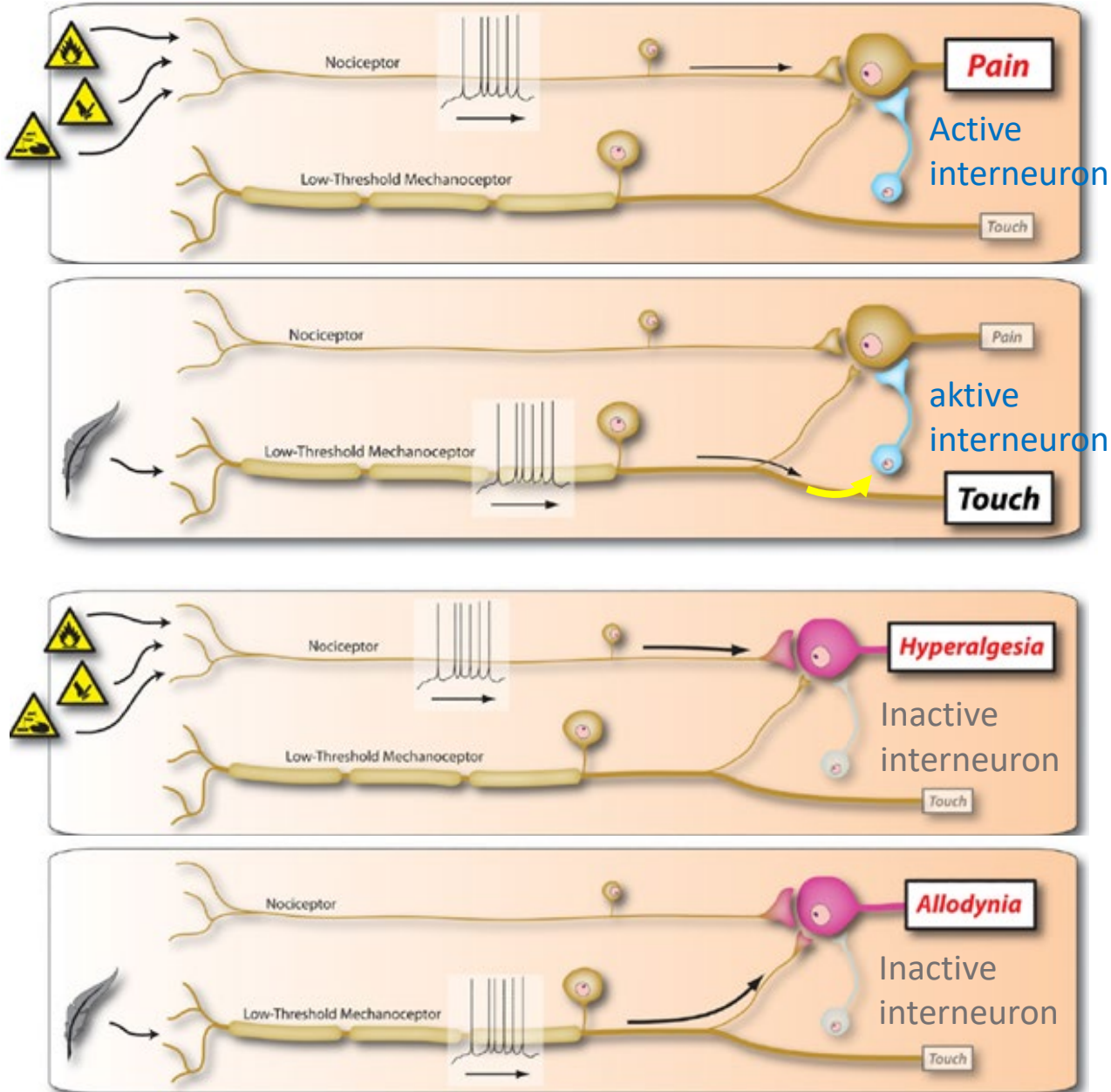


Several types of glial cells in the dorsal horn and infiltrating mononuclear cells can produce cytokines and other mediators under pathological conditions thereby contributing to central sensitization.

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Spinal mechanoreceptive and nociceptive projection to second order neurons

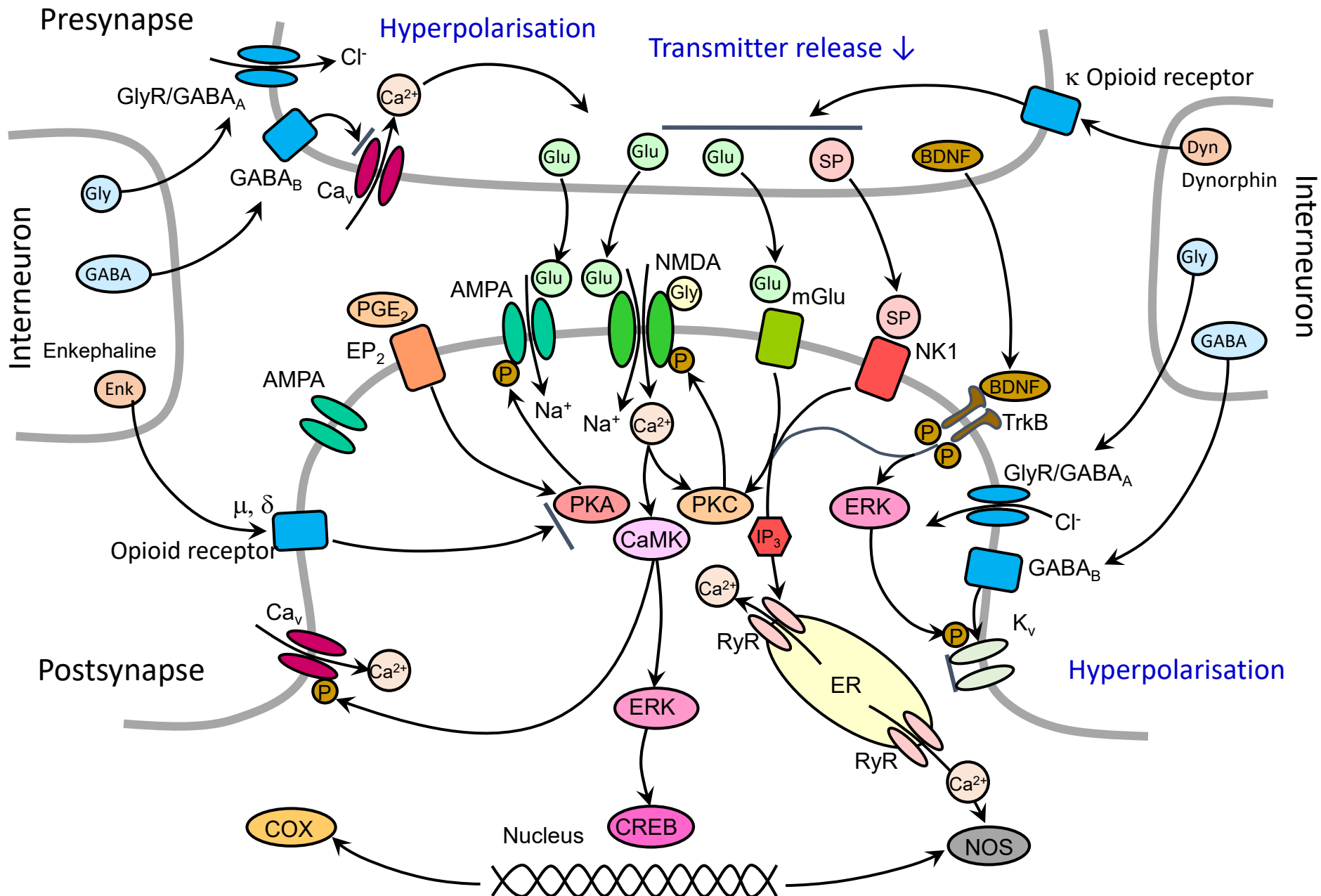


► WDR second order neurons receive input from both nociceptive and mechanoreceptive afferents. Inhibitory interneurons can modulate the activity of WDR neurons.

► Activated mechanoreceptive afferents ($A\beta$) can modulate the activity of WDR neurons through (inhibitory) interneurons („gate control“). The exact mechanism is not clear.

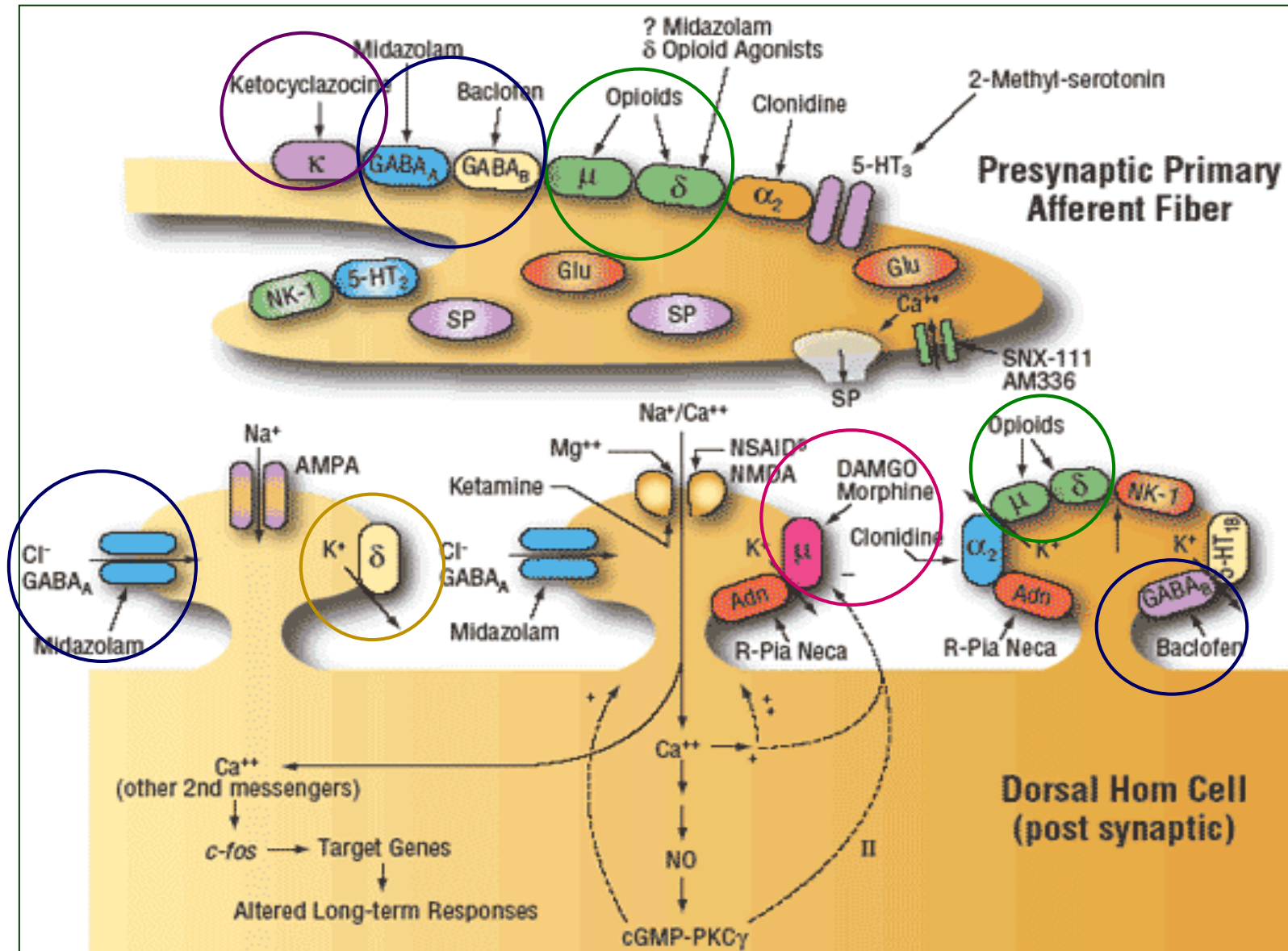
► Chronic pain syndromes like hyperalgesia and allodynia can significantly depend on the inactivation of inhibitory interneurons.

Inhibitory neurotransmitter actions at spinal neurons



► Inhibitory neurons in the spinal dorsal horn operate with neurotransmitters, which bind pre- or postsynaptically to inhibitory receptors like GABA, glycine or opioid receptors. On principle they counteract the excitatory mechanisms, i.e., they increase the membrane potential (hyperpolarisation) through inflow of Cl^- or outflow of K^+ , inhibit voltage-dependent Ca^{2+} channels and reduce the activity of sensitizing protein kinases.

Inhibitory receptor systems in the dorsal horn



► Besides the classical inhibitory neurotransmitters and receptors (GABA, glycin) three types of opioid receptors (μ , δ and κ) are involved in the pre- and postsynaptic inhibitory mechanisms.

Multiple receptor systems can contribute to the synaptic transmission in the spinal dorsal horn.

Endogenous opioids, opioid receptors and effects

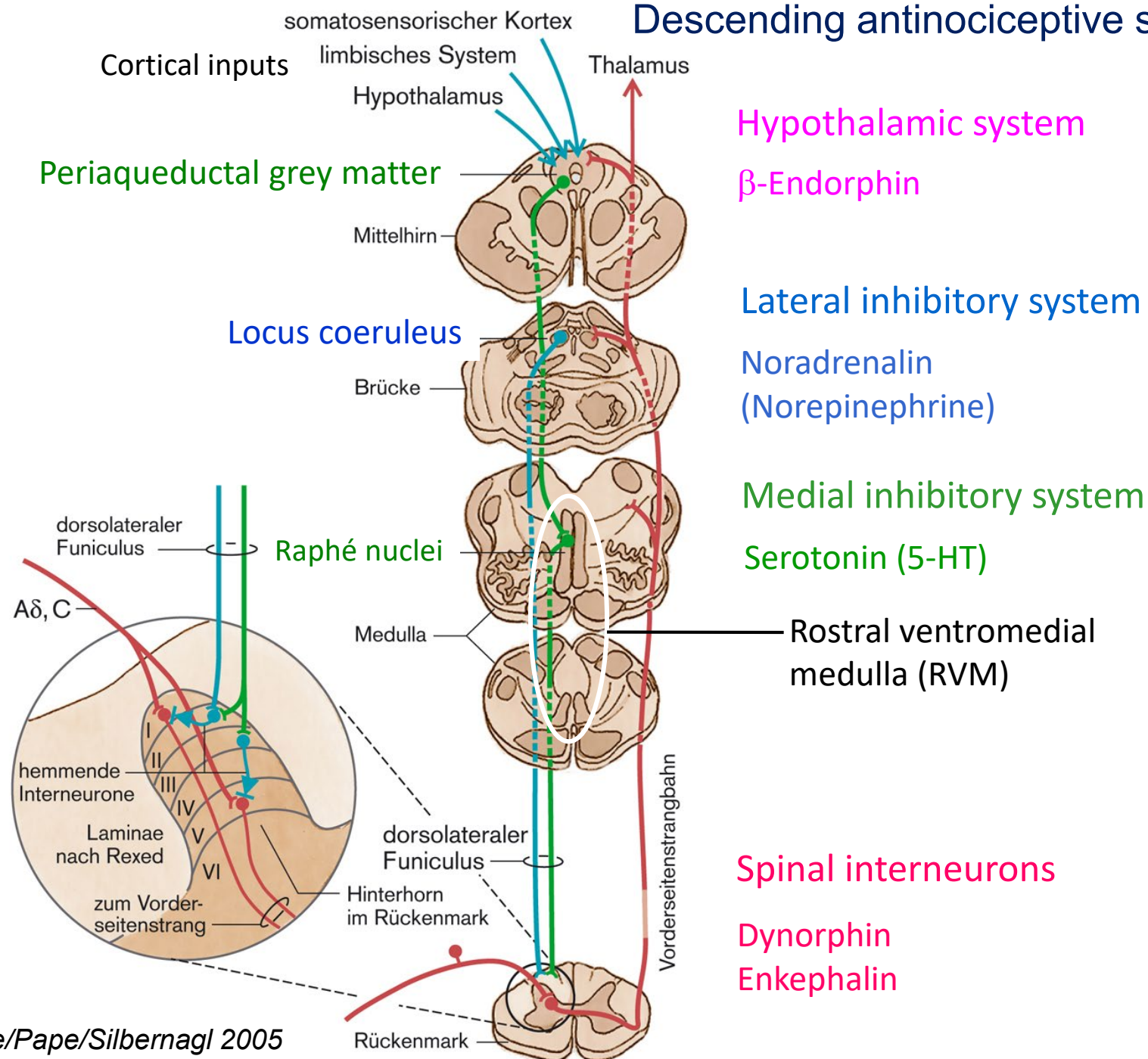
Endogenous opioids	Receptors	Production sites
Endorphins (β -Endorphin)	μ	Hypothalamus, Hypophysis
Enkephalins (Met-, Leu-Enkephalin)	μ , δ	CNS, Adrenal gland, Gut
Dynorphins (Dynorphin A, B)	κ	CNS, Gut

Receptors	Expression sites	Effects
μ_1	Brain	Analgesia , Bradykardia, Hypotonia
μ_2	Brain, Spinal cord, Peripheral organs	Analgesie , Constipation, Euphoria, Respiratory depression
κ	Brain, Spinal cord	Analgesia , Sedation, Dysphoria
δ	Brain, Spinal cord, Peripheral organs	Analgesia , Constipation
?		Miosis, Nausea

Adressed topics

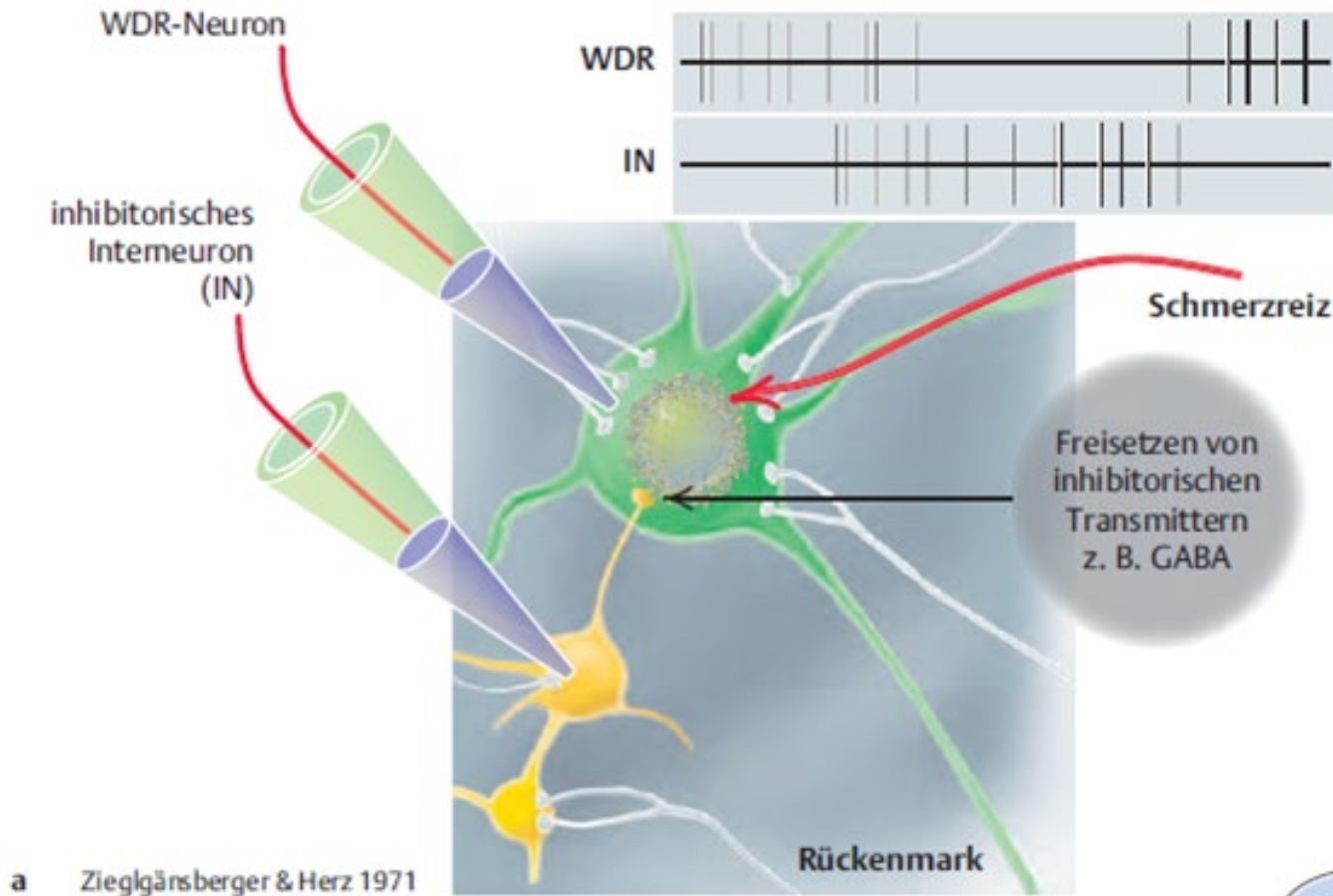
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Descending antinociceptive systems

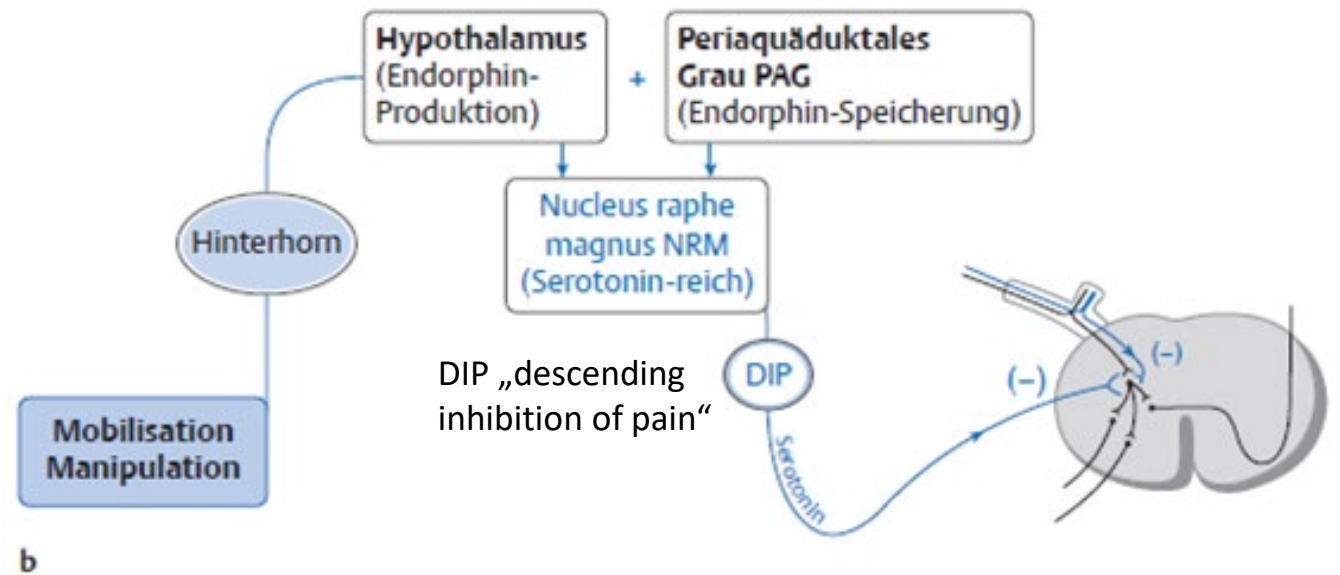


The descending pain modulating systems are classically divided into a more laterally descending **noradrenergic system** and a more medially descending **serotonergic system**. Both systems interact with neurons in the rostral ventromedial medulla (RVM). The descending axons synapse to **spinal interneurons**, many of which operate with opioids as neuromodulators. Opioids are also released from the **hypothalamus**, operating as hormones.

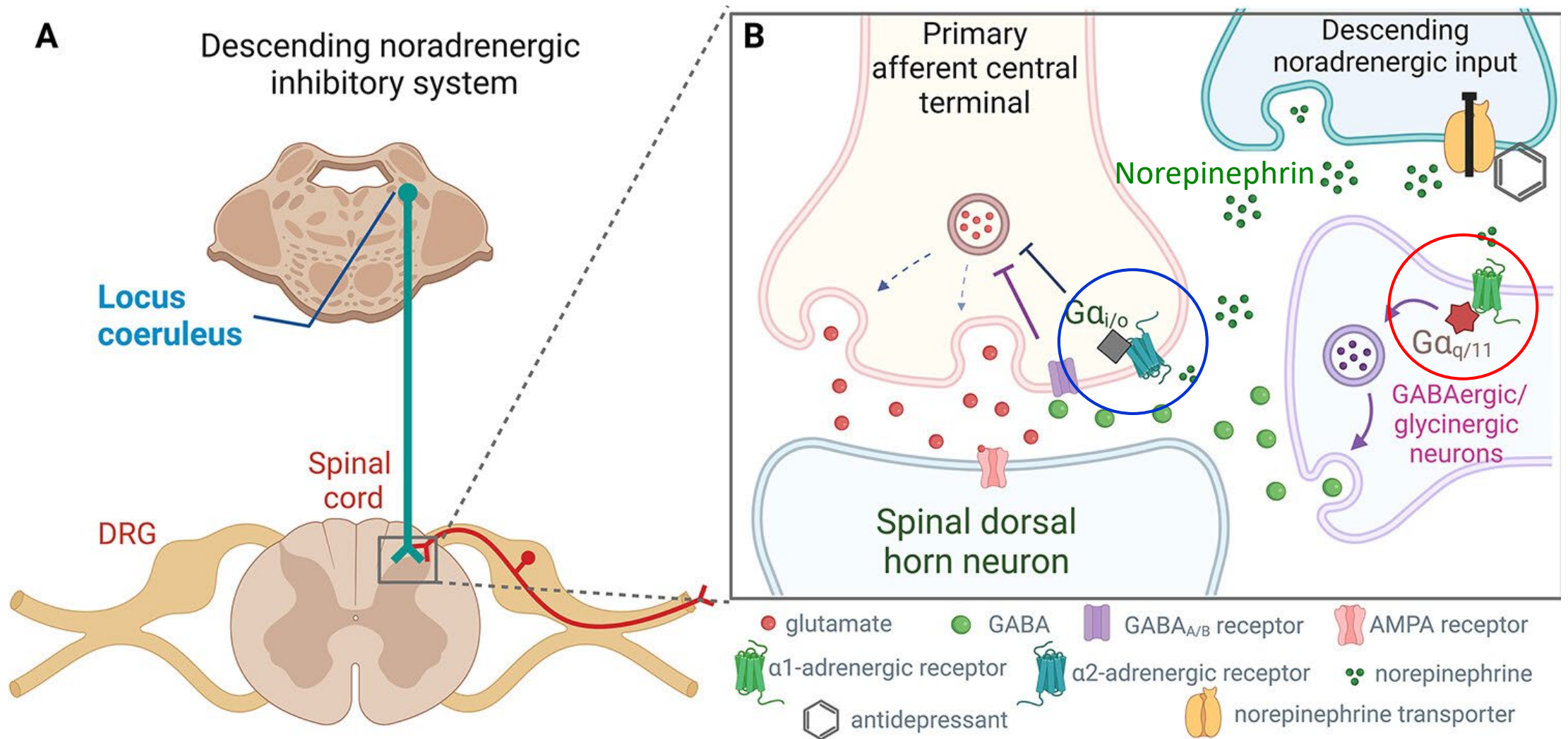
Basic principle of spinal WDR neuron inhibition in the dorsal horn



► Inhibition of spinal dorsal horn WDR neurons is managed by interneurons releasing inhibitory neurotransmitters (mainly GABA). Interneurons are activated by descending neurons, e.g. serotonergic neurons from the raphe nuclei in the midbrain, which receive descending input from the hypothalamus and the periaqueductal grey matter (PAG).



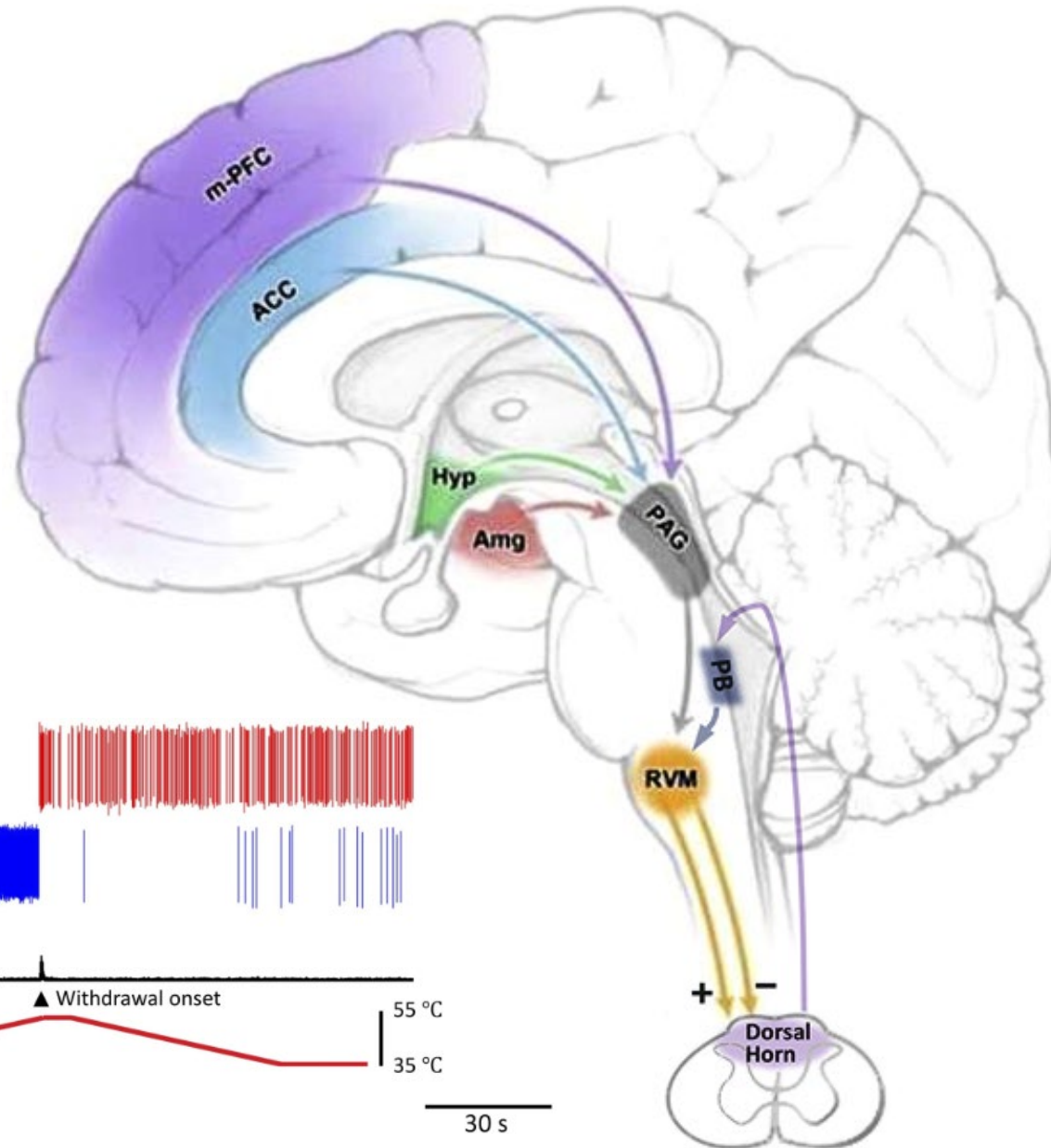
Transmission effects of the descending noradrenergic system



- Norepinephrin released from descending noradrenergic neurons activates α -receptors coupled with inhibitory G-proteins on primary afferent terminals (presynaptic) decreasing neurotransmitter release and α -receptors coupled with excitatory G-proteins on inhibitory interneurons causing increase in GABA and glycin release.

Cortical inputs to the descending control system

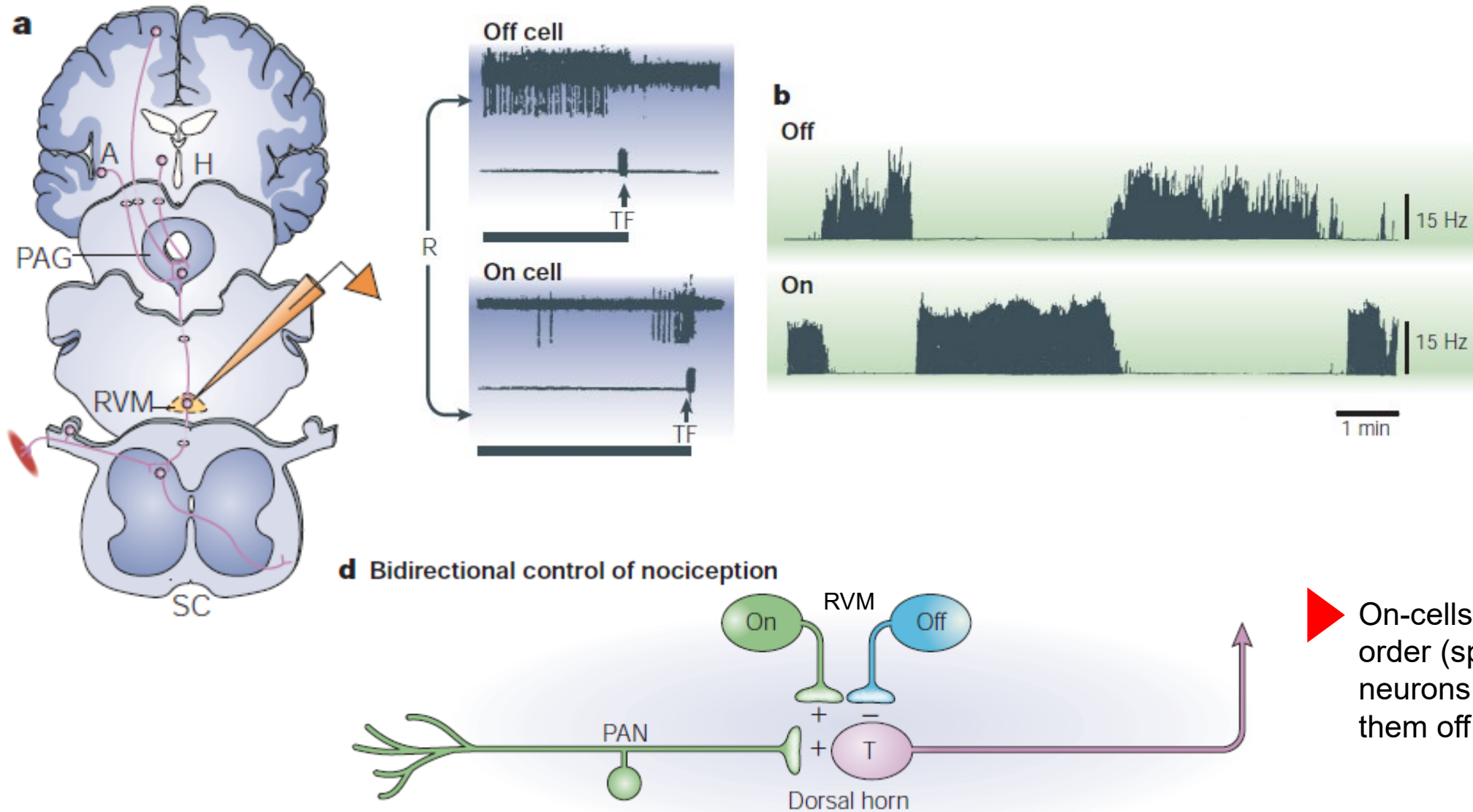
ACC, anterior cingular cortex
Amg, Amygdala
Hyp, Hypothalamus
m-PFC, medial prefrontal cortex
PAG, periaqueductal grey matter
PB, parabrachial nuclei
RVM, rostral ventromedial medulla



The rostral ventromedial medulla (RVM) contains a network of activating and inhibitory neurons which project to the spinal dorsal horn. Via the periaqueductal grey matter (PAG) the RVM receives cortical input from different areas processing *rational* (m-PFC), *emotional* (ACC) *autonomic* (Hyp) and *affective* information (Amg).

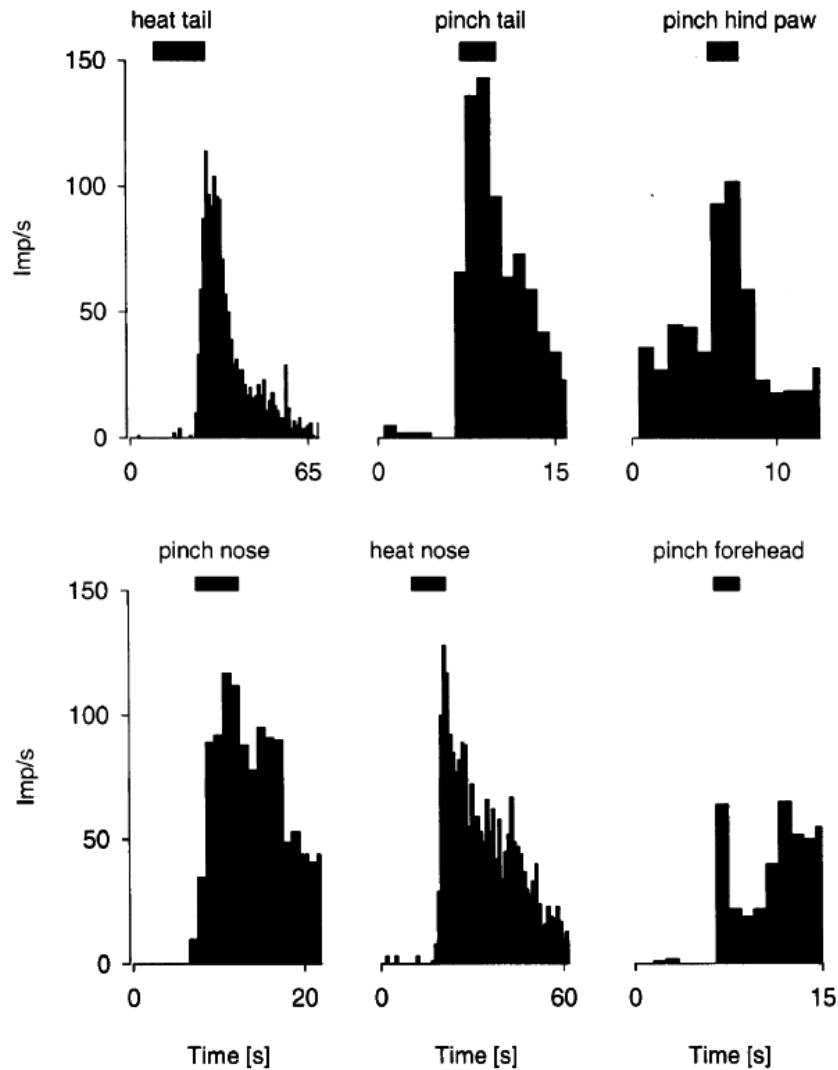
This means that descending control can be influenced by conscious, emotional and autonomic motives.

On- and Off-cells in the Rostral Ventromedial Medulla (RVM) and their impact on dorsal horn neurons

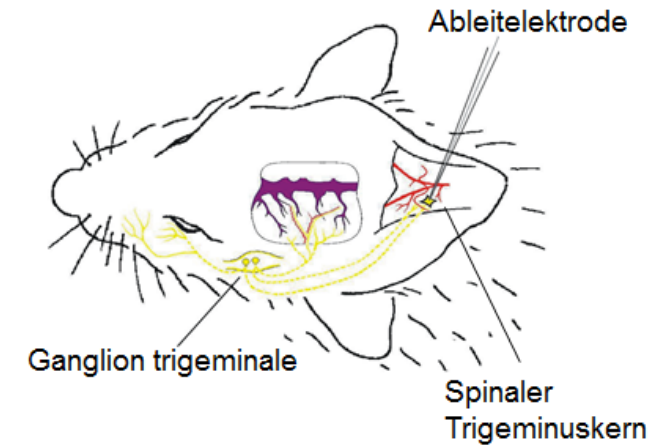
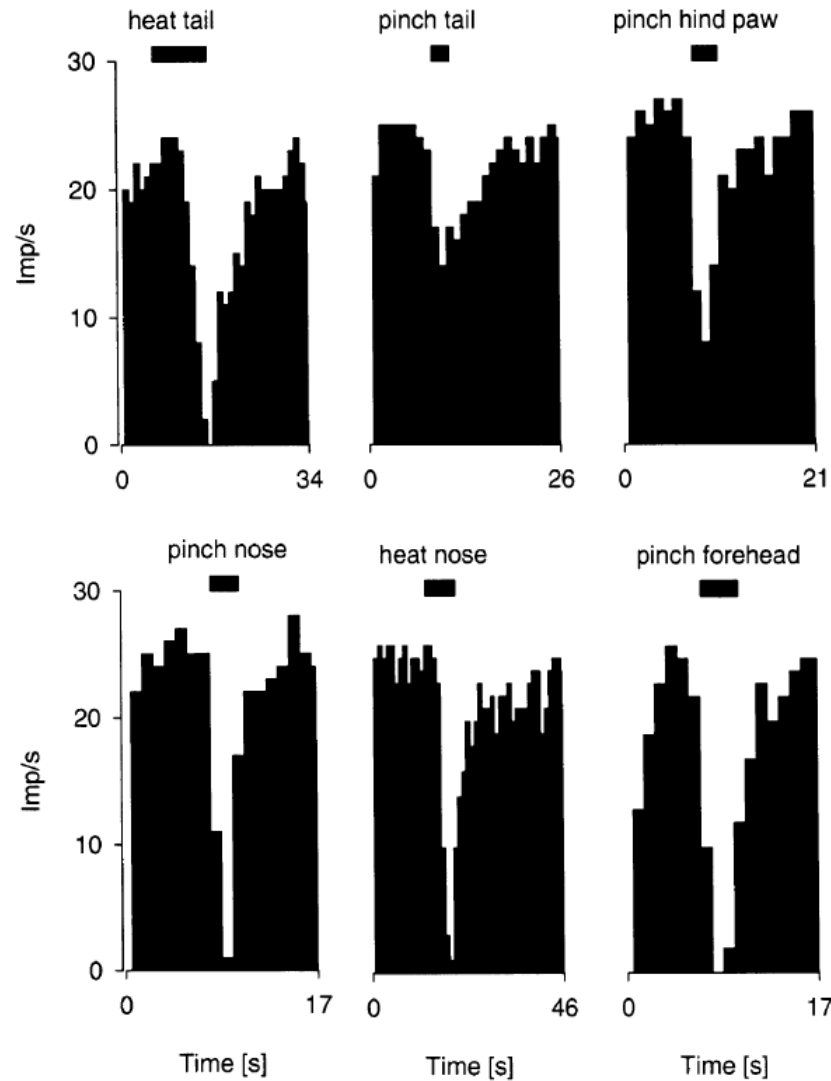


Activity of On- and Off-cells in der RVM during different noxious stimuli

On-cell



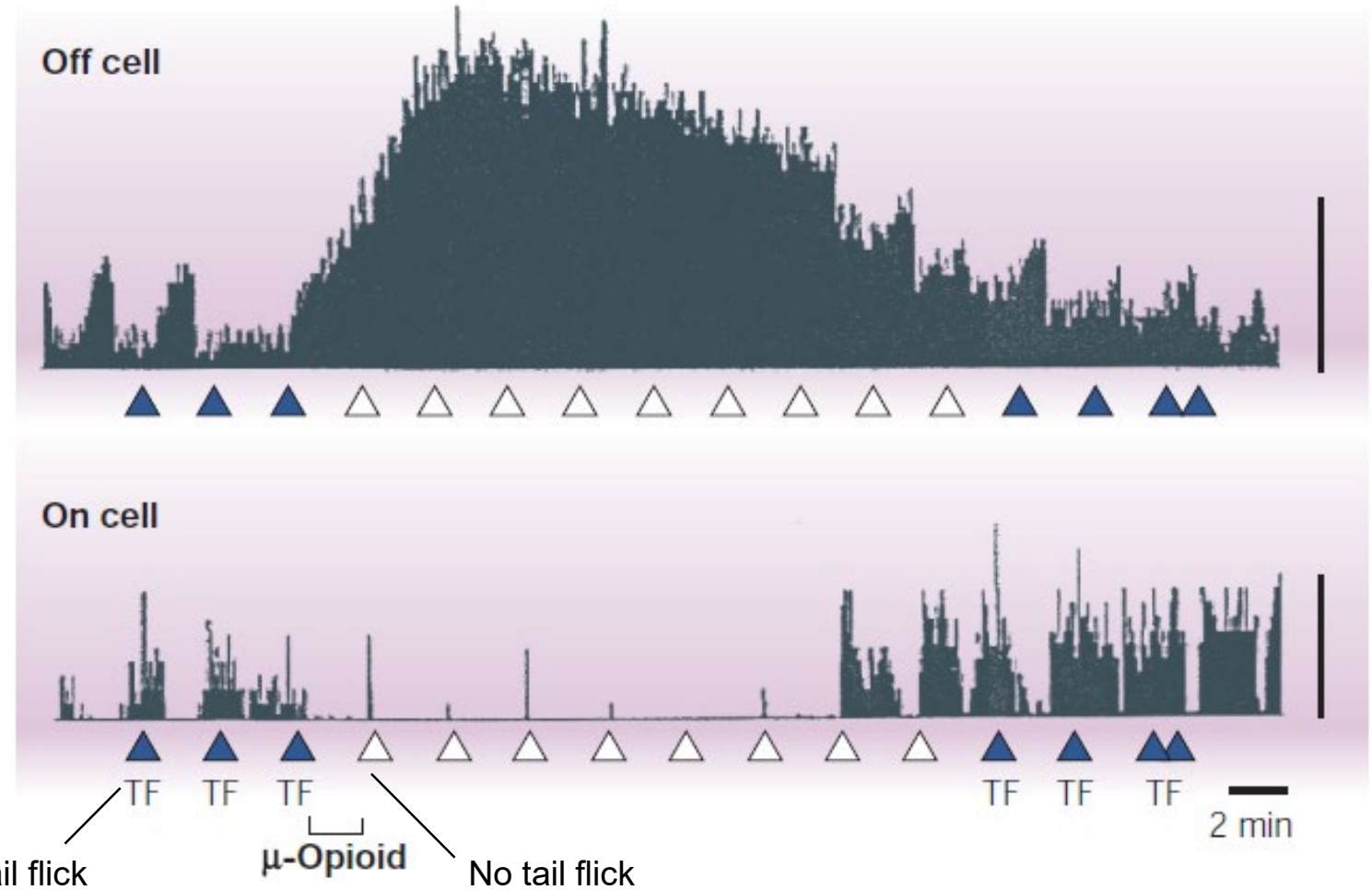
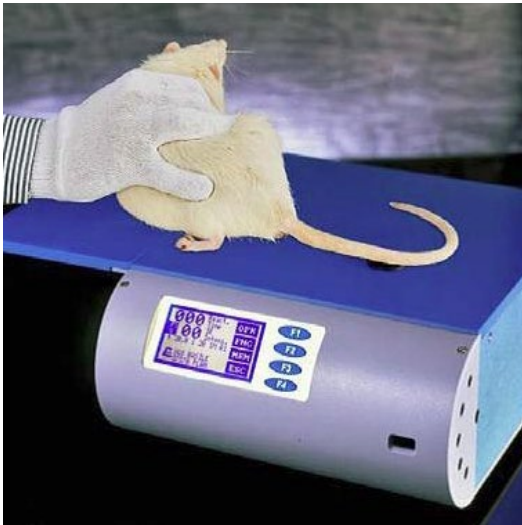
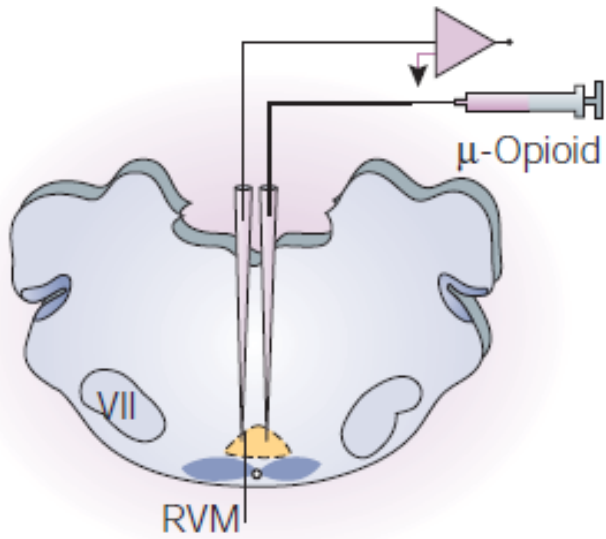
Off-cell



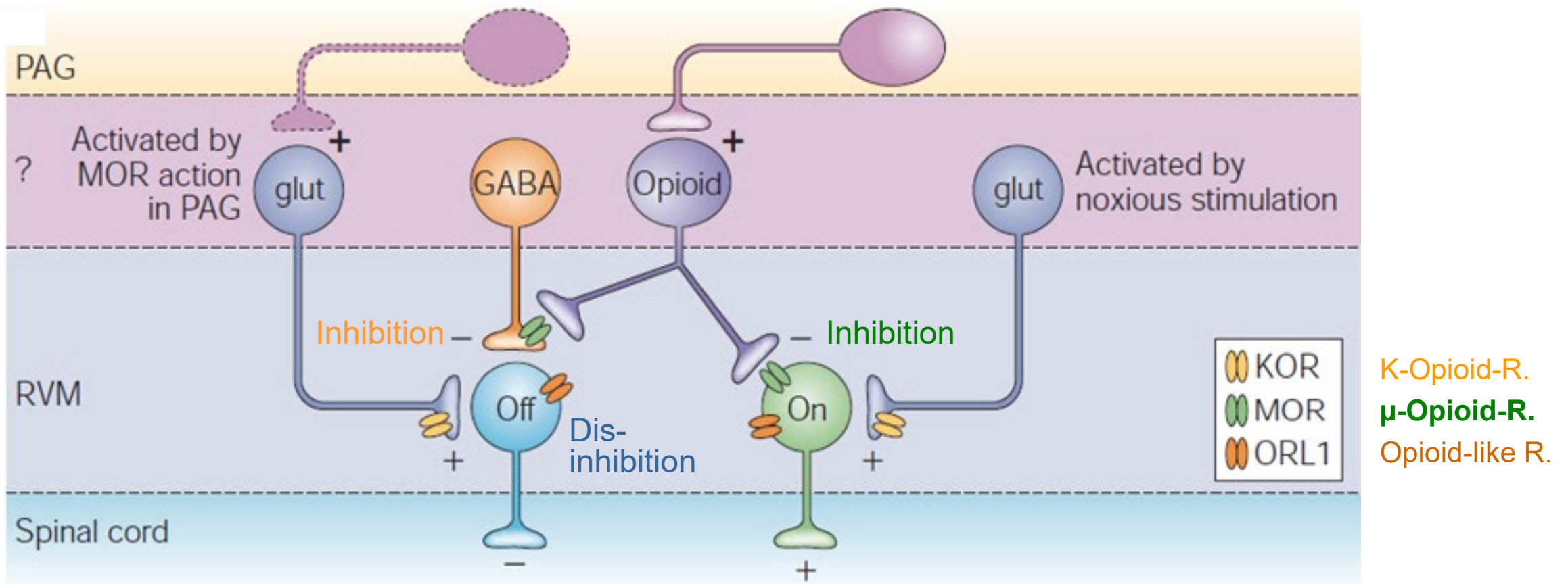
► On-cells show usually no ongoing activity but are activated during noxious stimulation of nearly all body areas. Off-cells are spontaneously active and are silenced upon noxious stimulation.

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Morphin activates Off-cells and inhibits On-cells in the RVM



Effects of opioids on On- and Off-cells in the rostral ventromedial medulla

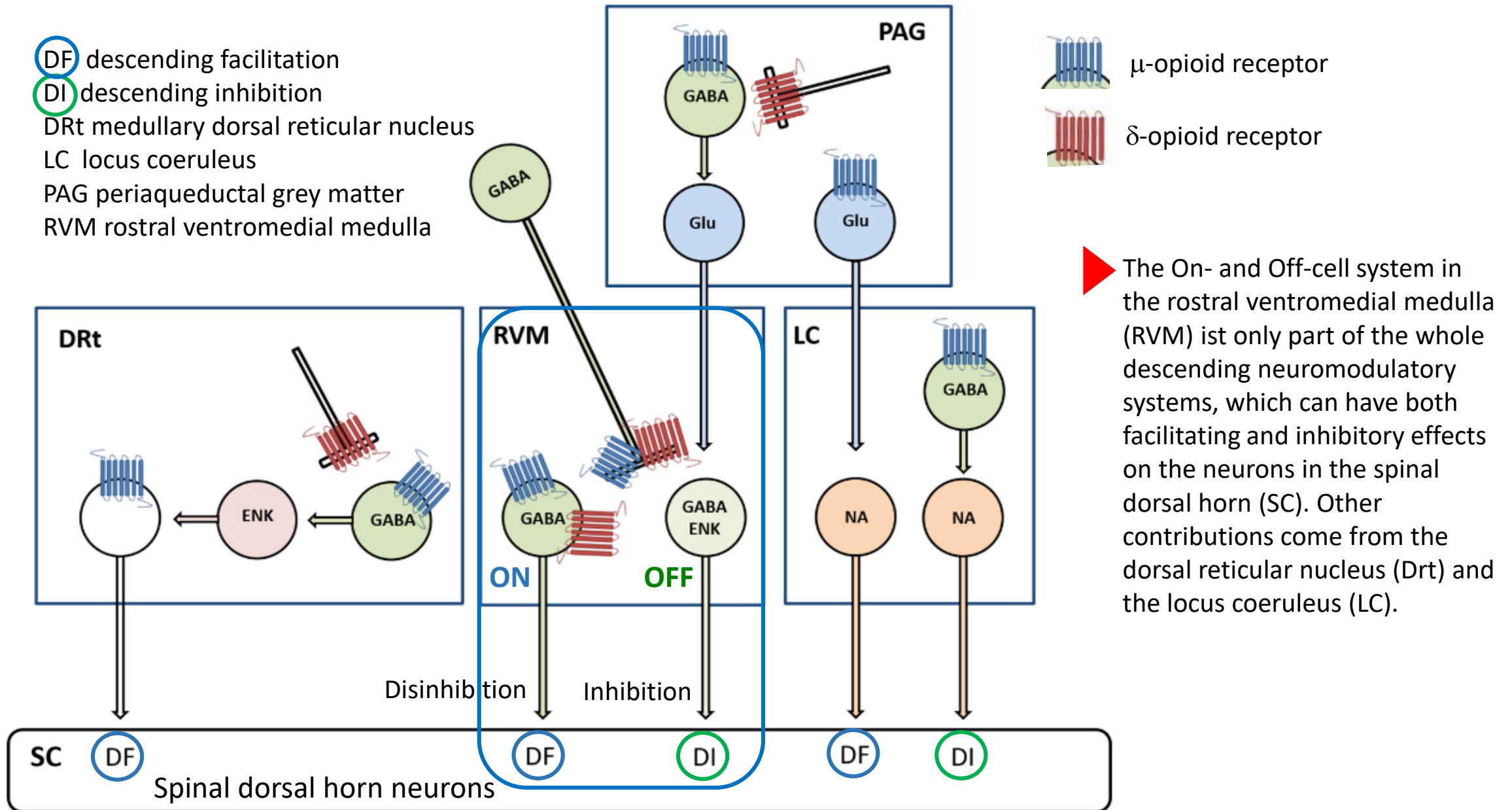


► **On-cells** in the RVM are activated by nociceptive input and can be inhibited by opioids directly (postsynaptic mechanisms).

Off-cells in the RVM are under GABAergic inhibitory control and are disinhibited by opioidergic neurons (presynaptic inhibition of GABAergic neurons).

The On- and Off-cell system may explain immediate pain control by modulating the activity of spinothalamic neurons.

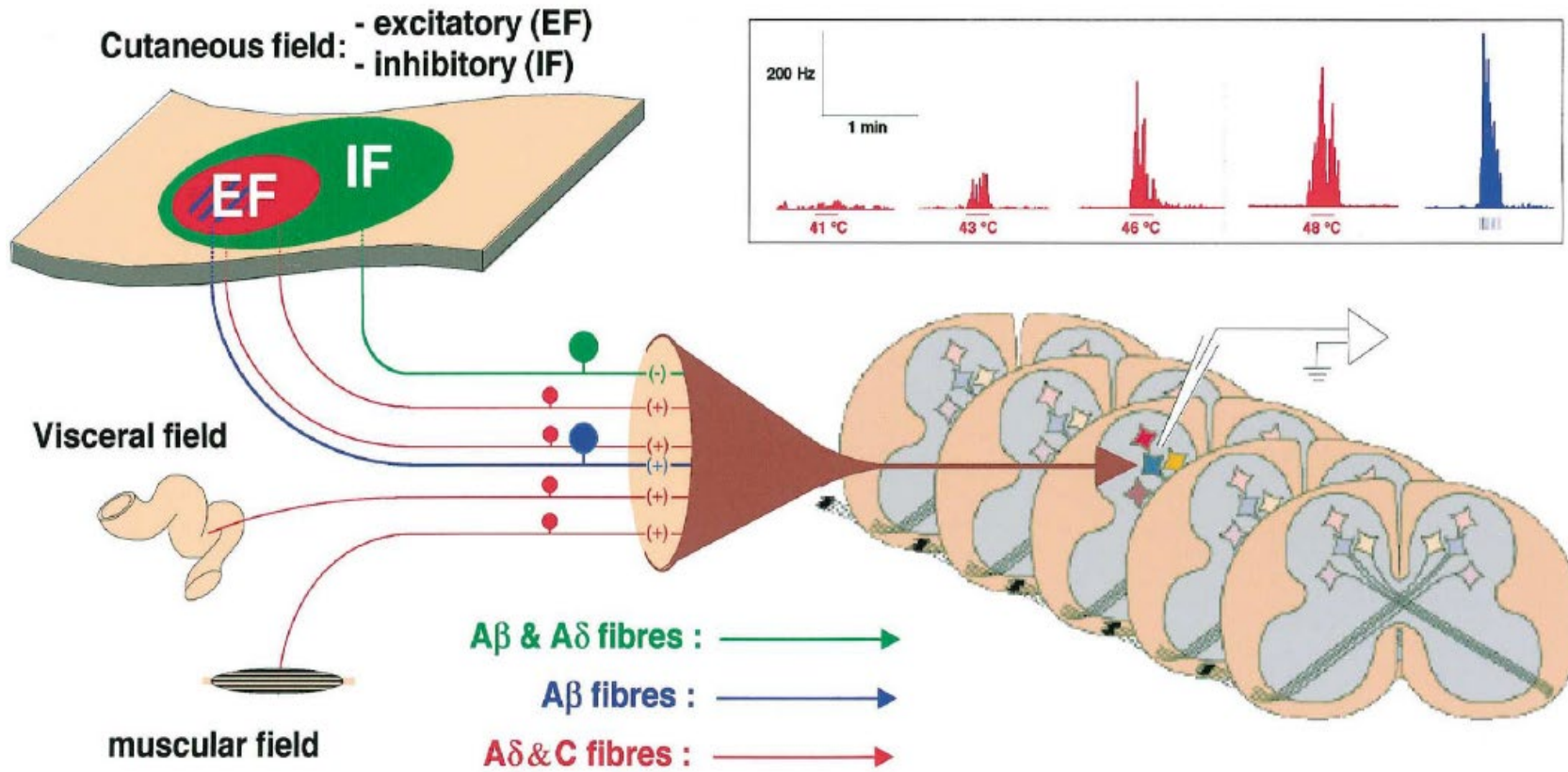
Complexity of the descending neuromodulatory system



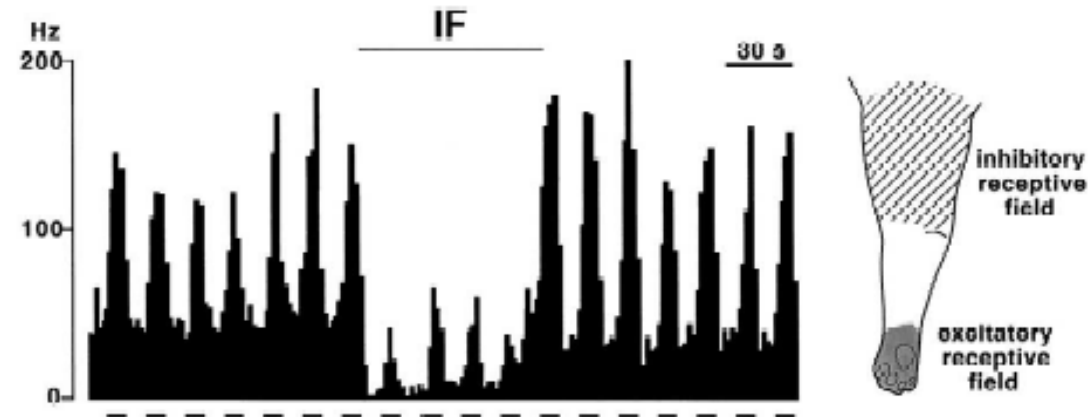
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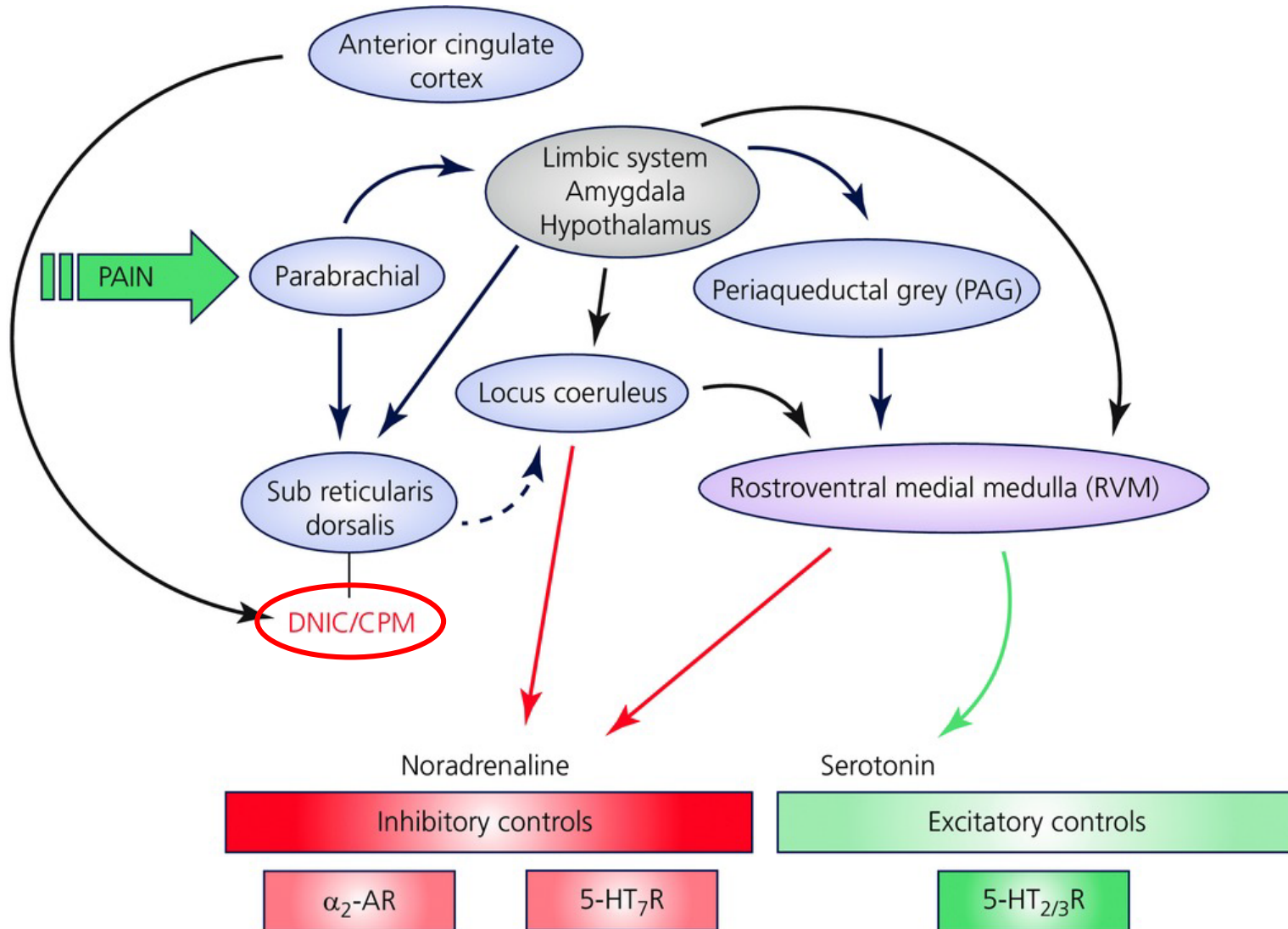
Multireceptive neurons in the Ncl. dorsalis reticularis



► Multireceptive neurons in the spinal dorsal horn receive input from low- and high-threshold afferents of the skin, the deep tissues and the viscera. Their cutaneous receptive fields have a inhibitory periphery. They are inhibited by noxious stimuli of many different regions of the body. The dorsal reticular nucleus in the caudal medulla is involved in the “diffuse noxious inhibitory controls” (DNIC) activated by Aδ- and C-fibers.

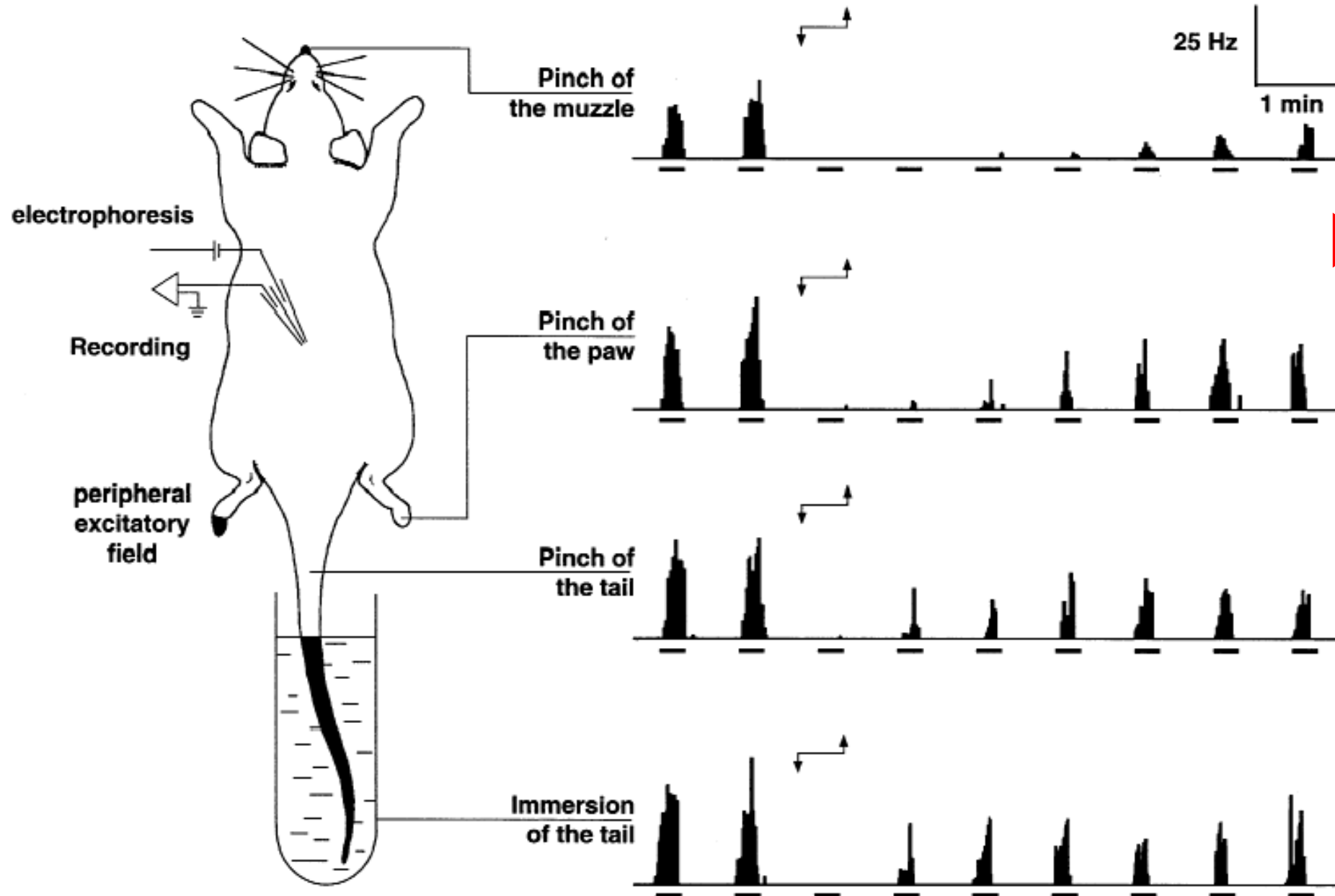


Descending pain modulatory system



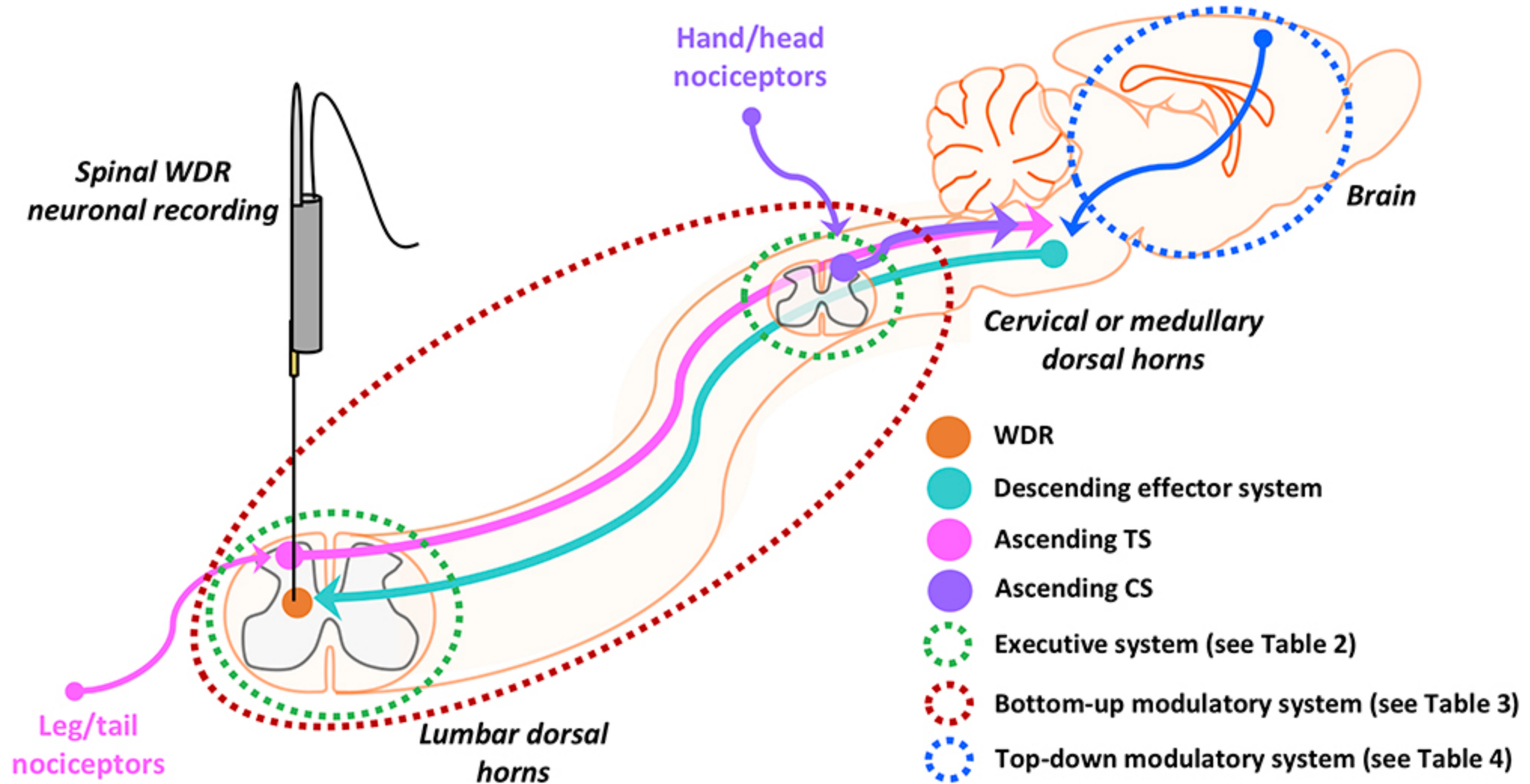
The descending pain modulating systems operate with **inhibitory** (α_2 adenergetic receptors) and **excitatory** mechanisms (5-HT₂ receptors) in the RVM. Both can have antinociceptive effects. The subreticular nucleus is involved in the **Diffuse Noxious Inhibitory Controls (DNIC)** phenomenon clinically visible as **Conditioned Pain Modulation (CPM)**.

Multireceptive neurons in the Ncl. dorsalis reticularis respond to DNIC

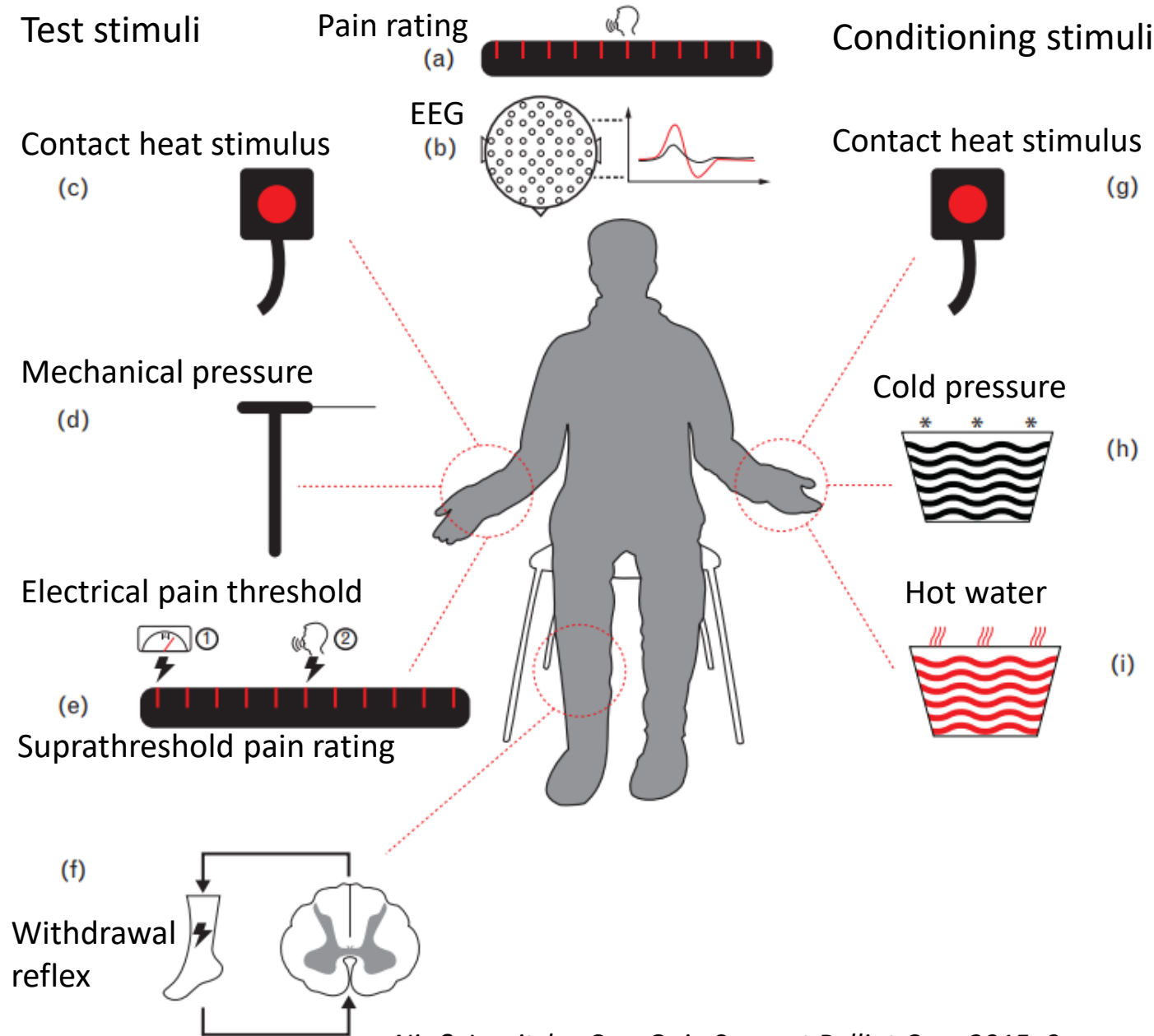


▶ Multireceptive neurons in the dorsal horn are silenced by short noxious stimulation of multiple areas (A δ - and C-fibers input) for *many minutes*. The dorsal reticular nucleus in the caudal medulla is involved in the “diffuse noxious inhibitory controls” (DNIC).

Hypothetic system of diffuse noxious inhibitory controls (DNIC)

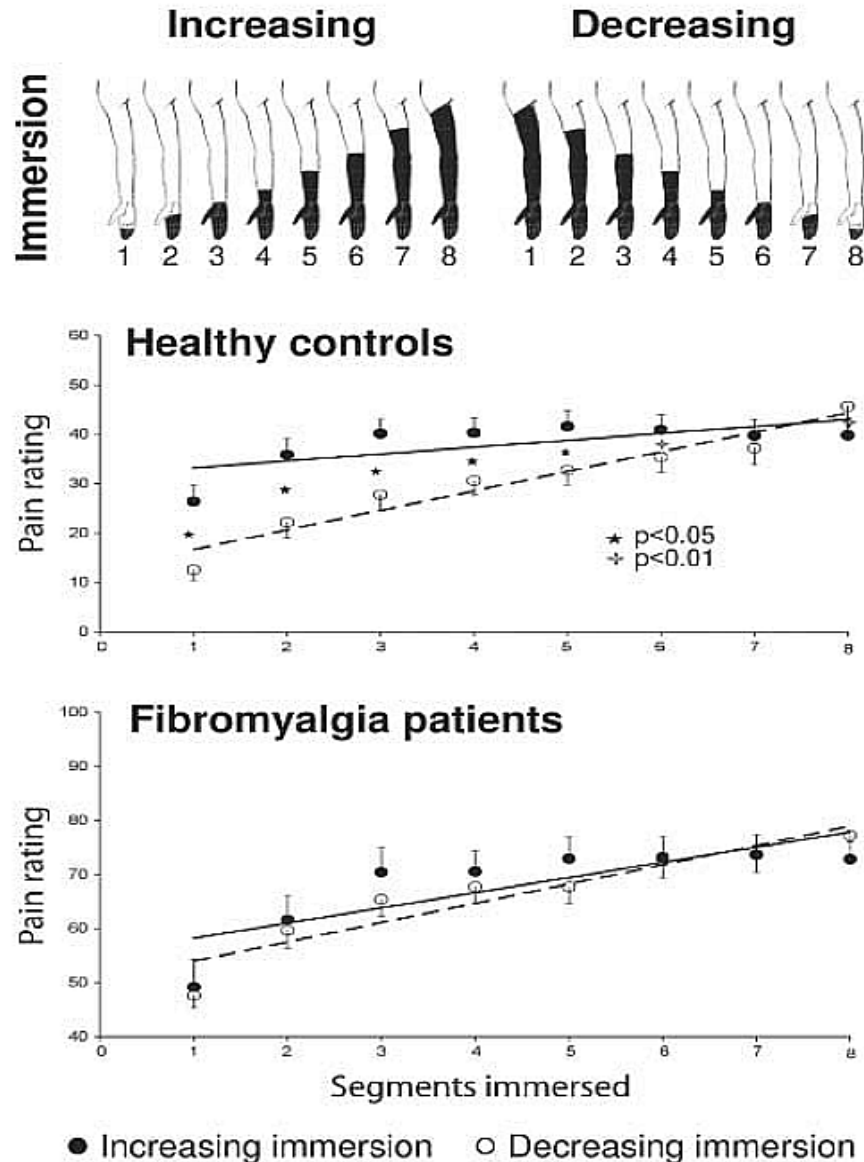


Conditioned pain modulation testing DNIC in humans



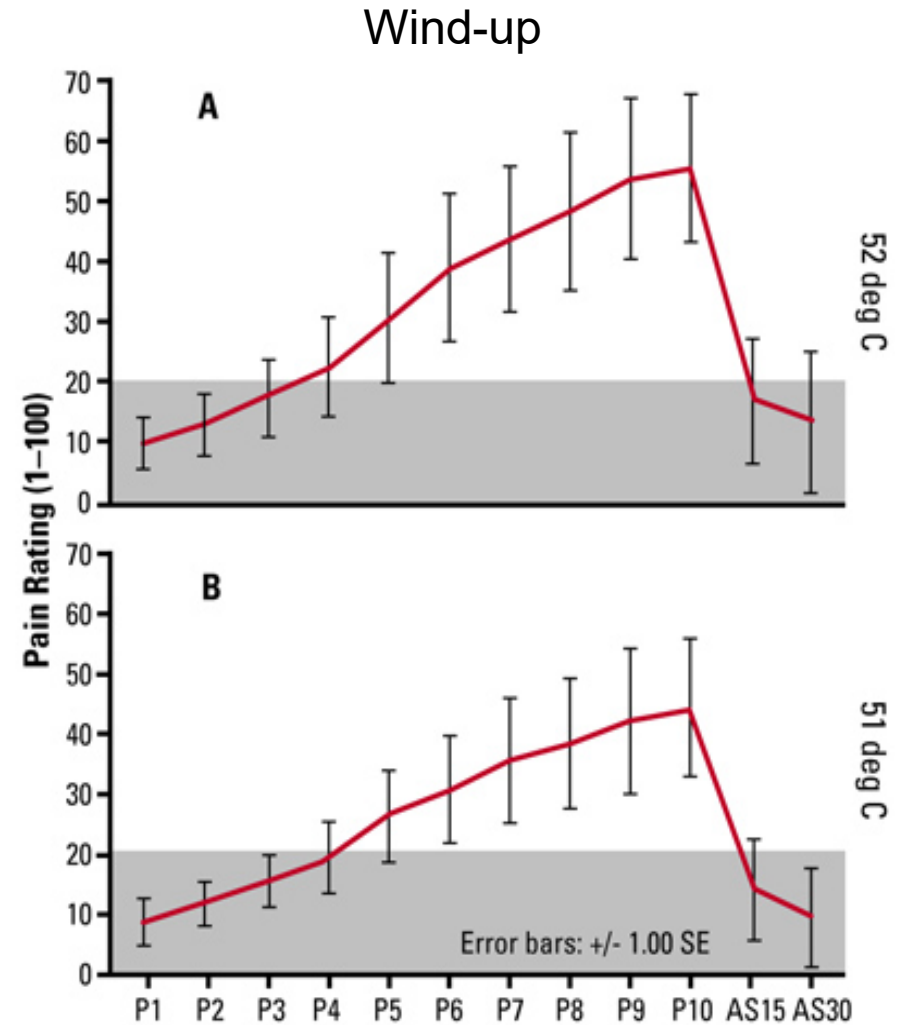
► Conditioned pain modulation tests are appropriate for testing the effect of DNIC in humans. Thereby, pain rating or objective parameters of pain (EEG) to noxious thermal, mechanical or electrical stimuli are assessed before and after a *homo- or heterotopic* conditioning stimulus. Decreased responses to test stimuli indicate DNIC.

Pathological DNIC and wind-up phenomenon of pain ratings in fibromyalgia patients

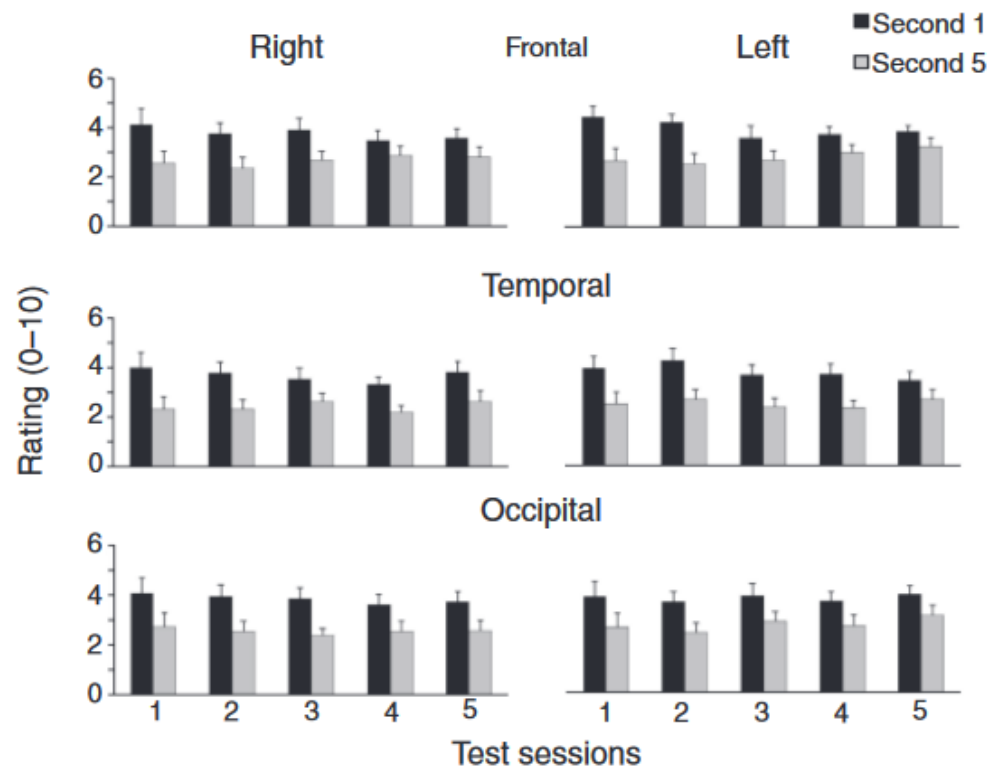
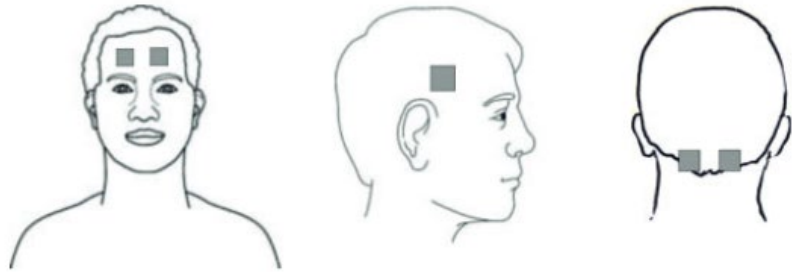


▶ Noxious stimulation (heat) of an increasing followed by a decreasing area is usually followed by less pain rating (habituation). This habituation is lacking in patients with fibromyalgia.

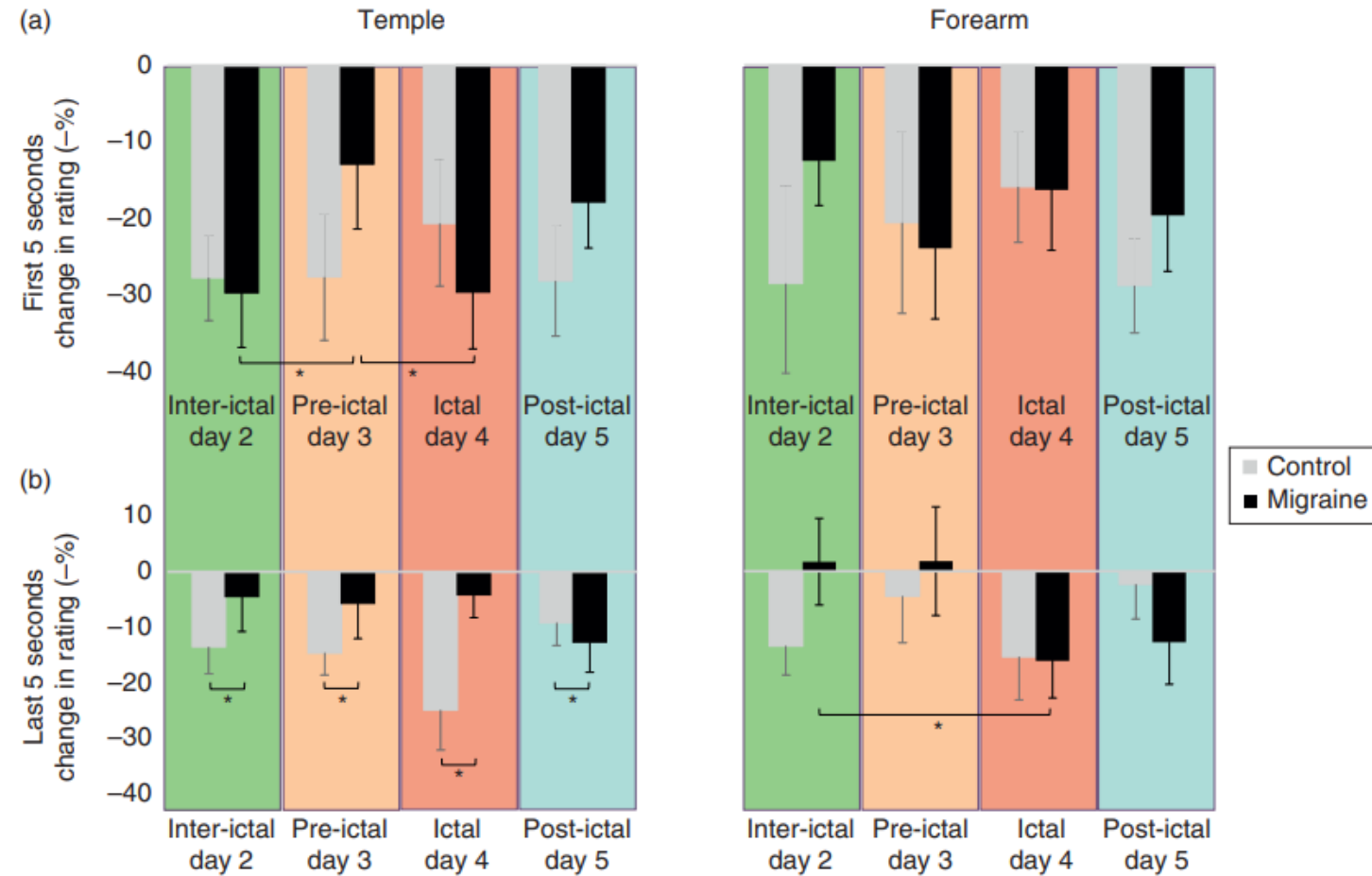
▶ Repetitive noxious stimulation (heat) at same intensity causes an increase in pain rating („wind-up“ phenomenon) in fibromyalgia patients.



DNIC may underlie habituation to painful stimuli – habituation deficit in migraine



▶ Periodic painful electrical stimulation is experienced as less painful after stimulation of some seconds (habituation). This phenomenon is disturbed in migraine patients (habituation deficit), particularly in the ictal phase.

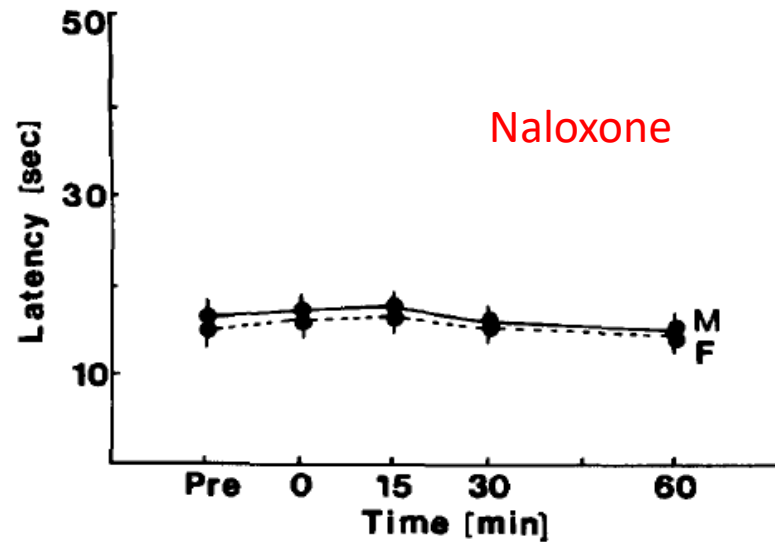
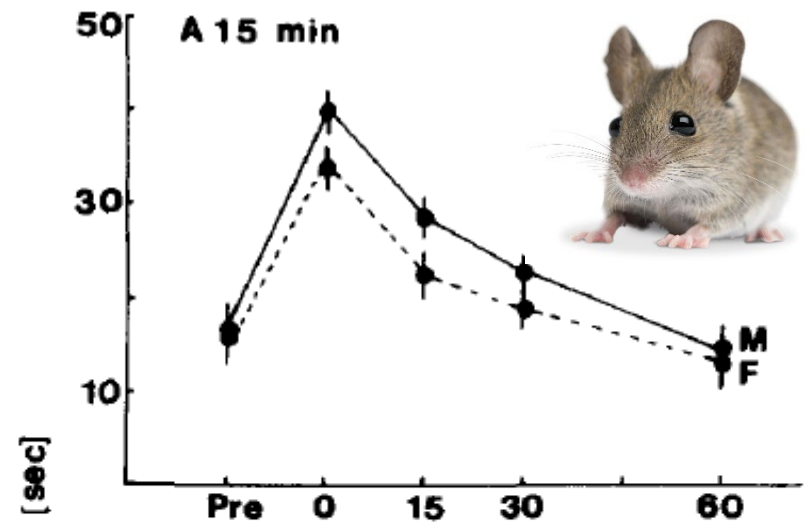


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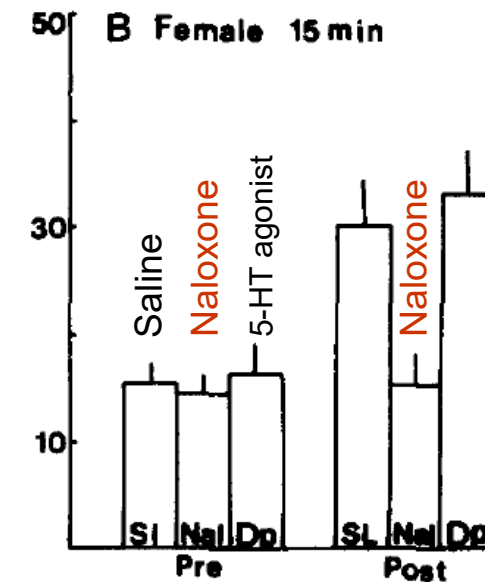
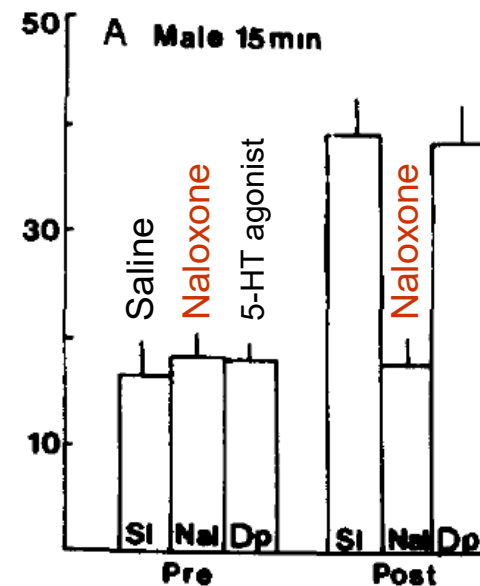
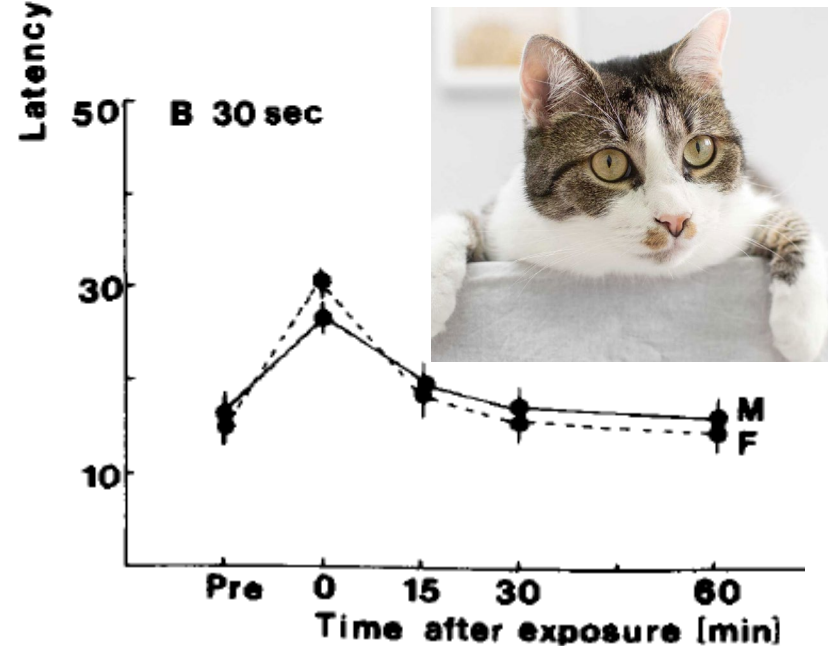
- **Retrospection:** Basic organization of ascending and descending pain modulating pathways
- **Recollection:** Molecular mechanisms of nociceptive transmission and central sensitization
- **Controlling spinal nociception:** Interneurons and molecular antinociceptive mechanisms
- **Descending pathways:** On and Off cells in the rostral ventromedial medulla
- **Supraspinal antinociceptive mechanisms:** Diffuse inhibitory control systems (DNIC)
- **Central pain inhibition:** Stress- and exercise-induced hypoalgesia and acupuncture

Predator-induced opioid-dependent analgesia in mice

Latency to heat (50°C) for licking behavior

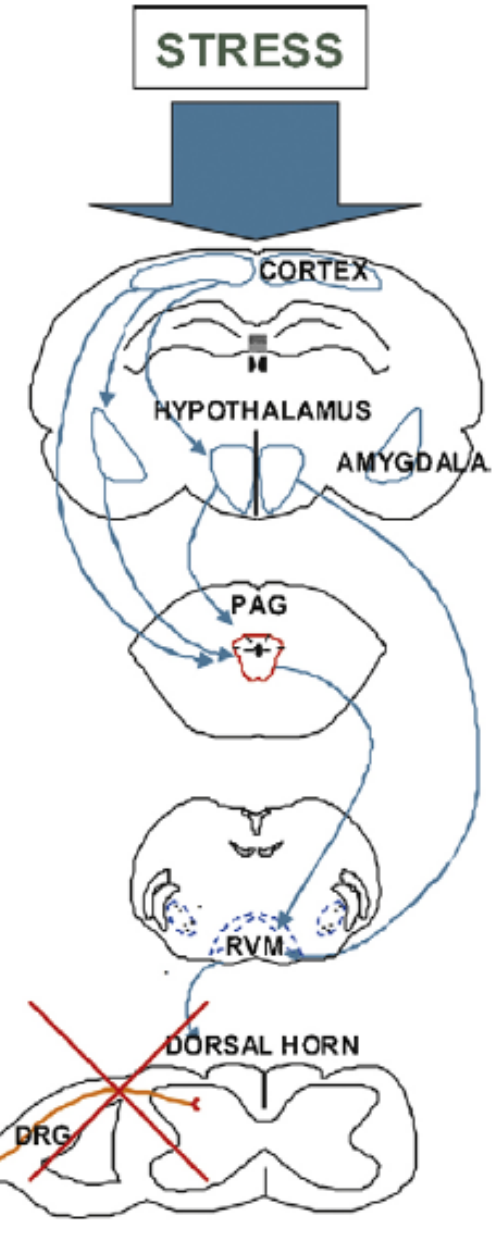
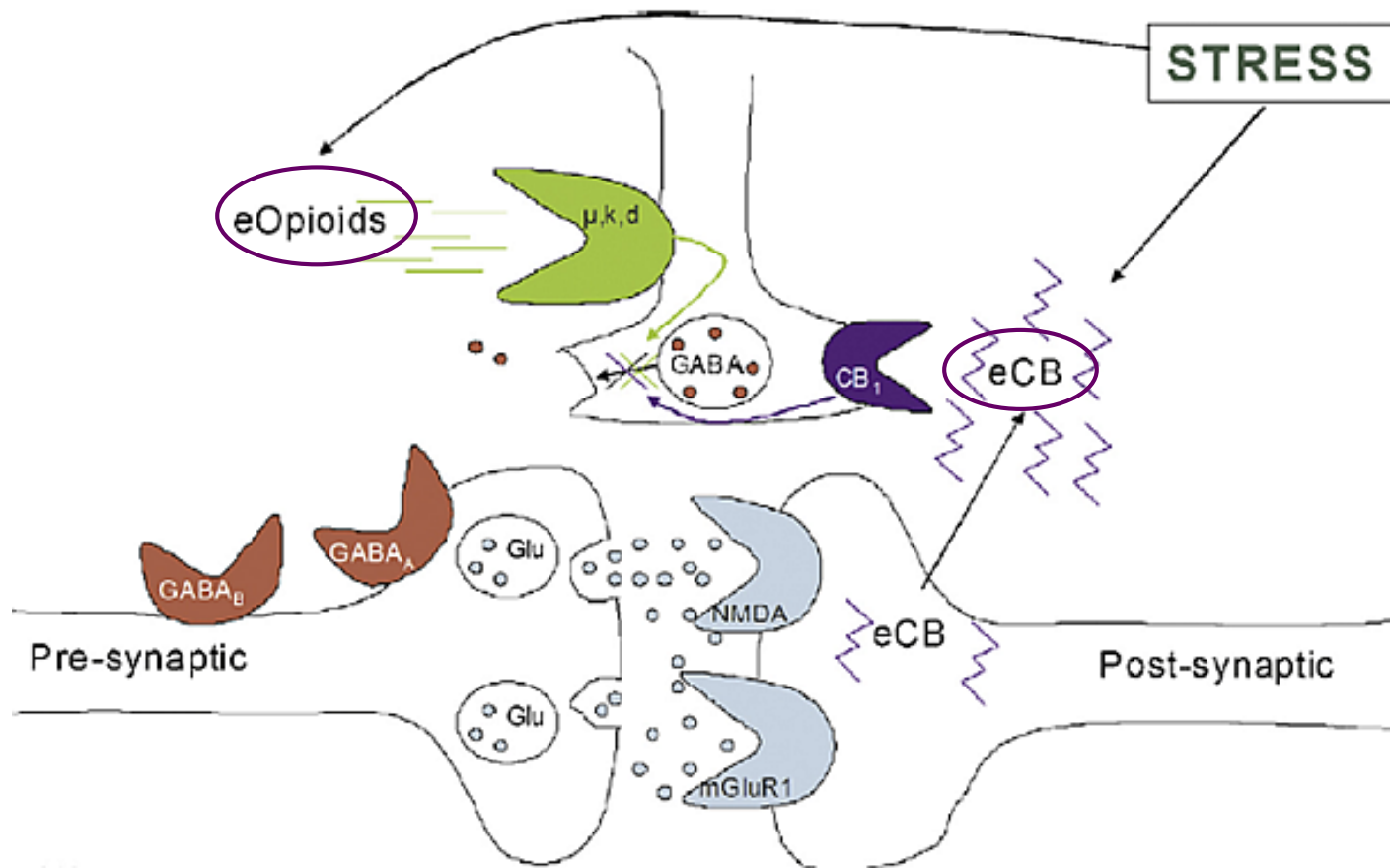


▶ Seeing a predator decreases licking behavior as a response to heat stimuli in mice. This analgesic effect is abolished by the opioid antagonist naloxone (opoid antagonist) showing the involvement of opioidergic mechanisms.



Stress-induced analgesia can be explained by central opioid mechanisms.

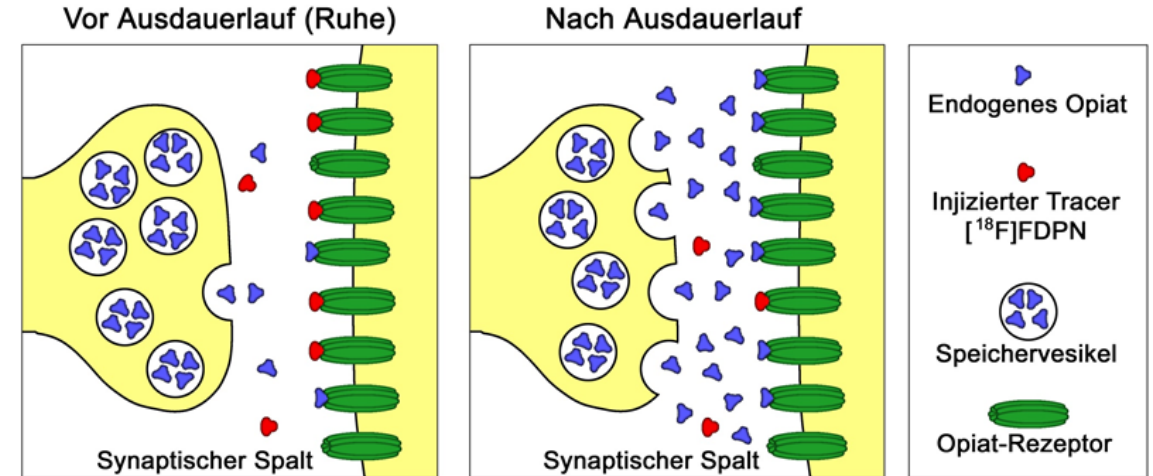
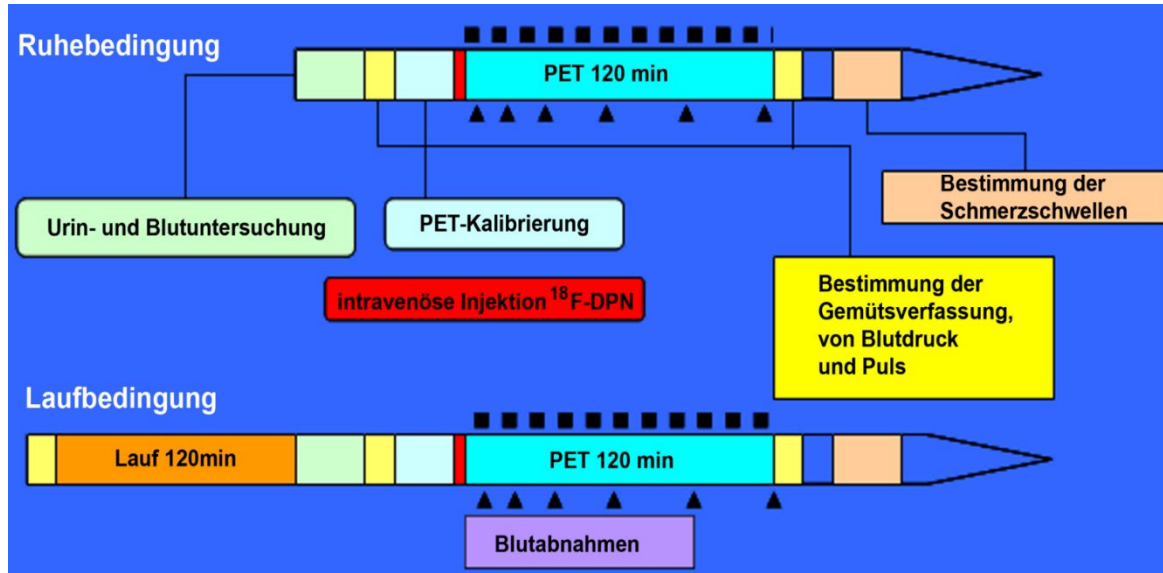
Proposed presynaptic mechanism of stress-induced analgesia



- Acute stress may cause secretion of endogenous opioids and endocannabinoids causing spinal analgesia.

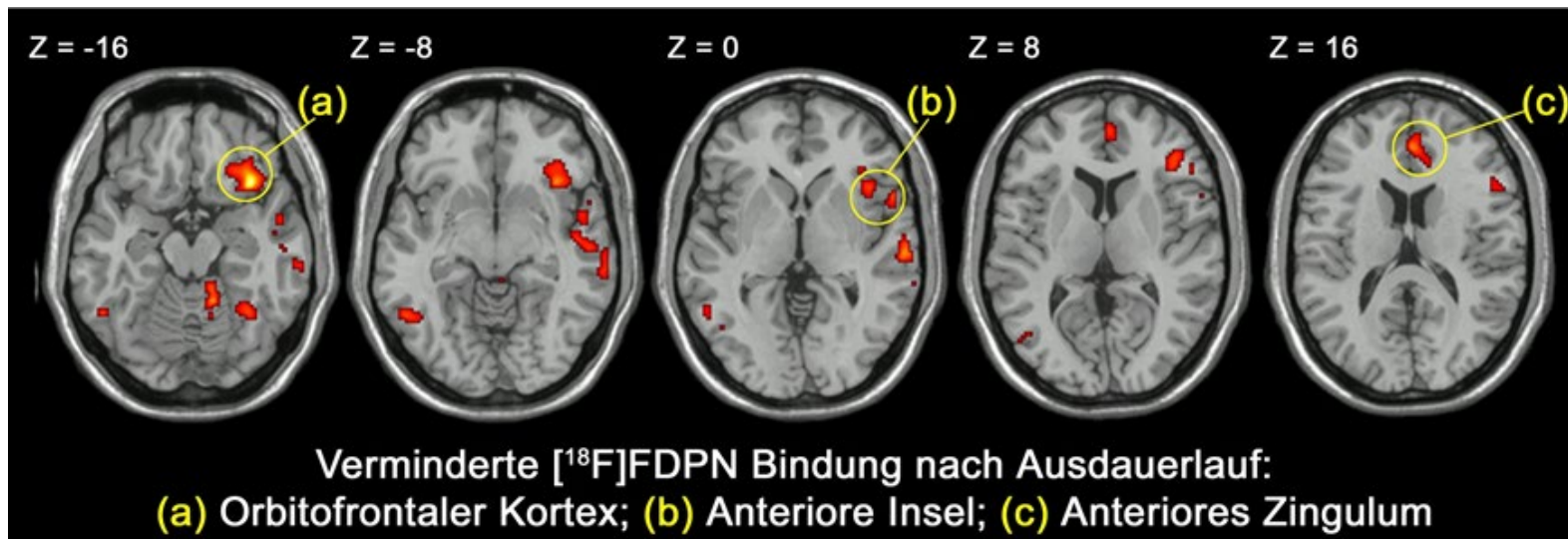
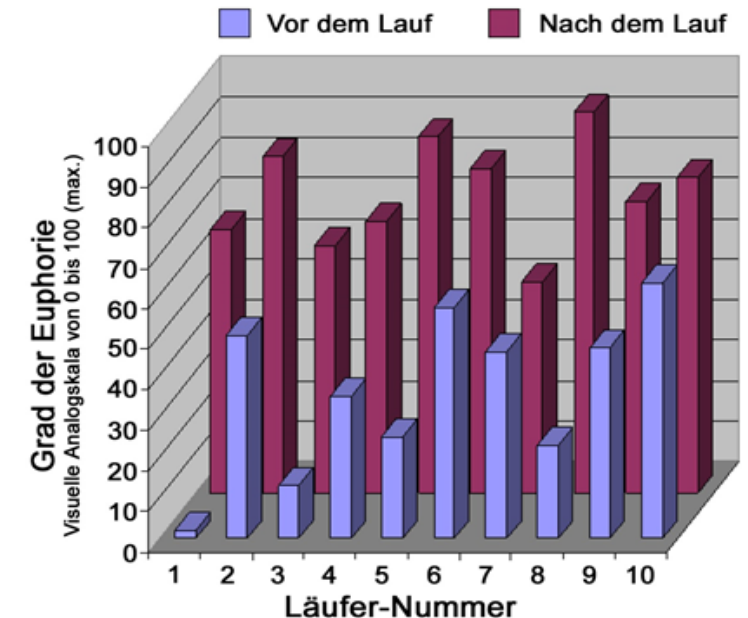
Opioid release in long-distant runners („runners high“)

Versuchsablauf



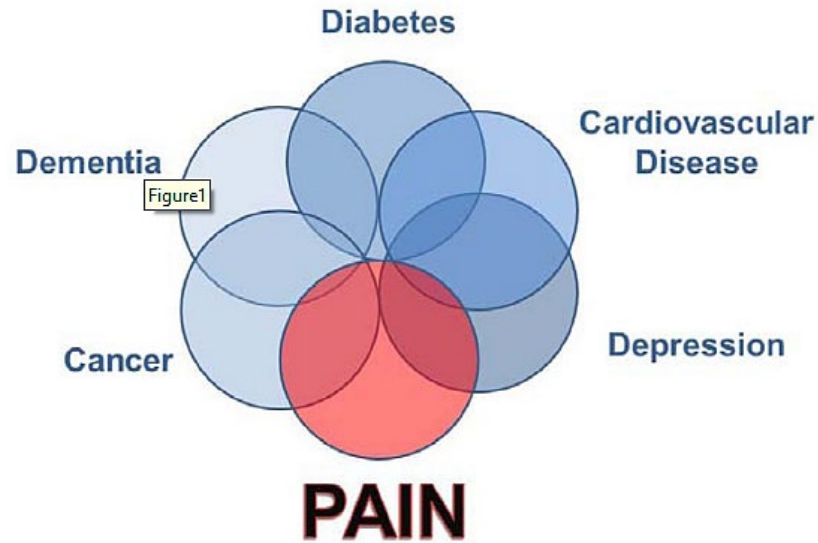
^{18}F -DPN = Fluor-markiertes Diprenorphin

Ausmaß der Euphorie bei 10 Läufern vor und nach dem Ausdauerlauf



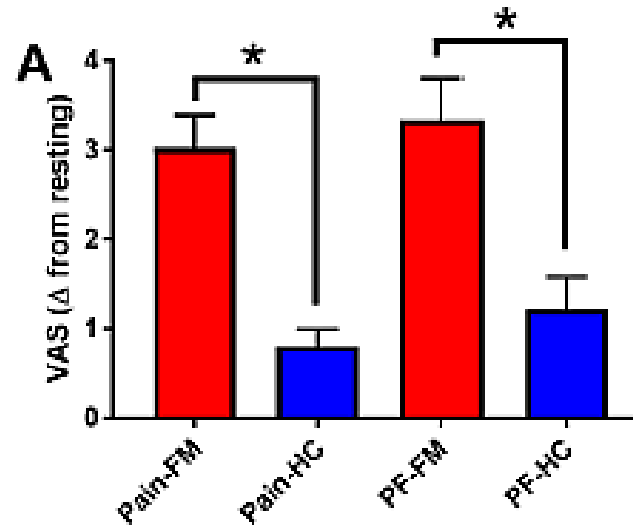
Pain, fatigue and exercise in fibromyalgia

Diseasome of Physical Inactivity

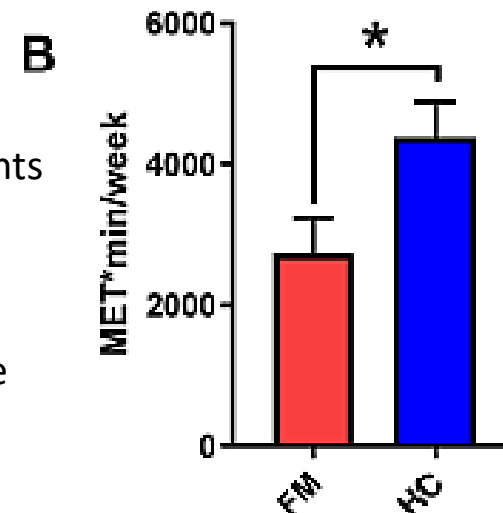


FM = fibromyalgia patients
HC = healthy controls
MET = activity measure
PF = physical fatigue
VAS = visual analog scale

Pain and fatigue



Physical activity



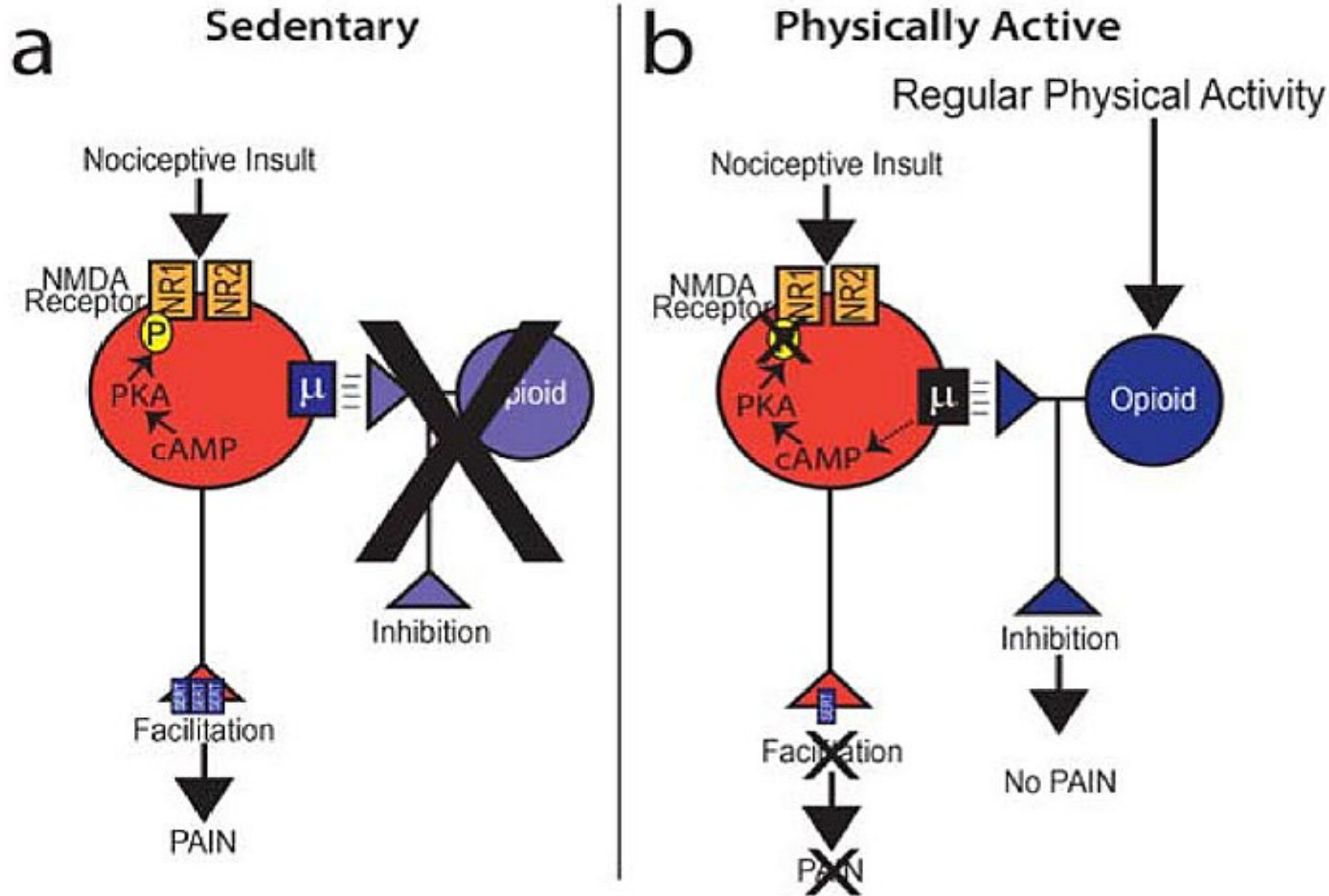
► **A.** In fibromyalgia patients both pain (measured on a visual analog scale) and physical fatigue are significantly increased.

B. This is the main reason why fibromyalgia patients are significantly less active than healthy controls.

Chronic painful diseases like fibromyalgia are frequently accompanied by decreased activity, which finally increases the pain.

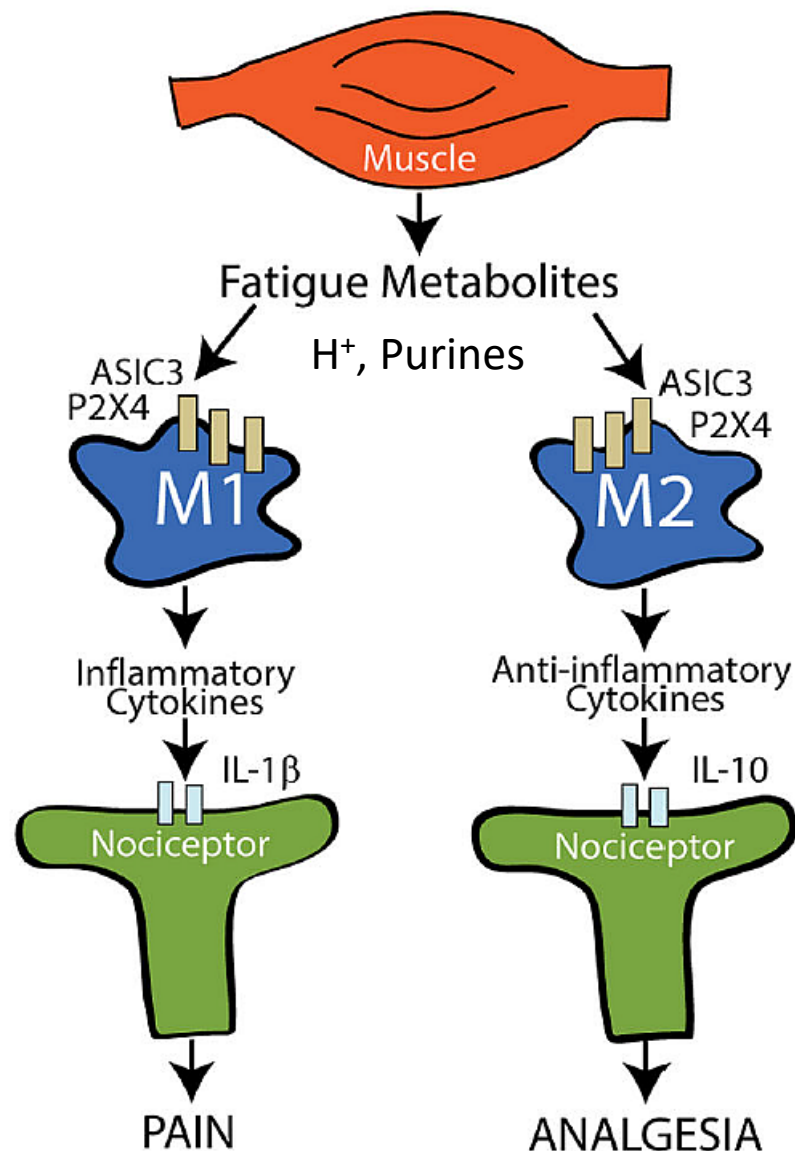
Therefore, light exercise is essential for these pain patients.

Changes in spinal neurotransmission and activity of macrophages



► Sedentary behavior facilitates nociceptive transmission in the dorsal horn at NMDA receptors and suppresses the antinociceptive activity of opioidergic interneurons, while physical activity increases opioidergic effects.

Changes in spinal neurotransmission and activity of macrophages

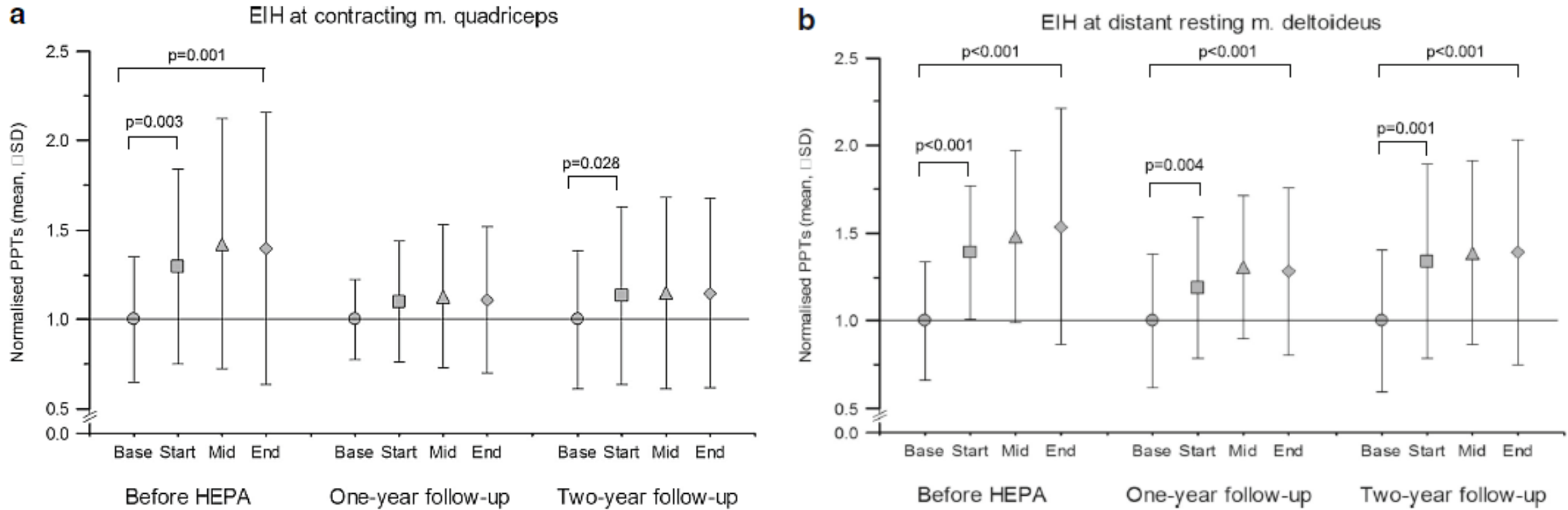


► Fatigue muscles release metabolites like protons and purines, which induce the formation of type-1-macrophages that secrete inflammatory and noxious cytokines like IL-1b.

Light exercise causes formation of type-2-macrophages with anti-inflammatory and anti-nociceptive cytokines like IL-10.

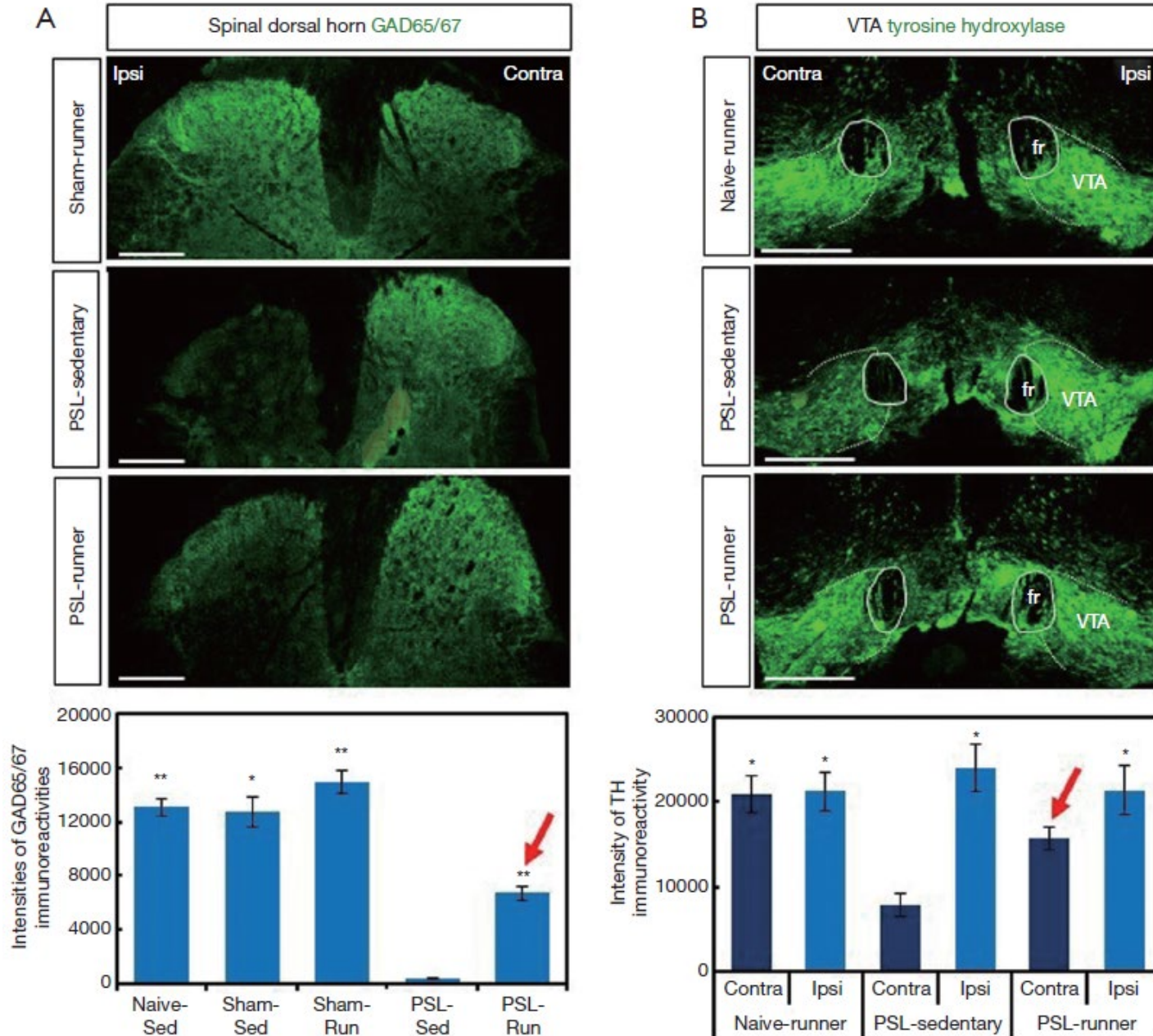
Muscle activity can significantly contribute to pain therapy due to its anti-inflammatory effect.

Impact of physical activity on pain sensation in patients with rheumatoid arthritis



► In patients with rheumatoid arthritis after physical training and moderate aerobic (HEPA) the pain pressure thresholds (PPTs) increase (i.e., the pain decreases). This is seen both at the trained muscle (M. quadriceps, **a**) as well as at not trained muscles (M. deltoideus, **b**). This indicates multisegmental improvement of the pain symptoms.

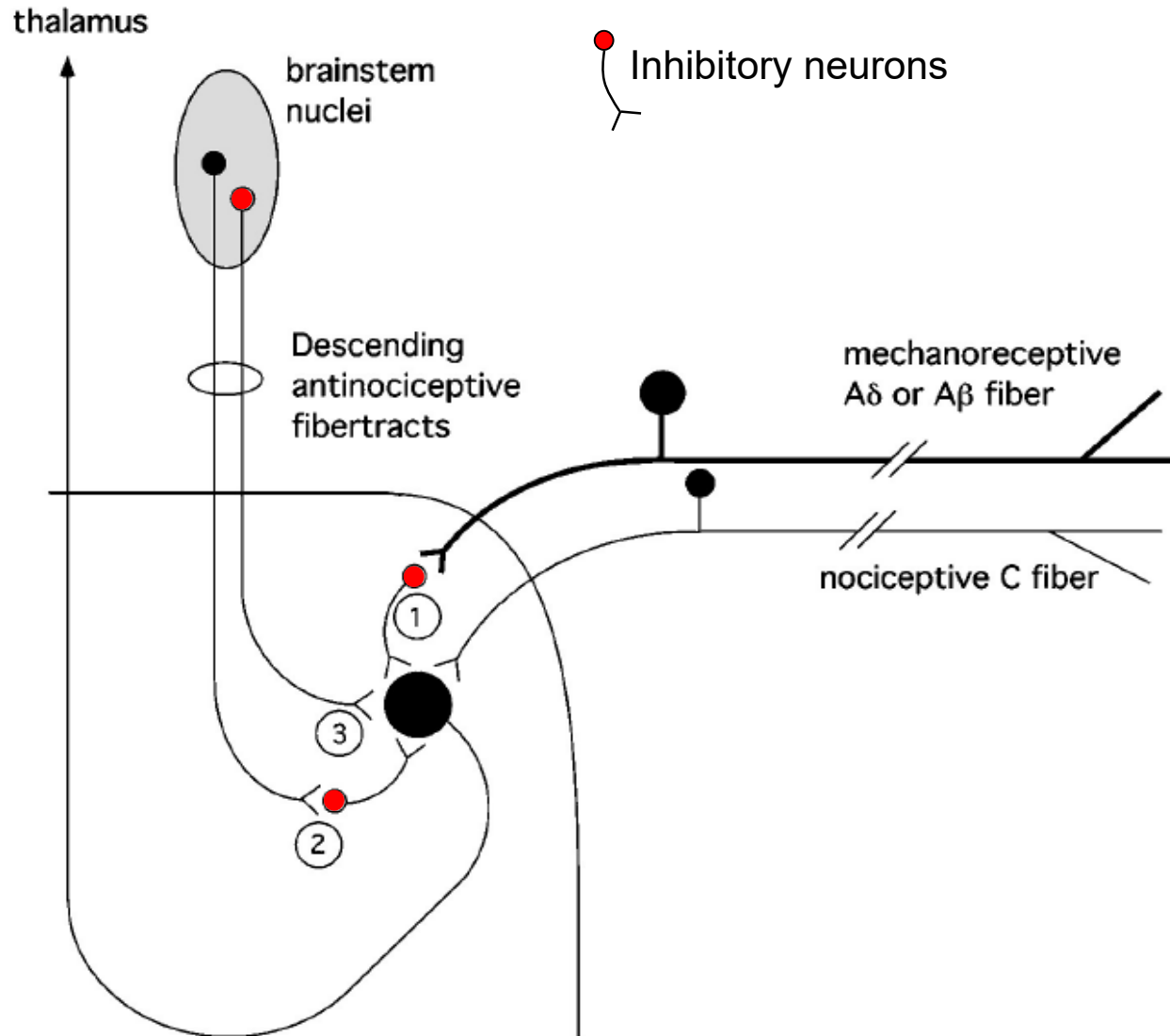
Impact of physical activity on the production of GABA and dopamine



After partial ligature of the sciatic nerve (N. ischiadicus) as a model of neuropathic pain, GAD65/67 (glutamate decarboxylase, a marker for GABA production) in the ipsilateral dorsal horn and tyrosine hydroxylase (a marker for dopamine synthesis) in the contralateral ventral tegmentum are decreased. Following physical activity (running wheel) this loss of GABA and dopamine is partly compensated (**red arrows**) and the pain thresholds are normalized.

Neuropathic pain, which is partly due to the loss of GABAergic inhibition in the dorsal horn and the anti-depressive dopamine production in the tegmental area, can be alleviated through physical activity, which partly compensates this loss.

Principles of descending and segmental inhibitory systems in the dorsal horn

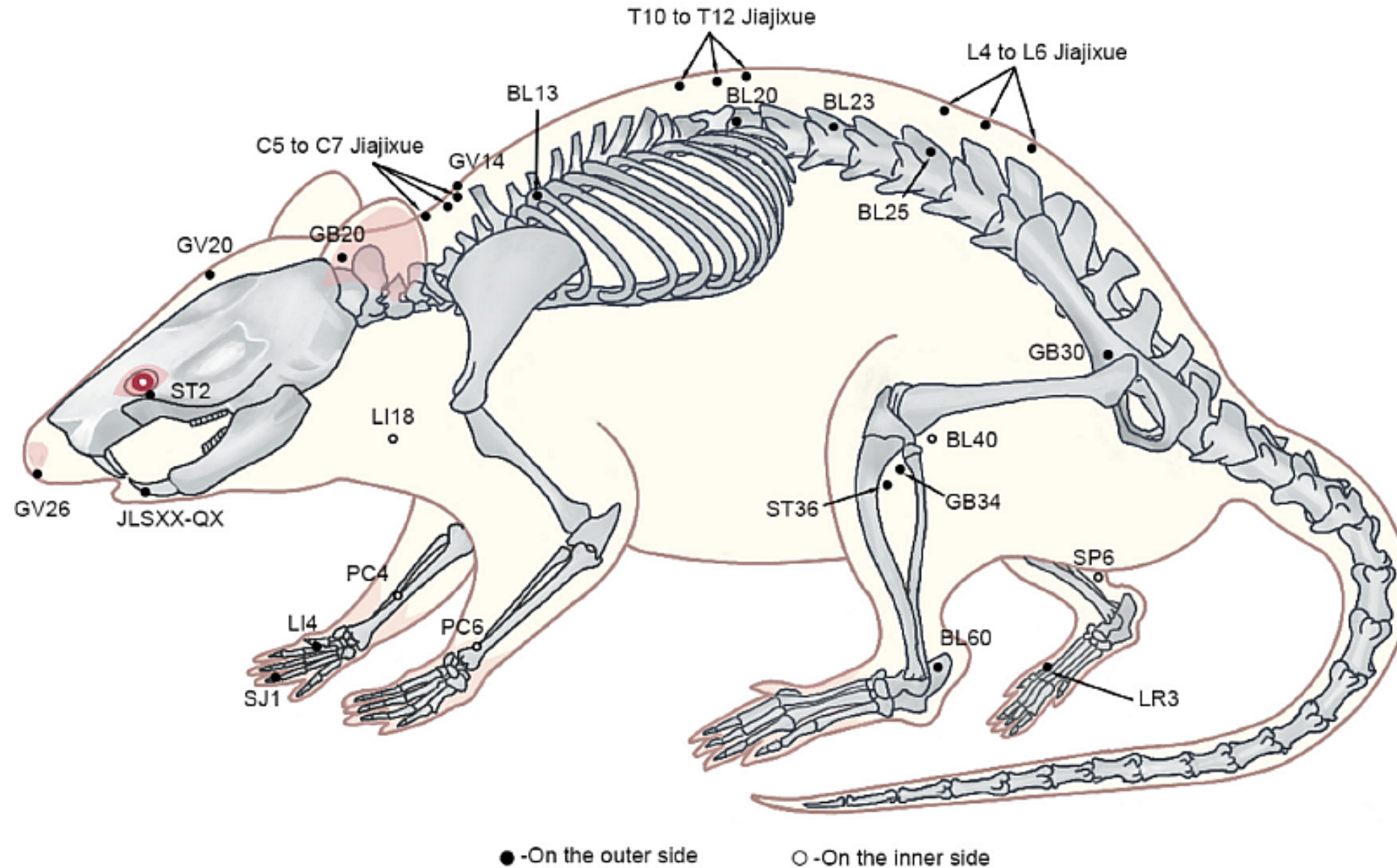


Zeilhofer, Cell. Mol. Life Sci. 2005

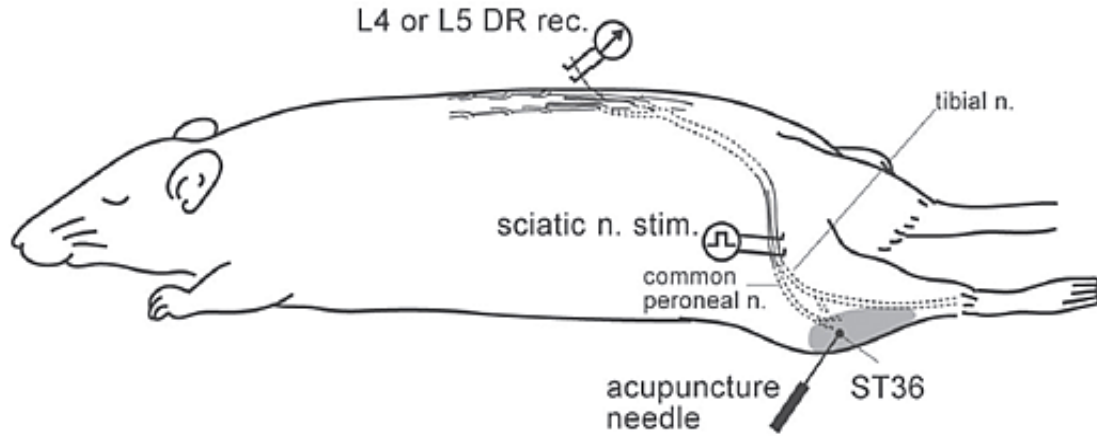


- ① Inhibitory interneuron, activated by mechanoreceptive afferent pathway
- ② Inhibitory interneuron, activated by descending efferent pathway
- ③ Directly inhibitory, descending pathway

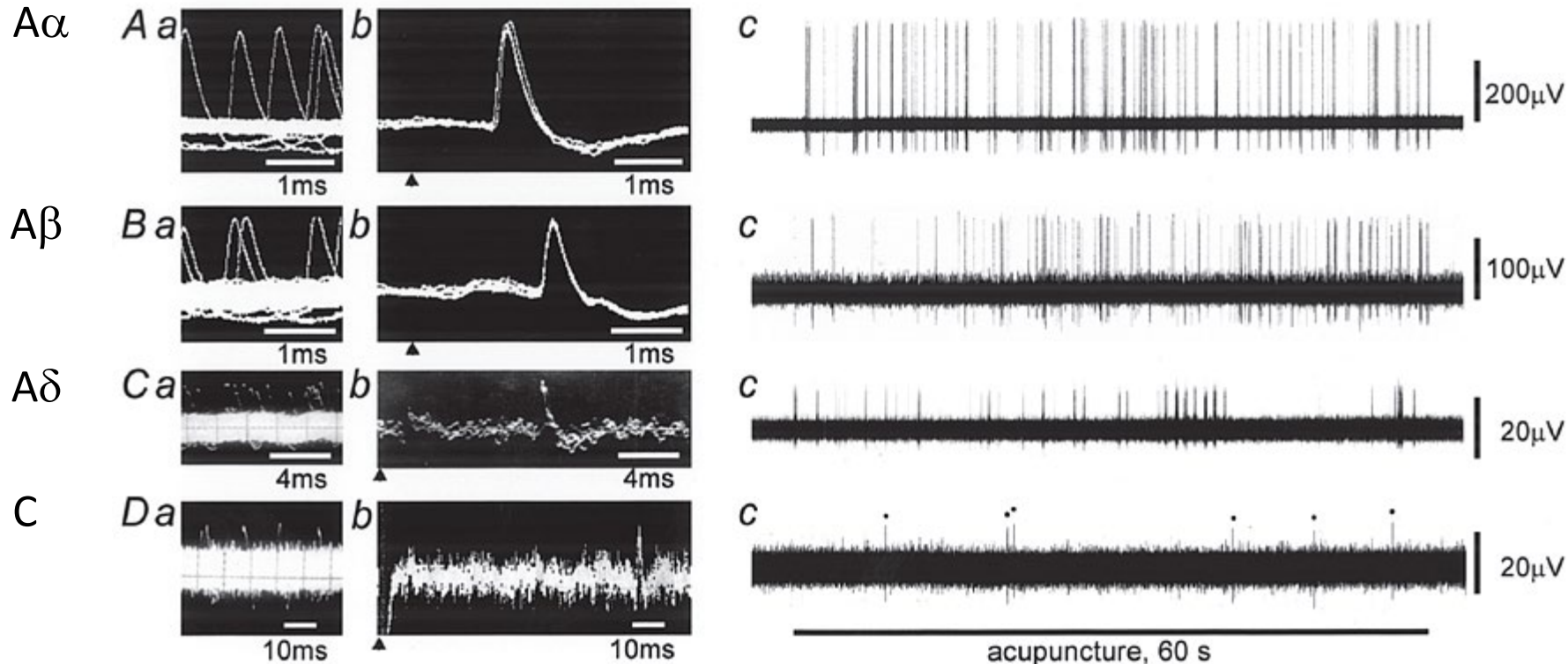
Acupuncture points in the rat



Activation of afferent fibers of all groups through acupuncture



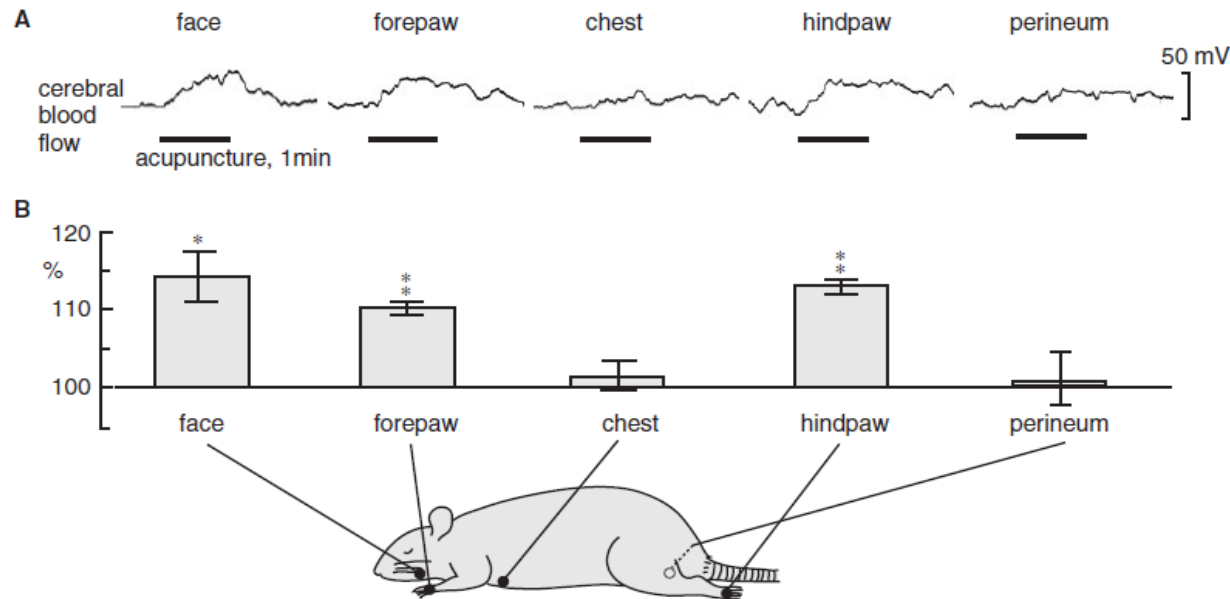
▶ Recordings from single afferent fibers in the lumbar dorsal roots show activation of all fibers during acupuncture.



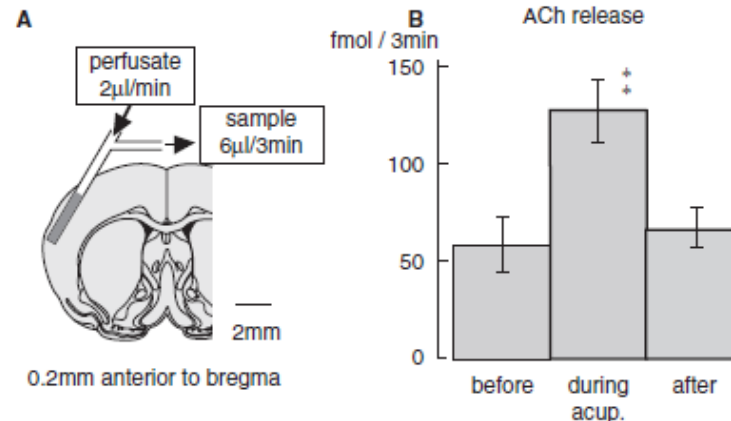
Acupuncture activates both low- and high-threshold (nociceptive) afferents. The pain inhibitory effect of acupuncture may (at least partly) depend on counter-irritation.

Effects of local „acupuncture“ and „acupressure“ in animals

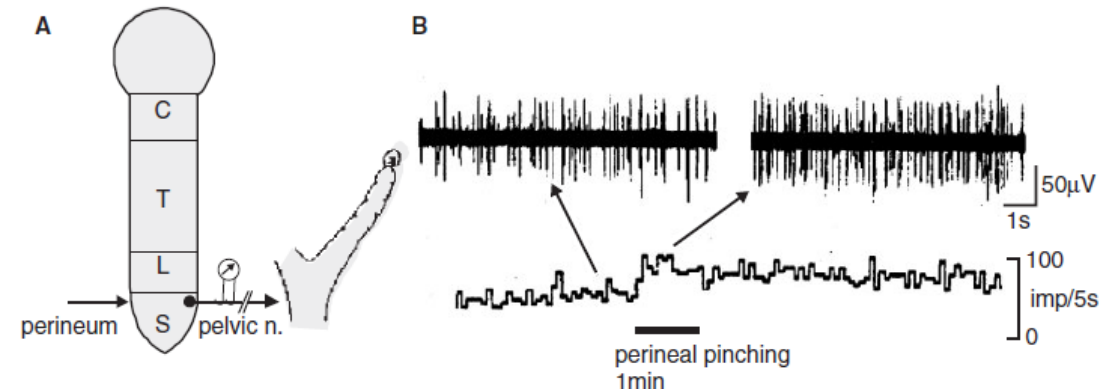
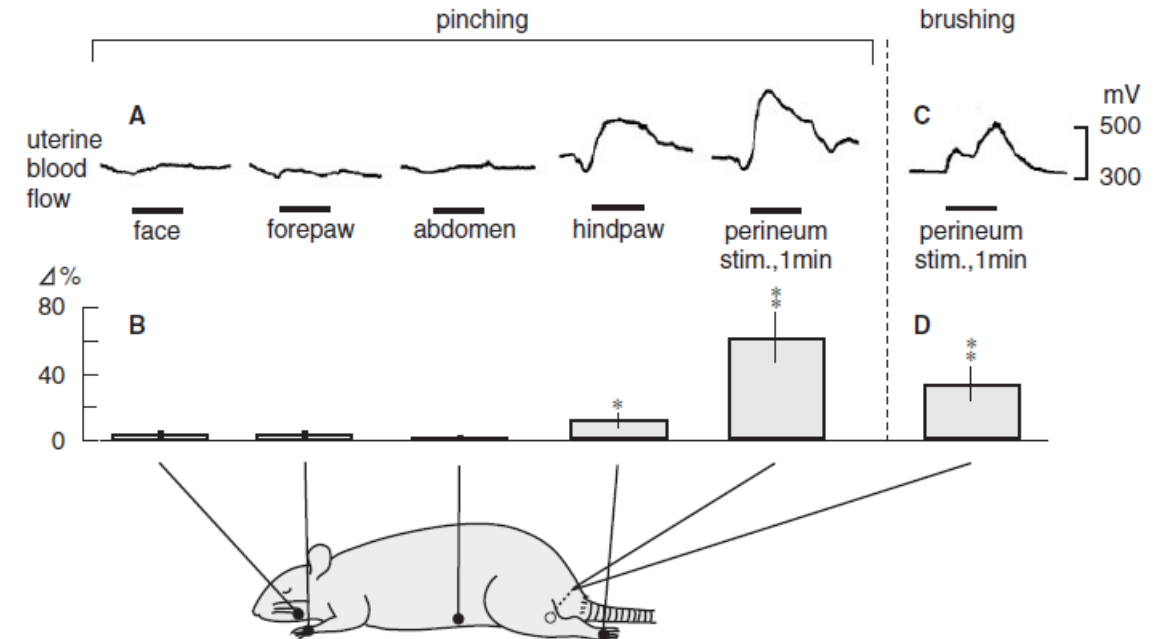
- Central effect: Experimental acupuncture in the face as well for- and hindlegs increases **cerebral blood flow** and the release of acetyl choline in the parietal cortex.



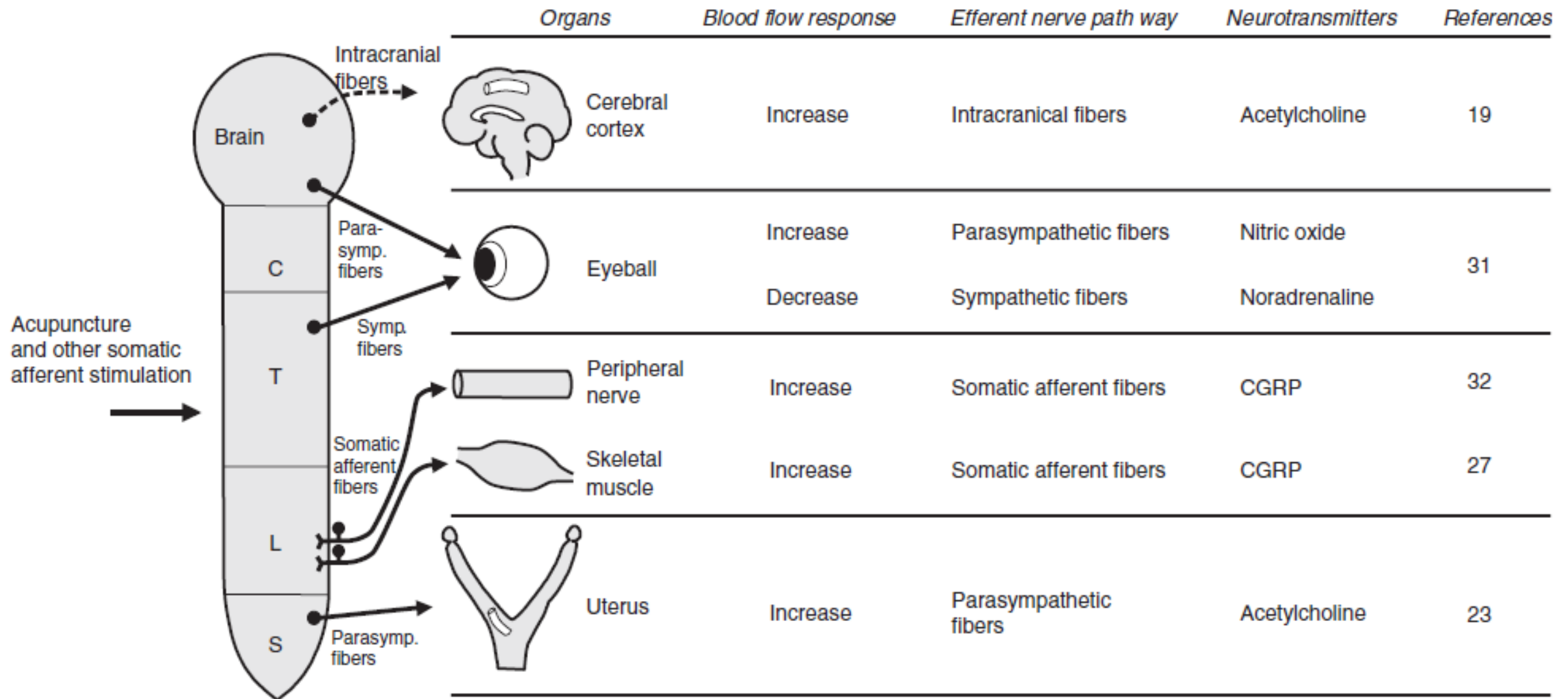
„Acupuncture“ and „acupressure“ have central and peripheral autonomic effects.



- Segmental effect: noxious and non-noxious stimulation of hind foot and tail increases the **uterine blood flow** and the neuronal activity of the pelvic nerve.



Effects of local „acupuncture“ and „acupressure“ in animals



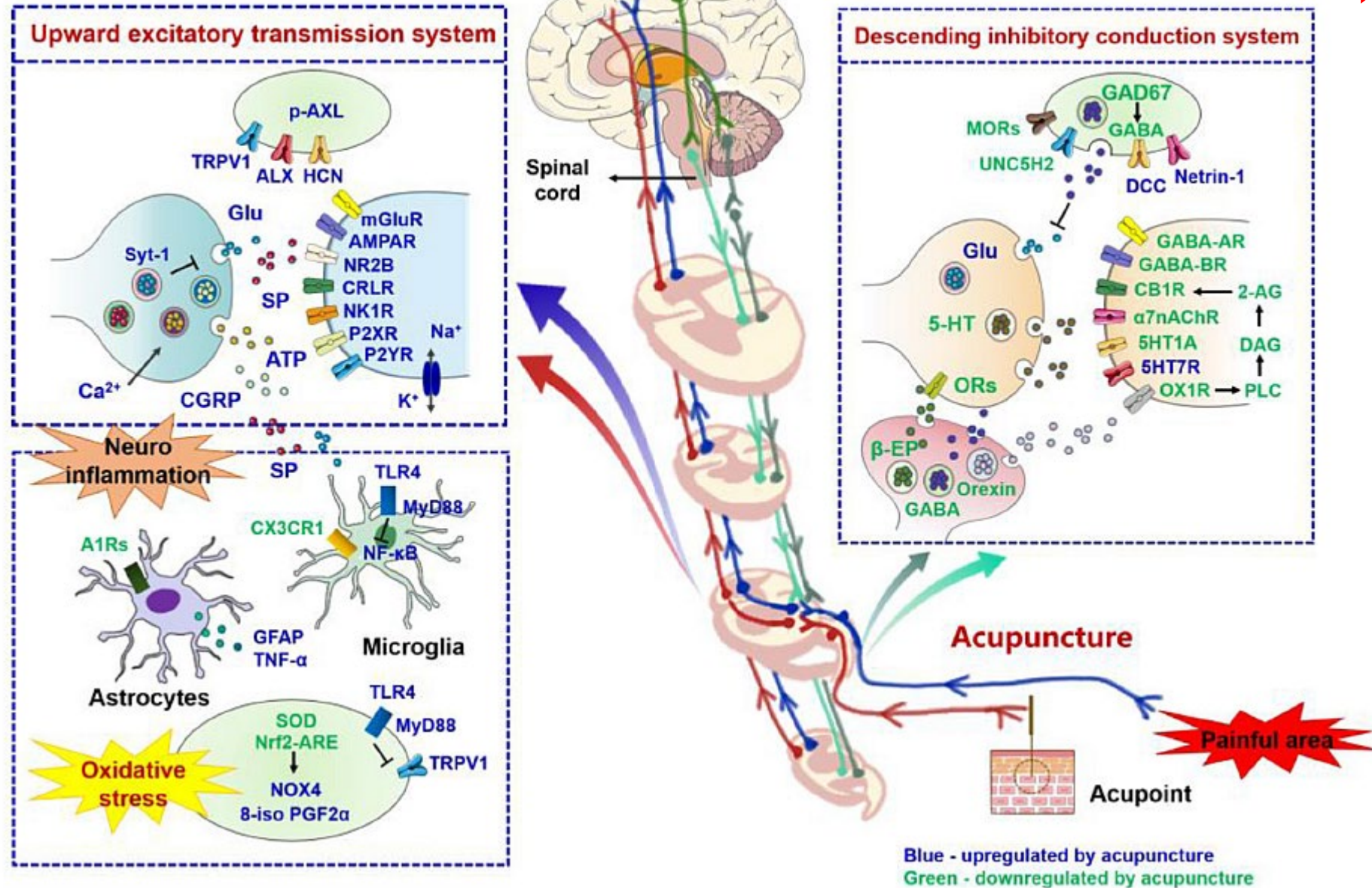
Effects of acupuncture on nociceptive functions during acupuncture of rodents

References	Pain model	Species	Intervention methods	Acupoints	Acupuncture parameter	Pain-related behavior	Test site	Biochemical measurements
Wang et al. (31)	SNI	Rat	EA	ST36/SP6	2 Hz, 0.5–1.5–2 mA, 30 min	PWL	Lateral hypothalamus	c-Fos-Positive Orexin Neurons↑
Zhu et al. (32)	SNI	Mouse	EA	ST36/SP6	2 Hz, 0.1 mA, 30 min	PWT	Brains	rACC Glu -viPAG↓
Xia et al. (33)	SNI	Rat	EA	ST36/SP6	2 Hz, 1–2–3 mA, 30 min	PWT	L4–L6 spinal cord	HMGB1↓, TLR4↓, MyD88↓, NF-κB p65↓, CD11b↓
Ali et al. (34)	SNL	Rat	EA	ST36/SP6	2 Hz, 2–3 mA, 20 min	PWT	Spinal microglial	IL-10↓, β-endorphin↑
Wei et al. (35)	SNL	Rat	EA	GB30/GB34	2 Hz, 1–2–3 mA, 30 min	PWT	L5 DRGs	p-AXL↓, AXL↓
Zheng et al. (36)	SNL	Rat	EA	ST36/BL60	2/100 Hz, 1.5 mA, 30 min	MWT and TWL	L4–6 spinal cords	Iba-1↓, BDNF↓, P2X4↓, GABA _A γ2↑
Liang et al. (37)	SNL	Rat	EA	ST36/BL60	2 Hz, 0.5–1.0–1.5 mA, 30 min	PWT and PFD	L4 DRG	P2X3R↓
Wu et al. (38)	SNL	Rat	EA	ST36/BL60	2 Hz, 1.5 mA, 30 min	MWT and TWL	L4–6 DRG	P2X7R↓, p-p38↓, Iba1↓, BDNF↓, IL-1β↓, IL-6↓, TNF-α↓, IL-10↑
Wang et al. (39)	SCI	Rat	EA	PC5/PC6	2 Hz, 2 mA, 20 min	PWT and PWL	L4–6 spinal dorsal horn	p-mTOR↓, p-S6K1↓, p-4E-BP1↓, SP↓, CGRP↓
Ji et al. (40)	SCI	Rat	EA	EX-B 2/BL25/BL40/BL60	1–2–3 mA, 2/100 Hz, 20 min	TWL	L4–L6 spinal cord	COX 2↓
Hou et al. (41)	BPAI	Rat	EA	T10–T12 Jiajixue	2/15 Hz, 30 min	TWL	SC, MC, Cpu, DLT	Metabolic alterations of brain↑
Xu et al. (42)	BPAI	Rat	EA	C5–C7 Jiajixue	2/15 Hz, 1.5 mA, 30 min	MWT and TWL	Spleen and lymph nodes	CD4+ T cells↑, β-endorphin↑, IL-17A↑, p-p65 NF-κB↑
Fei et al. (43)	DNP	Rat	EA	ST36/BL60	1 mA and 2 Hz	PWT and PWL	L4–6 DRGs	P2X3R↓
He et al. (44)	DNP	Rat	EA	ST36/BL60	2/100 Hz, 1–2 mA, 30 min	PWL	L4–L6 DRGs	P2X3 receptors↓, CGRP↓
Tang et al. (45)	DNP	Rat	MA	BL14/BL21/BL24	20 min	MWT and TWL	L4–6 DRG	P2X4↓, OX42↓
							Serum	CXCR3↓, TNF-α↓, IL-1β↓, IL-6↓, GSP↓, lipid metabolisms↓
Zhou et al. (46)	DNP	Rat	EA	ST36/BL60	2 Hz, 1 mA, 15 min	PWT	L4–L6 DRGs	P2X3R↓, p-PKC↓
Li et al. (47)	PHN	Rat	EA	GB30/GB34	2 Hz, 1 mA, 30 min	MWT	L4–L6 DRGs	Netrin-1↓, DCC↓, UNC5H2↑
Gao et al. (48)	Neck-incision pain model	Rat	EA	LI18, LI4-PC6, or ST36-GB34	2/100 Hz, 1 mA, 30 min	PT	C2–C5 dorsal cervicospinal cord	ATP↑, P2X7R↑, fractalkine↓, CX3CR1↓
Qiao et al. (49)	Neck-incision pain model	Rat	EA	LI18, LI4-PC6, or ST36-GB34	2/100 Hz, 1 mA, 30 min	PT	C3–C6 DRG	SP↓, CGRP↓, GAD67↑
Qiao et al. (50)	Neck-incision pain model	Rat	EA	LI18/LI4-PC6/ST36-GB34	2/100 Hz, 1 mA, 30 min	PT	C3–6 DRGs	SP↓, GFAP↓, GABA A α2R↓, GABA B R1↑

▶ The multiple effects comprise inhibitory actions on the transduction (downregulation of TRPV1, HCN), decrease in neuropeptide release (substance P, CGRP) and pro-nociceptive cytokines (TNF-α, IL-1β), inhibition of spinal transmission (NMDA receptors, BDNF) and neuroinflammation (JAK2/STAT3) as well as strengthening of inhibitory transmission (GABA receptors, SOM) and the descending inhibition (β-endorphin, μ-receptors).

Effects of acupuncture on nociceptive functions during acupuncture of rodents

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Thank you for your patience



